

1 **Artisanal fortified beers: brewing, enrichment, HPLC-DAD analysis and preliminary**
2 **screening of antioxidant and enzymatic inhibitory activities**

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26 **ABSTRACT**

27 In this study an **artisanal “Porter style” beer has been enriched with diverse natural bioactive**
28 **substances**, ~~low fermentation enriched beers have been made~~ following *all-grain* brewing method.
29 Common beer generally contains **a poor phenolic content in the class** of phenolic acids which confers
30 a low antioxidant power and nutritional value, also due to the presence of ethanol. However in this
31 work we aimed to enrich beer with different flavonoids and other food supplements like taurine,
32 resveratrol and caffeine, thus enhancing its nutritional value and energizing properties. A series of
33 flavonoid/phenol-enriched artisanal beers have been prepared, then sample of each has been tested *in*
34 *vitro* to evaluate antioxidant activity, chelating power and enzymatic inhibition capacity. Beer
35 samples were also analysed with HPLC-DAD system to determinate flavonoid and phenol contents.
36 Results show increased nutritional values and significant antioxidant properties in comparison with
37 ~~blank artisanal~~ **not-fortified beer as control** and commercial beer, thus paving the way to the potential
38 use of these beverages as new food supplements.

39 **Keywords:** beer, flavonoids, antioxidants, ~~food supplements~~, fermentation, **drink**

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53 1. Introduction

54 Beer is one of the most popular alcoholic drinks consumed in the world and its production is known
55 since ancient times. Beer brewing is typically realised through alcoholic fermentation of barley malt
56 operated by different types of yeasts, with the final adjunction of hops. (Olaniran, Hiralal, Mokoena,
57 & Pillay, 2017) The main ingredient is malt, which derives from some types of grasses, especially by
58 barley. Malt is the result of an enzymatic digestion operated by a series of amylases produced by the
59 cereal in the germination step, during which starch is hydrolysed into small carbohydrates such as
60 disaccharide **maltose**, the trisaccharide **maltotriose**, monosaccharides like glucose, fructose, and
61 sucrose. (Liu & Quek, 2016; Olaniran et al., 2017) These are fermentable sugars representing the
62 nourishment of a great number of yeasts; *Saccharomyces cerevisiae* and *Saccharomyces*
63 *carisbergensis* are the most common yeast species used to beer brewing. (Liu & Quek, 2016; Olaniran
64 et al., 2017). Literature reports many methods for beer brewing and various type of malts, hops and
65 yeasts that affect beer taste, aroma, colour and texture. ~~The final step before fermentation of beer~~
66 ~~production is the adjunction of hops.~~ Hopping step consists on adding dried hops to the wort during
67 boiling step. (Olaniran et al., 2017) The addiction of hops gives the typical bitter taste which is
68 desirable by the main consumers. (Olaniran et al., 2017) Hops contain a complex series of organic
69 compounds *e.g.* hop acids, hop oil and various flavonoids; the hop acid class of isohumulones is
70 particularly relevant for beer bitter taste. (Olaniran et al., 2017) **Natural phenolic antioxidants**
71 ~~Flavonoids~~ like chlorogenic acid, syringic acid, resveratrol and carvacrol are also added during
72 hopping step **by fruits**. (Olaniran et al., 2017) (Liu & Quek, 2016; Olaniran et al., 2017) Moreover,
73 beer contains a series of volatile compounds responsible for taste and aroma. (Olaniran et al., 2017)
74 In addition to ethanol other alcohols are formed during fermentation step, like propanol, hexanol,
75 benzyl alcohol and isoamyl alcohol; different esters and carbonyl compounds like ethyl acetate,
76 isoamyl acetate, benzyl acetate, acetaldehyde and 2,3-pentanedione derive from amino acids and fatty
77 acid metabolism. These molecules have a fundamental role to confer different aromas and tastes,
78 depending on their concentration. (Olaniran et al., 2017) In fact, isoamyl acetate is responsible of a
79 banana taste, while phenyl acetate gives a honey taste; ethyl acetate confers an undesirable “solvent
80 flavour”, high concentration of hexanol is related to a greasy aroma badly affecting beer taste.
81 (Olaniran et al., 2017) Phenolic content is also a key factor in determining beer taste, haze and
82 bitterness. (Piazzon, Forte, & Nardini, 2010; Shopska et al., 2021; Šibalić, Planinić, Jurić, Bucić-
83 Kojić, & Tišma, 2021) Beer has a moderate content of **phenolic compounds** ~~phenols and flavonoids~~
84 normally present in fruits, vegetables and some beverages ~~with very high antioxidant power~~. They
85 also exhibit a series of different biological activities like antiallergic, antiviral and anti-inflammatory
86 activity, but the most important is the capacity to stabilize and inactivate free radical species deriving

87 from pro-oxidative cellular stress. (Anand David, Arulmoli, & Parasuraman, 2016; Bertuzzi et al.,
88 2020; de Gaetano et al., 2016) **Flavonoids are also present in hops, but** a significant fraction of **them**
89 **flavonoids are** **is** not assimilated by human organism, in fact, intestinal villi can't adsorb flavonoid
90 bonded with matrix **such as** cellulose and other insoluble fibres. ~~however acid condition associated~~
91 ~~with high temperature can hydrolyse this bond.~~ (Schulz et al., 2019) **However in** a recent study by
92 Grieco and co-workers, was reported that lactic fermentation with *Lactobacillus rhamnosus* can
93 hydrolyse this bond, thus increasing the phenolic absorbable fraction in date fruit bars. (Maisto et al.,
94 2021) Most common flavonoids are **quercetin, resveratrol, gallic acid, syringic acid, ferulic and**
95 **caffeic acid, but also rutin, iso-quercetin, chlorogenic acid and catechin.** Quercetin is a coumaric
96 compound belonging to the class of tetrahydroxy-flavonols largely contained in fruit, vegetables and
97 seeds. (Anand David et al., 2016) Different studies have highlighted numerous biological activities
98 like anti-inflammatory activity, anti-depressive, anti-obesity, **anti-allergic and anti-asthmatic activity,**
99 anti-hypercholesterolemic effects, but also anti-atherosclerotic and vasodilator activity related to this
100 compound. (Anand David et al., 2016; **Mlcek, Jurikova, Skrovankova, & Sochor, 2016**) ~~Thus the~~
101 ~~scientific interest towards quercetin and his similar compounds, like rutin, iso quercetin and catechin~~
102 ~~is growing.~~ (Anand David et al., 2016) Numerous studies have demonstrated that quercetin can act
103 as inhibitor of cyclooxygenase, lipoxygenase, NO synthase, and human reactive C protein leading to
104 a reduction of inflammatory states. (Anand David et al., 2016) Some preclinical studies conducted in
105 rats have highlighted an inhibition of both acute and chronic inflammation. (Anand David et al., 2016)
106 ~~Many cardiovascular positive effects are related to the consumption of quercetin: in fact, Quercetin~~
107 ~~can prevent platelet aggregation, can reduce oxidation caused by LDL and can reduce fat~~
108 ~~accumulation in human cells, also enhancing fat cell apoptosis.~~ (Anand David et al., 2016) As a
109 result, quercetin has the capacity to reduce atherosclerosis, hypertension and to improve the health of
110 blood vessels. (Anand David et al., 2016) Other studies reported anticancer and antiproliferative
111 properties, but also antibacterial and antiviral proprieties. (Anand David et al., 2016) Thanks to its
112 antioxidant effects, quercetin consumption has been associated to minor neuroinflammation, and less
113 reduction in cognitive performance and age depended neurodegeneration. (Anand David et al., 2016)
114 Many other studies report that quercetin has an anti-H1 and anti-H2 effect, so it can act as antiallergic
115 and anti-asthmatic molecule and can inhibit gastric acid secretion with prevention of gastritis. (Anand
116 David et al., 2016) Quercetin anti-H1, anti-H2 and antiallergic capacity is due to the inhibition of
117 enzymes, inflammatory mediators and mast cellules activation through the block of calcium,
118 prostaglandins, leukotrienes, and histamine releases. (Mlcek, Jurikova, Skrovankova, & Sochor,
119 2016)

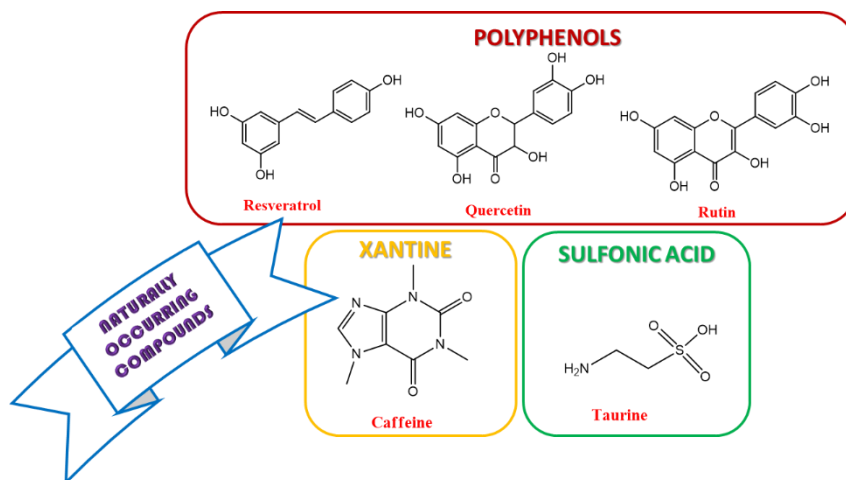
120 Resveratrol, largely present in grapes and red wine (Mollica et al., 2021) has been also recently
121 identified in beer in lower concentration; it is a stilbene derivate with high antioxidant power and
122 potential preventing activity in several human diseases. (Chiva-Blanch et al., 2011) Resveratrol
123 glycoside piceid, was found in some hop varieties and pellets used to produce beer in a concentration
124 between 0.5 to 11.7 mg/Kg; it was also detected in a concentration between 0.3 to 1.2 mg/kg, as free
125 form of *cis/trans*-resveratrol. (Chiva-Blanch et al., 2011) ~~However,~~ Hops are added in small amount
126 during beer production thus a low quantity of resveratrol is transferred in beer, that's why ~~content of~~
127 ~~*cis/trans*-resveratrol in beer is very low; thus,~~ normal beers can't be considered a dietary source of
128 resveratrol. (Chiva-Blanch et al., 2011) **The polyphenol content in beer confers** an interesting
129 nutritional value, ~~because it contains a significant polyphenol content,~~ depending on the quality and
130 quantity of malt and hops, and to the production method used. (Piazzon et al., 2010; Šibalić et al.,
131 2021) ~~Besides the aforementioned compounds beer contains many different phenolic substances,~~
132 ~~principally resveratrol, quercetin, rutin and other phenolic acids, but also tannins, proanthocyanins~~
133 ~~and amino phenolic acids. (Deng et al., 2019)~~ Phenols malt wort content depends to the temperature
134 applied during the mashing step, in fact a too long boiling step (> 60 min) decreases the total phenol
135 content and antioxidant activity of beer, because phenols polymerise during this passage. (Piazzon et
136 al., 2010; Šibalić et al., 2021) On the other hand, total phenolic content depends also to the type of
137 beer considered; in fact, there was found significant differences in phenol content of lager beer, ale
138 beer and other commercial beers. (Piazzon et al., 2010; Šibalić et al., 2021) Dealcoholized beers have
139 a lower antioxidant power and phenolic content than strong and dark beer according to the following
140 order: dealcoholized beer < lager < pilsner < wheat < ale < abbey < bock; probably because
141 dealcoholized beer is usually brewed with a minor quantity of original wort extract. (Piazzon et al.,
142 2010; Šibalić et al., 2021) Malt has a fundamental role in beer's production as antioxidant molecules,
143 phenols and starch source. (Piazzon et al., 2010; Šibalić et al., 2021) Malt provides between 86% and
144 95% of total antioxidant activity and about 80% of total phenolic content, instead hops provide less
145 amount of phenolic content without significantly affecting antioxidant capacity of beer. (Shopska et
146 al., 2021) Germination, kilning and roasting are the main processes that influence the phenolic profile
147 and antioxidant power of malts, thus determining beer properties. (Shopska et al., 2021) It's possible
148 to significantly increase beer phenolic content using different raw materials or by adding one or more
149 of them for example, diverse cereals instead of barley. Many studies described the production of beer
150 by buckwheat. (Deng et al., 2019) The best benefit obtained with buckwheat beer brewing is to
151 increase its quercetin content of about 60 times. (Deng et al., 2019) On the other hand, buckwheat is
152 gluten-free food and it is also a source of proteins, amino acids, lipids and dietary fibres. (Deng et al.,
153 2019) However, phenolic content and nutritional value of buckwheat strongly depend on different
154 factors, like buckwheat species, cultivars and thermal processes. (Bai et al., 2015; Ge & Wang, 2020;

155 Qin, Wang, Shan, Hou, & Ren, 2010) Another strategy to improve the phenolic content and
156 antioxidant power of beer is the adjunction of some fruits like cherry, peach, orange, and many others
157 during fermentation step. (Deng et al., 2020) In a recent study conducted by Deng and co-workers,
158 ale beers have been enriched with omija fruit (*Schisandra chinensis*) at diverse boiling times. (Deng
159 et al., 2020). Adding omija fruit at initial boiling step produces a typical red colour beer due to some
160 phenolic compounds formed during *Maillard reactions*, among them high quantity of lignans; for this
161 reason, omija fruit can be considerate as a valid solution to improve nutritional value of beer. (Deng
162 et al., 2020) Similar results have been obtained by Adamenko and co-workers with Cornelian cherry
163 (*Cornus mas* L.) (Adamenko, Kawa-Rygielska, & Kucharska, 2020) Authors have developed a non-
164 alcoholic beer brewed with special yeast *saccharomyces ludwigii* added to cornelian cherry juice,
165 characterised by natural aroma and sour taste. (Adamenko et al., 2020) The final beer presents a
166 significant content of phenolic compounds, anthocyanins and iridoids, lower energy value, major
167 antioxidant capacity and new sensory attributes compared to standard beer, thanks to the presence of
168 cornelian cherry juice. (Adamenko et al., 2020)

169 Beer brewing methods can be promptly selected to prepare gluten-free beers for coeliac. Common
170 beer contains gluten, which derives from the cereal used to prepare malts, (Cela et al., 2020) however
171 beer can be brewed with a gluten-free malt obtained by natural gluten-free cereals, to afford a gluten-
172 free beer. (Cela et al., 2020) An example of gluten-free cereal is teff. (Gebremariam, Zarnkow, &
173 Becker, 2013) Teff has a good malting characteristic and brewing potential. (Gebremariam et al.,
174 2013) Its attitude for malting was investigated by Grebremarian and co-workers, which have tested
175 five different teff varieties for malting procedure revealing that *Kuncho* teff variety has the best
176 enzyme activity and fermentable sugar content. (Gebremariam et al., 2013) Malting conditions, like
177 temperature and germination time are also important to obtain a high quality of teff malt. (Di Ghionno
178 et al., 2017) It was found that optimal malting condition for teff is 4 days of germination at 48% of
179 steeping degrees under 24° C. (Di Ghionno et al., 2017) Furthermore, the total di-methyl-sulphur teff
180 content can be below 7 mg/Kg to ensure a good quality malt for brewing. (Gebremariam et al., 2013)
181 Other examples of natural gluten-free cereals that can be malted are rice, maize, oat, sorghum, millet,
182 buckwheat, quinoa, and amaranth. (Cela et al., 2020).

183 Beer's nutritional value is due to the content of micro and macro-nutrients, vitamins, phenolic
184 substances, minerals, and fibres, ~~Phenolic compounds confer to beer numerous beneficial proprieties~~
185 ~~like anti-inflammatory and antioxidant activity but also a protective activity on cardiovascular and~~
186 ~~neuro-protective systems. These properties~~ that make beer an excellent base for nutritional enriched
187 ~~drinks~~ beverages brewing. The main intent of this research is to develop a new beer-based beverage
188 ~~drink~~ enriched with flavonoids (e.g. quercetin and rutin), resveratrol, caffeine and taurine according

189 to the dosages contained in food supplements **in order** to preserve the reintegrative propriety and to
190 enhance **its** nutritional value (Figure 1).



191

192 **Figure 1.** Naturally occurring compounds selected for artisanal beer's fortification.

193 HPLC-DAD analysis was performed on each sample using **not-fortified** blank beer as **control**
194 **reference**. Their antioxidant and enzyme inhibitory activities were also investigated and compared to
195 the **control** standard and commercial Porter style.

196 2. Materials and methods

197 2.1 Reagents, solvents and standard phenolic and flavonoid compound

198 Gallic acid, catechin, chlorogenic acid, *p*-OH benzoic acid, vanillic acid, epicatechin, syringic acid,
199 3-OH benzoic acid, 3-OH-4-MeO benzaldehyde, *p*-coumaric acid, rutin, sinapinic acid, *t*-ferulic acid,
200 naringin, 2,3-diMeO benzoic acid, benzoic acid, *o*-coumaric acid, quercetin, harpagoside, *t*-cinnamic
201 acid, naringenin, resveratrol, and carvacrol were purchased from Sigma Aldrich (Milan, Italy).
202 Methanol, acetonitrile (both HPLC-grade) and formic acid (pure 99%) were obtained from Carlo Erba
203 Reagenti (Milan, Italy). Double-distilled water was obtained using a Millipore Milli-Q Plus water
204 treatment system (Millipore Bedford Corp., Bedford, MA, USA).

205 2.2 Beer brewing

206 Pale Ale malts, Crystal 100 and 300 malts, East Kent Golding hop and Safale-4 yeast were bought to
207 MrMalt (Prato, IT). Quercetin, rutin, taurine and resveratrol were purchased **by** **from** Sigma (Milano,
208 IT). Coffee bean extract was prepared starting from 5 g of commercial coffee bean (Castroni, Rome,
209 IT), which was added in boiling water at 100 °C and left to stand for 5 minutes. The suspension was
210 filtered and the solution was lyophilized, the so obtained brown powder was used as such for beer's
211 fortification. Beer brewing (*all-grain*) was developed using KLARSTEIN (di Chal-Tec GmbH

212 Wallstr.16, 10179 Berlin-Deutschland) masher according to the following procedure: Malts (4.5 Kg)
213 were ground and put into KLARSTEIN masher with mineral water (25 L); temperature was brought
214 to 68 °C for 60 minutes during *meshing* step. Then temperature was raised to 72 °C for 10 minutes,
215 and to 78 °C for 5 minutes during the *mesh out* step. Then *sparging* step was raised at 78 °C for 15
216 minutes, the grain waste was removed and the first portion of hop was added (30 g). Temperature
217 was raised to 100 °C for 50 minutes (*hopping* step), then a second portion of hop was added and left
218 to boil in must **boiling** for 10 minutes. Finally, the must was cooled at to 24-25 °C. Hydrated yeast
219 (7.5 g) was added. Must was then left to ferment for 7 days at 20 °C or until completeness; after
220 fermentation step, sugar (10 g/L), resveratrol (250 mg/ 330 mL), quercetin dihydrate (200 mg/ 330
221 mL), rutin (300 mg/ 330 mL), coffee been extract (150 mg/ 330 mL), coffee been extract *plus* taurine
222 (150 mg + 105 mg/ 330 mL) were added to crude beer (20 L) during bottling and refermented for 30
223 days. After refermentation, 100 mL of each ~~mixture~~ **beer type sample** was lyophilized and analysed
224 by HPLC-DAD to evaluate their total phenol and flavonoid contents, antioxidant activity and enzyme
225 inhibitory activity.

226 2.3 HPLC-DAD analysis

227 HPLC analyses were performed on a Waters liquid chromatograph equipped with a model 600 solvent
228 pump and a 2996 photodiode array detector (PDA). *Empower v.2* Software (Waters Spa, Milford,
229 MA, USA) was used for data acquisition. A C18 reversed-phase packing column (Prodigy ODS(3),
230 4.6 × 150 mm, 5 µm; Phenomenex, Torrance, CA, USA) was used for the separation. The column
231 oven (Jetstream2 Plus) was set at 30 ± 1°C. The UV/Vis acquisition wavelength was set in the range
232 of 200 - 500 nm. The quantitative analyses were achieved at maximum wavelength for each
233 compound. The injection volume was 20 µL. The mobile phase was directly *on-line* degassed by
234 using Biotech DEGASi, mod. Compact (LabService, Anzola dell'Emilia, Italy). Gradient elution was
235 performed using the mobile phase water-acetonitrile (93:7, v:v, 3% acetic acid) as reported in Table
236 1. **Samples for HPLC-PDA analysis were prepared as follows: the lyophilized sample was weighted**
237 **and solubilize in mobile phase A (milliQ water + 3% acetic acid): B (acetonitrile +3% acetic acid)**
238 **(93:7, v: v). The samples were prepared at concentration of 1 mg/250 µL. All samples were vortexed**
239 **for 1/2 min, sonicated for 15 min and then an aliquot of 20 µL was injected in the chromatographic**
240 **system for the analysis. Table 2 reports** ~~while in Table 2 were reported~~ the retention times and the
241 maximum wavelengths used for the quantitative analyses.

242

243

Table 1. Gradient elution program used for HPLC analyses

| TIME (min) | FLOW (mL min ⁻¹) | %A | %B |
|------------|------------------------------|----|----|
| 0 | | 93 | 7 |
| 0.1 | | 93 | 7 |
| 30 | | 72 | 28 |
| 38 | | 75 | 25 |
| 45 | 1 | 2 | 98 |
| 47 | | 2 | 98 |
| 48 | | 93 | 7 |
| 58 | | 93 | 7 |

Table 2. Analytes, retention times, and maximum wavelengths used for quantitative analyses

| Analytes | Retention Times (min) | λ max |
|---------------------------------|-----------------------|---------------|
| Gallic acid | 4.99 | 271 nm |
| Catechin | 13.36 | 278 nm |
| Chlorogenic acid | 14.29 | 324 nm |
| 4-hydroxybenzoic acid | 14.71 | 256 nm |
| Vanillic acid | 17.31 | 260 nm |
| Epicatechin | 18.30 | 278 nm |
| Syringic acid | 18.50 | 274 nm |
| 3-hydroxybenzoic acid | 19.41 | 275 nm |
| 3-hydroxy-4-methoxybenzaldehyde | 22.08 | 278 nm |
| <i>p</i> -coumaric acid | 22.65 | 310 nm |
| Rutin | 25.38 | 256 nm |
| Sinapinic acid | 26.18 | 324 nm |
| <i>t</i> -ferulic acid | 27.75 | 315 nm |
| Naringin | 29.78 | 285 nm |
| 2,3-dimethoxybenzoic acid | 30.36 | 299 nm |
| Benzoic acid | 31.20 | 275 nm |
| <i>o</i> -coumaric acid | 34.81 | 276 nm |
| Quercetin | 40.57 | 367 nm |
| Harpagoside | 45.49 | 280 nm |
| <i>t</i> -cinnamic acid | 45.87 | 276 nm |
| Naringenin | 46.74 | 290 nm |
| Carvacrol | 49.95 | 275 nm |

247 **2.4 Biological activity**

248 **2.4.1 *In vitro* antioxidant assays**

249 The antioxidant assays [1,1-diphenyl-2-picrylhydrazyl (DPPH) and 2,2'-azino-bis(3-
250 ethylbenzothiazoline) 6-sulfonic acid (ABTS) radical scavenging, cupric ion reducing antioxidant
251 capacity (CUPRAC), ferric ion reducing antioxidant power (FRAP), metal chelating ability (MCA)
252 and phosphomolybdenum assay (PBD)] were previously described (Zengin and Aktumsek, 2014;
253 Mollica et al. 2021). For DPPH, ABTS, CUPRAC and FRAP assays data were expressed as mg
254 Trolox equivalents (TE)/g extract. ~~whereas in~~ MCA and PBD **were expressed as** mg EDTA
255 equivalents (EDTAE)/g extract and mmol TE/g extract respectively ~~were used~~. Total phenolic content
256 (TPC) and total flavonoid content (TFC) were determined as previously described (Zengin and
257 Aktumsek, 2014), and expressed as mg gallic acid equivalents (GAE)/g extract (TPC) and mg rutin
258 equivalents (RE)/g extract (TFC), **respectively**.

259 **2.4.2 Enzyme inhibitory activity**

260 The protocols used for acetylcholinesterase (**AChE**), butyrylcholinesterase (**BChE**), tyrosinase,
261 amylase and glucosidase assays were previously provided (Zengin, 2016; Sinan, 2021). In
262 cholinesterase assays, data were expressed as mg galanthamine equivalents (GALAE)/g extract,
263 whereas mg kojic acid equivalents (KAE)/g extract were used in tyrosinase assay. For amylase and
264 glucosidase assays, the results were reported as mmol acarbose equivalents (ACAE)/g extract.

265 **2.5. Statistical analysis**

266 All the experiments were performed in three replicates, with the results presented as mean \pm standard
267 deviation (S.D.). One-way analysis of variance (ANOVA) with Turkey's post-hoc test was
268 conducted; $p < 0.05$ was considered statistically significant. The correlation analysis between TPC,
269 TFC and biological activities was reported as Pearson's coefficients, calculated using Graph software
270 (9.0).

271 **3. Results and discussion**

272 **3.1 Determination of the total content of flavonoid and phenol compounds**

273 The total content of phenols and flavonoids in beer samples was examined using spectrophotometric
274 methods. The results are shown in the Table 3.

275 **Table 3. Beers antioxidant activity and total content of flavonoid and phenol compounds**

| Samples | TPC (mg GAE/g) | TFC (mg RE/g) | DPPH (mg TE/g) | ABTS (mg TE/g) | FRAP (mg TE/g) | CUPRAC (mg TE/g) | PHD (mmol TE/g) | MCA (mg EDTAE/g) |
|-------------------------|-------------------------|------------------------|-------------------------|------------------------|-------------------------|-------------------------|-------------------------|------------------------|
| Blank Beer control | 3.21±0.07 ^f | 0.03±0.01 ^e | 1.09±0.25 ^d | 2.63±0.25 ^d | 7.94±0.25 ^c | 15.37±0.39 ^c | 0.27±0.03 ^{ab} | na |
| Rutin beer | 3.43±0.10 ^e | 0.11±0.01 ^d | 2.13±0.24 ^{bc} | 2.77±0.19 ^d | 6.89±0.06 ^e | 14.04±0.24 ^d | 0.27±0.02 ^{ab} | na |
| Coffee beer | 3.37±0.03 ^{ef} | 0.01±0.01 ^e | 0.39±0.02 ^e | 0.14±0.02 ^f | 7.39±0.05 ^d | 14.05±0.23 ^d | 0.22±0.01 ^b | na |
| Quercetin beer | 5.21±0.04 ^c | 0.64±0.03 ^a | 2.65±0.29 ^b | 4.65±0.25 ^c | 8.70±0.17 ^b | 16.91±0.47 ^b | 0.25±0.01 ^b | 2.28±0.29 ^c |
| Resveratrol beer | 5.77±0.04 ^b | 0.25±0.04 ^c | 2.01±0.23 ^c | 7.19±0.41 ^b | 8.85±0.14 ^b | 17.26±0.20 ^b | 0.27±0.04 ^{ab} | 3.60±0.53 ^b |
| Coffee and taurine beer | 7.18±0.04 ^a | 0.40±0.04 ^b | 5.62±0.30 ^a | 8.59±0.39 ^a | 13.14±0.09 ^a | 26.18±0.45 ^a | 0.32±0.02 ^a | 5.17±0.63 ^a |
| Commercial beer | 3.89±0.03 ^d | 0.13±0.03 ^d | 0.68±0.08 ^{de} | 1.03±0.08 ^e | 6.54±0.07 ^e | 13.33±0.20 ^d | 0.21±0.02 ^b | 0.34±0.05 ^d |

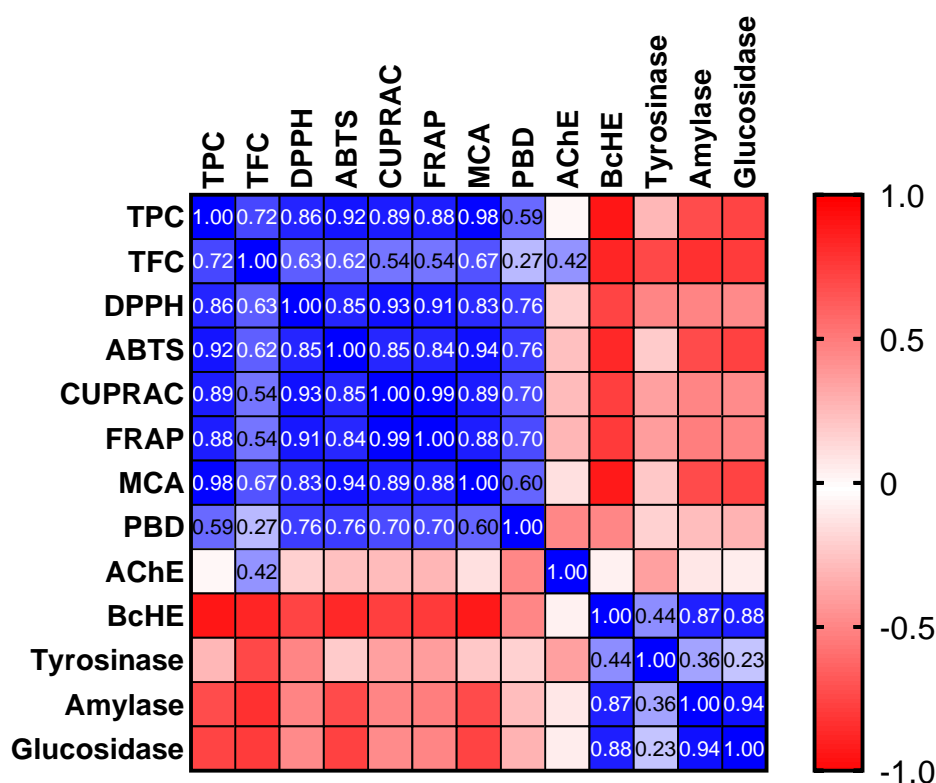
276 * Values expressed are means ± S.D. of three parallel measurements. TPC: Total phenolic content; TFC: Total flavonoid
277 content; PHD: Phosphomolybdenum; MCA: Metal chelating; FRAP: Ferric reducing antioxidant power; CUPRAC:
278 Cupric ion-reducing activity; TE: Trolox equivalent; EDTAE: EDTA equivalent. GAE: Gallic acid equivalent; RE: Rutin
279 equivalent; nt: not tested; na: no active. Different letters in the same column indicate significant differences in the tested
280 samples (p<0.05)

281 The content of best result in terms of total phenols follows this order of magnitude: content is in
282 coffee / taurine-enriched beer was determined as (7.18 mg GAE/g) > followed by resveratrol-enriched
283 beer (5.77 mg GAE/g) > quercetin-enriched beer (5.21 mg GAE/g). In terms of total flavonoid
284 content, the richest sample was quercetin fortified beer (0.64 mg RE / g), followed by coffee / taurine
285 (0.40 mg RE/g) and resveratrol (0.25 mg RE/g). The lowest values for total flavonoid were found in
286 control beer (0.03 mg RE/g) and coffee beer (0.01 mg RE/g). The enrichment with quercetin,
287 resveratrol or coffee / taurine clearly increased the total phenol content compared to the control beer.
288 Diverse antioxidant activities could be explained with total content of phenolic compounds, different
289 preparation techniques and geographical locations of malt and hops (Vidyalakshmi et al., 2022;
290 Rahman et al., 2020; Zhao et al., 2010; Nardini and Garaguso, 2020; Nescitelli et al. 2016).

291 3.2 Antioxidant activity

292 The antioxidant properties of the tested beer samples were evaluated by different chemical assays.
293 The results are depicted in Table 3. DPPH and ABTS radicals are widely used to assess the radical
294 scavenging abilities of plant extracts. In both assays, the best scavenging ability was determined in
295 coffee extract/taurine enriched beer (DPPH: 5.62 mg TE/g and ABTS: 8.59 mg TE/g), followed by
296 quercetin enriched beer (2.65 mg TE/g) in the DPPH assay and resveratrol enriched beer (7.19 mg
297 TE/g) in the ABTS assay. The electron-donating abilities of the tested beer samples were examined
298 by reducing power assays, namely CUPRAC and FRAP. The assays are based on the reduction of
299 cupric/ferric to copper/ferrous form by antioxidant compounds. In both of them, the best reduction

300 ability was observed for coffee extract/taurine enriched beer (CUPRAC: 26.18 mg TE/g and FRAP:
 301 13.14 mg TE/g). The phosphomolybdenum assay consists in the conversion of Mo (VI) to Mo (V) by
 302 antioxidant compounds, thus forming a blue Mo (V) / phosphate complex **which has a maximum of**
 303 **absorbance at 695 nm**. In this sense, the assays could be considered as a reducing power assay, as
 304 well CUPRAC and FRAP but all antioxidant components are susceptible to this reaction, thus the
 305 assay is considered as an antioxidant assay. The best ability was observed for coffee extract/taurine
 306 enriched beer in phosphomolybdenum assay (0.32 mmol TE/g), while other beer samples showed
 307 very similar activities (0.21-0.27 mmol TE/g). These results are consistent with the total phenolic
 308 content in the extracts, further confirming that they actively contribute to the free radical and reducing
 309 abilities of the tested beer samples. The correlation analysis was performed between the test systems
 310 and the obtained results (Figure 2).



311

312 **Figure 2.** Pearson's correlation values (R) in the performed biological activity assays; AChE,
 313 acetylcholinesterase; BcHE, butyrylcholinesterase; CUPRAC, cupric ion reducing antioxidant
 314 capacity; DPPH, 1,1-diphenyl-2-picrylhydrazyl; FRAP, ferric ion reducing antioxidant power; MCA,
 315 metal chelating activity; PBD, phosphomolybdenum assay; TFC, total flavonoid content; TPC, total
 316 phenolic content.

317 A strong correlation between total phenolic and radical scavenging/reducing power assays ($R > 0.7$)
 318 was found, in line with several literature reports (Nardini and Garaguso, 2020; Moura-Nunes et al.,

319 2016; Baigts-Allende et al., 2021). Chelation of transition metals is closely linked to manage the
320 production of hydroxyl radicals in Fenton reaction. In the current study, the best chelating ability was
321 detected by coffee/taurine enriched beer (5.17 mg EDTAE/g), followed by resveratrol (3.60 mg
322 EDTAE/g), **and** quercetin-enriched beers ~~and commercial beer (0.34 mg EDTAE/g)~~ (2.28 mg
323 EDTAE/g). Overall our data demonstrate that the enriched-beer samples exhibit stronger antioxidant
324 abilities than the ~~blank control and commercial beers~~. The use of quercetin and coffee extract/taurine
325 in the artisanal beer's fortification increases its antioxidant properties, representing an extremely
326 appealing starting point for the development of **low-fermentation** beer based-functional **drinks**.

327

328 **3.3 Identification and quantification of flavonoid and phenol compounds by HPLC-DAD**

329 ~~Samples for HPLC-PDA analysis were prepared as follows: the lyophilized sample was weighted and~~
330 ~~solubilize in mobile phase A (milliQ water + 3% acetic acid): B (acetonitrile + 3% acetic acid) (93:7,~~
331 ~~v: v). The samples were prepared at concentration of 1mg/250 μ L. All samples were vortexed for 1/2~~
332 ~~min, sonicated for 15 min and then an aliquot of 20 μ L was injected in the chromatographic system~~
333 ~~for the analysis. In Table 4 are reported the phenolic quantitative data (in μ g/mg dry extract) observed~~
334 ~~for the different beer samples: beer control, rutin beer, coffee beer, quercetin beer, resveratrol beer,~~
335 ~~and coffee-taurin beer, respectively. The reported values are mean \pm standard deviation of three~~
336 ~~independent measures. The value below Limit of Quantification (0.02 μ g/mL) have been reported as~~
337 ~~BLQ (Below Limit of Quantification). Detectable quantity of rutin ($0.83 \pm 0.7 \mu$ g/mg) is in rutin beer~~
338 ~~sample, *p*-coumaric acid ($0.05 \pm 0.01 \mu$ g/mg) and sinapinic acid ($0.19 \pm 0.01 \mu$ g/mg) are in coffee beer~~
339 ~~sample, quercetin ($0.19 \pm 0.01 \mu$ g/mg) in quercetin enriched-beer and chlorogenic acid ($0.24 \pm$~~
340 ~~0.01μ g/mg) in coffee extract/taurine beer sample.~~

Table 4. Flavonoids and phenols quantification by HPLC-DAD analysis

| Concentration ($\mu\text{g}/\text{mg}$) | Galic acid | Catechin | Chlorogenic acid | <i>p</i> -OH benzoic acid | Vanillic acid | Epicatechin | Syringic acid | 3-OH benzoic acid | 3-OH-4-MeO benzaldehyde | <i>p</i> -coumaric acid | Rutin | Sinapinic acid | <i>t</i> -ferulic acid | Naringin | 2,3-diMeO benzoic acid | Benzoic acid | <i>o</i> -coumaric acid | Quercetin | Harpagoside | <i>Trans</i> -cinnamic acid | Carvacrol |
|--|------------|----------|---------------------|---------------------------|---------------|-------------|---------------|-------------------|-------------------------|-------------------------|------------------------|---------------------|------------------------|----------|------------------------|--------------|-------------------------|---------------------|-------------|-----------------------------|-----------|
| BEER SAMPLES | | | | | | | | | | | | | | | | | | | | | |
| Beer control | | | | | | | | | | | | | | | | | | | | | |
| Rutin beer | | | | | | | | | | | 0.83 (± 0.07) | | | | | | | | | | |
| Coffee beer | | | | | | | | | | 0.05 (± 0.01) | | 0.19 (± 0.01) | | | | | | | | | |
| Quercetin beer | | | | | | | | | | BLQ | | | | | | | | 0.27 (± 0.02) | | | |
| Resveratrol beer | | | | | | | | | | | | | | | | | | | | | |
| Coffee-taurin beer | | | 0.24 (± 0.01) | | | | | | | BLQ | | | | | | | | | | | |

BLQ: below Limit of Quantification.

1 3.4 Enzyme inhibitory activity

2 The enzyme inhibition theory plays an important role in treating global health problems such as
3 diabetes mellitus, Alzheimer's disease and obesity. In particular, the inhibition of some key enzymes
4 (*e.g.* cholinesterase, amylase, and lipase) can reduce the pathological observations in these diseases
5 (Rauf and Jehan, 2017). Several drugs have been manufactured as enzyme inhibitors in the
6 pharmaceutical industry, but most of them show side effects such as toxicity and gastrointestinal
7 diseases (Jagadeesan et al., 2022; Meziat et al., 2021), thus the discovery of new and natural enzyme
8 inhibitors has become one of the most attractive topic in the scientific platform (Mollica et al. 2018).
9 According to these, we investigated the enzyme inhibitory properties of the tested beer samples using
10 cholinesterases, tyrosinase, amylase and glucosidase as target enzymes. The results are shown in
11 Table 5. In AChE inhibition assay, two beer samples (quercetin enriched ~~and commercial~~-beer) are
12 able to inhibit the enzyme at 2.66 and 3.07 mg GALAE/g, respectively. All beer samples are active
13 against BChE, the best result was obtained by rutin-enriched beer (7.95 mg RE/g) while the lowest
14 BChE inhibitory effect was observed for quercetin-enriched beer (5.29 mg GALAE/g). Tyrosinase is
15 a main enzyme involved in the synthesis of melanin; its inhibition represents a key strategy in the
16 treatment of hyperpigmentation (Della Valle et al. 2020). The best tyrosinase inhibitory effect was
17 detected in resveratrol-enriched beer (59.19 mg KAE/g), while the weakest ability was provided by
18 quercetin-enriched beer (49.90 mg KAE/g). Other beer samples exhibited similar tyrosinase
19 inhibitory effects (53.00-56.74 mg KAE/g). All samples showed very similar abilities against amylase
20 and glucosidase enzymes. To the best of our knowledge, the information regarding the enzyme
21 inhibitory effect of beer samples are very scarce in literature (Merino et al., 2018; Szwajgier, 2013),
22 therefore the present study provides insightful data in this field, thus paving the way to the
23 development of **low-fermentation** functional beers ~~samples~~ in the food and drink industries.

24 **Table 5. Beer's enzymatic inhibition activity.**

| Samples | AChE (mg GALAE/g) | BChE (mg GALAE/g) | Tyrosinase (mg KAE/g) | Amylase (mmol ACAE/g) | Glucosidase (mmol ACAE/g) |
|----------------------------|------------------------|-------------------------|-------------------------|-------------------------|---------------------------|
| Beer -Control | na | 7.87±0.16 ^{ab} | 55.16±0.52 ^b | 0.17±0.01 ^{ab} | 1.20±0.01 ^a |
| Rutin Beer | na | 7.95±0.22 ^a | 55.86±0.60 ^b | 0.17±0.01 ^a | 1.20±0.01 ^a |
| Coffee Beer | na | 7.52±0.10 ^b | 56.74±0.19 ^b | 0.17±0.01 ^{ab} | 1.20±0.01 ^a |
| Quercetin Beer | 2.66±0.03 ^b | 5.29±0.24 ^{cd} | 49.90±0.85 ^d | 0.11±0.01 ^c | 1.14±0.01 ^c |
| Resveratrol Beer | na | 5.69±0.03 ^c | 59.19±0.44 ^a | 0.12±0.02 ^c | 1.14±0.01 ^c |
| Coffee and taurine beer | na | 5.14±0.07 ^d | 53.00±0.45 ^c | 0.14±0.01 ^{bc} | 1.17±0.01 ^b |
| Commercial beer | 3.07±0.01 ^a | 7.83±0.06 ^{ab} | 56.72±1.30 ^b | 0.18±0.01 ^a | 1.20±0.01 ^a |

25 * Values expressed are means \pm S.D. of three parallel measurements. AChE: Acetylcholinesterase; BChE:
26 Butyrylcholinesterase; GALAE: Galatamine equivalent; KAE: Kojic acid equivalent; ACAE: Acarbose equivalent; na:
27 not active. Different letters in the same column indicate significant differences in the tested samples ($p < 0.05$)

28 4. Conclusions

29 In this study artisanal beers brewed by *all grain* method and low fermentation ~~with fresh and pleasant~~
30 ~~flavour~~ have been prepared. Samples were analysed by HPLC-DAD system in order to determine the
31 polyphenolic profile. The highest total polyphenol content was found in resveratrol, quercetin and in
32 taurine /plus coffee extract enriched beers. HPLC-DAD analysis reveals that flavonoid enriched beer
33 has a content of bioactive substances like common food supplement and this result highlights the
34 possibility to use enriched beer as vehicle of bioactive compounds. ~~Antioxidant activity and chelation~~
35 ~~capacity was evaluated *in vitro* trough DPPH scavenging and FRAP test;~~ The major antioxidant
36 activity was found in resveratrol beer and coffee-aurine beer. ~~Enzymatic inhibition activity was~~
37 ~~detected *in vitro* to estimate the inhibition capacity against some enzymes involved in glucose~~
38 ~~metabolism and neurodegeneration.~~ According to our study flavonoids-enriched beer has a good
39 inhibitory activity on enzyme tyrosinase, moderate activity against BChE and very low activity on
40 other enzymes. These data encourage further studies on the development of enriched beers **and in the**
41 **optimization of food supplements**; the procedure applied in this work ~~can be extended to~~ **for** the
42 production of invigorating or energizing **drinks** beverages, as well as to prepare a lyophilized matrix
43 ~~for cosmetics and anti-aging products.~~

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47 References

- 48 Adamenko, K., Kawa-Rygielska, J., & Kucharska, A. Z. (2020). Characteristics of Cornelian cherry
49 sour non-alcoholic beers brewed with the special yeast *Saccharomyces ludwigii*. *Food*
50 *Chemistry*, 312, 125968. <https://doi.org/10.1016/j.foodchem.2019.125968>
- 51 Anand David, A. V., Arulmoli, R., & Parasuraman, S. (2016). Overviews of biological importance
52 of quercetin: A bioactive flavonoid. *Pharmacognosy Reviews*, 10(20), 84–89.
53 <https://doi.org/10.4103/0973-7847.194044>
- 54 Bai, C. Z., Feng, M. L., Hao, X. L., Zhong, Q. M., Tong, L. G., & Wang, Z. H. (2015). Rutin,
55 quercetin, and free amino acid analysis in buckwheat (*Fagopyrum*) seeds from different

- 56 locations. *Genetics and Molecular Research*, 14(4), 19040–19048.
57 <https://doi.org/10.4238/2015.December.29.11>
- 58 Baigts-Allende, D. K., Pérez-Alva, A., Ramírez-Rodrigues, M. A., Palacios, A., & Ramírez-
59 Rodrigues, M. M. (2021). A comparative study of polyphenolic and amino acid profiles of
60 commercial fruit beers. *Journal of Food Composition and Analysis*, 100, 103921.
- 61 Bertuzzi, T., Mulazzi, A., Rastelli, S., Donadini, G., Rossi, F., & Spigno, G. (2020). Targeted
62 healthy compounds in small and large-scale brewed beers. *Food Chemistry*, 310, 125935.
63 <https://doi.org/10.1016/j.foodchem.2019.125935>
- 64 Cela, N., Condelli, N., Caruso, M. C., Perretti, G., Cairano, M. Di, Tolve, R., & Galgano, F. (2020).
65 *Gluten-Free Brewing : Issues and Perspectives*. 1–26.
- 66 Chiva-Blanch, G., Urpi-Sarda, M., Rotchés-Ribalta, M., Zamora-Ros, R., Llorach, R., Lamuela-
67 Raventós, R. M., ... Andrés-Lacueva, C. (2011). Determination of resveratrol and piceid in
68 beer matrices by solid-phase extraction and liquid chromatography-tandem mass spectrometry.
69 *Journal of Chromatography A*, 1218(5), 698–705.
70 <https://doi.org/10.1016/j.chroma.2010.12.012>
- 71 de Gaetano, G., Costanzo, S., Di Castelnuovo, A., Badimon, L., Bejko, D., Alkerwi, A., ...
72 Iacoviello, L. (2016). Effects of moderate beer consumption on health and disease: A
73 consensus document. *Nutrition, Metabolism and Cardiovascular Diseases*, 26(6), 443–467.
74 <https://doi.org/10.1016/j.numecd.2016.03.007>
- 75 Della Valle, A., Dimmito, M.P., Zengin, G., Pieretti, S., Mollica, A., Locatelli, M., Cichelli, A.,
76 Novellino, E., Ak, G., Yerlikaya, S., Baloglu, M.C., Celik Altunoglu, Y., Stefanucci, A.
77 (2020). Exploring the Nutraceutical Potential of Dried Pepper *Capsicum annum* L. on Market
78 from Altino in Abruzzo Region. *Antioxidants*, 9, 400. <https://doi.org/10.3390/antiox9050400>
- 79 Deng, Y., Lim, J., Lee, G. H., Hanh Nguyen, T. T., Xiao, Y., Piao, M., & Kim, D. (2019). Brewing
80 rutin-enriched lager beer with buckwheat malt as adjuncts. *Journal of Microbiology and*
81 *Biotechnology*, 29(6), 877–886. <https://doi.org/10.4014/jmb.1904.04041>
- 82 Deng, Y., Lim, J., Nguyen, T. T. H., Mok, I. K., Piao, M., & Kim, D. (2020). Composition and
83 biochemical properties of ale beer enriched with lignans from *Schisandra chinensis* Baillon
84 (omija) fruits. *Food Science and Biotechnology*, 29(5), 609–617.
85 <https://doi.org/10.1007/s10068-019-00714-5>

- 86 Di Ghionno, L., Marconi, O., Lee, E. G., Rice, C. J., Sileoni, V., & Perretti, G. (2017). Gluten-Free
87 Sources of Fermentable Extract: Effect of Temperature and Germination Time on Quality
88 Attributes of Teff [*Eragrostis tef* (zucc.) Trotter] Malt and Wort. *Journal of Agricultural and*
89 *Food Chemistry*, 65(23), 4777–4785. <https://doi.org/10.1021/acs.jafc.7b01717>
- 90 Durga Prasad, C.G., Vidyalakshmi, R., & Baskaran, N. (2022). Influence of *Pichia myanmarensis* in
91 fermentation to produce quinoa based non-alcoholic beer with enhanced antioxidant
92 activity. *Journal of Cereal Science*, 103, 103390.
- 93 Ge, R. H., & Wang, H. (2020). Nutrient components and bioactive compounds in tartary buckwheat
94 bran and flour as affected by thermal processing. *International Journal of Food Properties*,
95 23(1), 127–137. <https://doi.org/10.1080/10942912.2020.1713151>
- 96 Gebremariam, M. M., Zarnkow, M., & Becker, T. (2013). Effect of teff (*Eragrostis tef*) variety and
97 storage on malt quality attributes. *Journal of the Institute of Brewing*, 119(1–2), 64–70.
98 <https://doi.org/10.1002/jib.65>
- 99 Jagadeesan, G., Muniyandi, K., Manoharan, A. L., Nataraj, G., & Thangaraj, P. (2022).
100 Understanding the bioaccessibility, α -amylase and α -glucosidase enzyme inhibition kinetics of
101 *Allmania nodiflora* (L.) R. Br. ex Wight polyphenols during in vitro simulated digestion. *Food*
102 *Chemistry*, 372, 131294.
- 103 Liu, S. Q., & Quek, A. Y. H. (2016). Evaluation of beer fermentation with a novel yeast *Williopsis*
104 *saturnus*. *Food Technology and Biotechnology*, 54(4), 403–412. <https://doi.org/10.17113/ft>
105 [b.54.04.16.4440](https://doi.org/10.17113/ft)
- 106 Maisto, M., Annunziata, G., Schiano, E., Piccolo, V., Iannuzzo, F., Santangelo, R., ... Grieco, P.
107 (2021). Potential functional snacks: Date fruit bars supplemented by different species of
108 *Lactobacillus* spp. *Foods*, 10(8), 1–9. <https://doi.org/10.3390/foods10081760>
- 109 Merino, P., Santos-López, J. A., Mateos, C. J., Meseguer, I., Garcimartín, A., Bastida, S., ... &
110 González-Muñoz, M. J. (2018). Can nonalcoholic beer, silicon and hops reduce the brain
111 damage and behavioral changes induced by aluminum nitrate in young male Wistar rats?. *Food*
112 *and Chemical Toxicology*, 118, 784-794.
- 113 Meziat, L., Bachir-bey, M., Bensouici, C., Saci, F., Boutiche, M., & Louaileche, H. (2021).
114 Assessment of inhibitory properties of flavonoid-rich fig (*Ficus carica* L.) peel extracts against
115 tyrosinase, α -glucosidase, urease and cholinesterases enzymes, and relationship with
116 antioxidant activity. *European Journal of Integrative Medicine*, 43, 101272.

- 117 Mlcek, J., Jurikova, T., Skrovankova, S., & Sochor, J. (2016). Quercetin and its anti-allergic
118 immune response. *Molecules*, *21*(5), 1–15. <https://doi.org/10.3390/molecules21050623>
- 119 Mollica, A., Scioli, G., Della Valle, A., Cichelli, A., Novellino, E., Bauer, M., Kamysz, W.,
120 Llorent-Martínez, E.J., Fernández-de Córdoba, M.L., Castillo-López, R., Ak, G., Zengin, G.,
121 Pieretti, S., Stefanucci, A. (2021). Phenolic Analysis and In Vitro Biological Activity of Red
122 Wine, Pomace and Grape Seeds Oil Derived from *Vitis vinifera* L. cv. Montepulciano
123 d’Abruzzo. *Antioxidants*, *10*, 1704. <https://doi.org/10.3390/antiox10111704>
- 124 Mollica, A., Stefanucci, A., Zengin, G., Locatelli, M., Macedonio, G., Orlando, G., Ferrante, C.,
125 Menghini, L., Recinella, L., Leone, S., Chiavaroli, A., Leporini, L., Di Nisio, C., Brunetti, L.,
126 Tayrab, E., Ali, I., Musa, T.H., Musa, H.H., Ahmed, A.A. (2018) Polyphenolic composition,
127 enzyme inhibitory effects ex-vivo and in-vivo studies on two Brassicaceae of north-central
128 Italy. *Biomedicine & Pharmacotherapy*, *107*, 129-138.
129 <https://doi.org/10.1016/j.biopha.2018.07.169>.
- 130 Moura-Nunes, N., Brito, T. C., da Fonseca, N. D., de Aguiar, P. F., Monteiro, M., Perrone, D., &
131 Torres, A. G. (2016). Phenolic compounds of Brazilian beers from different types and styles
132 and application of chemometrics for modeling antioxidant capacity. *Food chemistry*, *199*, 105-
133 113.
- 134 Nardini, M., & Garaguso, I. (2020). Characterization of bioactive compounds and antioxidant
135 activity of fruit beers. *Food chemistry*, *305*, 125437.
- 136 Nescatelli, R., Carradori, S., Marini, F., Caponigro, V., Bucci, R., De Monte, C., Mollica, A.,
137 Mannina, L., Ceruso, M., Supuran, C.T., Secci, D. (2017). Geographical characterization by
138 MAE-HPLC and NIR methodologies and carbonic anhydrase inhibition of Saffron
139 components. *Food Chem.* *221*(15), 855-863. doi: 10.1016/j.foodchem.2016.11.086.
- 140 Olaniran, A. O., Hiralal, L., Mokoena, M. P., & Pillay, B. (2017). Flavour-active volatile
141 compounds in beer: production, regulation and control. *Journal of the Institute of Brewing*,
142 *123*(1), 13–23. <https://doi.org/10.1002/jib.389>
- 143 Piazzon, A., Forte, M., & Nardini, M. (2010). Characterization of phenolics content and antioxidant
144 activity of different beer types. *Journal of Agricultural and Food Chemistry*, *58*(19), 10677–
145 10683. <https://doi.org/10.1021/jf101975q>
- 146 Qin, P., Wang, Q., Shan, F., Hou, Z., & Ren, G. (2010). Nutritional composition and flavonoids
147 content of flour from different buckwheat cultivars. *International Journal of Food Science and*

- 148 *Technology*, 45(5), 951–958. <https://doi.org/10.1111/j.1365-2621.2010.02231.x>
- 149 Rahman, M. J., Liang, J., Eskin, N. M., Eck, P., & Thiyam-Holländer, U. (2020). Identification of
150 hydroxycinnamic acid derivatives of selected canadian and foreign commercial beer extracts
151 and determination of their antioxidant properties. *LWT*, 122, 109021.
- 152 Rauf, A., & Jehan, N. (2017). Natural products as a potential enzyme inhibitors from medicinal
153 plants. *Enzyme Inhibitors and Activators; InTech: Rijeka, Croatia*, 165-177.
- 154 Schulz, M., Seraglio, S. K. T., Della Betta, F., Nehring, P., Valese, A. C., Daguer, H., ... Fett, R.
155 (2019). Blackberry (*Rubus ulmifolius* Schott): Chemical composition, phenolic compounds
156 and antioxidant capacity in two edible stages. *Food Research International*, 122(2018), 627–
157 634. <https://doi.org/10.1016/j.foodres.2019.01.034>
- 158 Shopska, V., Denkova-Kostova, R., Dzhivoderova-Zarcheva, M., Teneva, D., Denev, P., & Kostov,
159 G. (2021). Comparative study on phenolic content and antioxidant activity of different malt
160 types. *Antioxidants*, 10(7). <https://doi.org/10.3390/antiox10071124>
- 161 Šibalić, D., Planinić, M., Jurić, A., Bucić-Kojić, A., & Tišma, M. (2021). Analysis of phenolic
162 compounds in beer: from raw materials to the final product. *Chemical Papers*, 75(1), 67–76.
163 <https://doi.org/10.1007/s11696-020-01276-1>
- 164 Sinan, K.I., Dall'Acqua, S., Ferrarese, I., Mollica, A., Stefanucci, A., Glamočlija, J., Sokovic, M.,
165 Nenadić, M., Aktumsek, A., Zengin, G. (2021). LC-MS Based Analysis and Biological
166 Properties of *Pseudocedrela kotschyi* (Schweinf.) Harms Extracts: A Valuable Source of
167 Antioxidant, Antifungal, and Antibacterial Compounds. *Antioxidants*, 10, 1570.
168 <https://doi.org/10.3390/antiox10101570>
- 169 Szwajgier, D. (2013). Inhibition of cholinesterases by phenolic acids detected in beer: A dose-
170 response model approach. *African Journal of Biotechnology*, 12(14), 1675-1681.
- 171 Zengin, G., & Aktumsek, A. (2014). Investigation of antioxidant potentials of solvent extracts from
172 different anatomical parts of *Asphodeline anatolica* E. Tuzlaci: An endemic plant to
173 Turkey. *African Journal of Traditional, Complementary and Alternative Medicines*, 11(2),
174 481-488.
- 175 Zengin, G. (2016). A study on in vitro enzyme inhibitory properties of *Asphodeline anatolica*: New
176 sources of natural inhibitors for public health problems. *Industrial Crops and Products*, 83, 39-
177 43.

178 Zhao, H., Chen, W., Lu, J., & Zhao, M. (2010). Phenolic profiles and antioxidant activities of
179 commercial beers. *Food Chemistry*, *119*(3), 1150-1158.

180

181