Ther Adv Hematol

2022, Vol. 13: 1-20

DOI: 10.1177/ 20406207221090882

© The Author(s), 2022. Article reuse guidelines: sagepub.com/journalspermissions

Correspondence to: Fabio Ciceri

Hematology and Bone Marrow Transplantation Unit, IRCCS San Raffaele Scientific Institute, via Olgettina 60, Milano 20132, Italy Vita-Salute San Raffaele University, Milan, Italy

ciceri.fabio@hsr.it

Carmine Liberatore Maria Teresa Lupo-Stanghellini Matteo Giovanni Carrabba Raffaella Greco Sarah Marktel Andrea Assanelli Francesca Farina Consuelo Corti Massimo Bernardi Jacopo Peccatori Haematology and Bone Marrow Transplant Unit. IRCCS San Raffaele Scientific Institute, Milan, Italv

Francesca Lorentino

Haematology and Bone Marrow Transplant Unit, IRCCS San Raffaele Scientific Institute, Milan, Italy

PhD Program in Public Health, Department of Medicine and Surgery, University of Milano Bicocca, Milan, Italy

Luca Vago

Haematology and Bone Marrow Transplant Unit, IRCCS San Raffaele Scientific Institute, Milan, Italy

Unit of Immunogenetics, Leukemia Genomics and Immunobiology, IRCCS San Raffaele Scientific Institute, Milan, Italy

Azacitidine and donor lymphocytes infusions in acute myeloid leukemia and myelodysplastic syndrome relapsed after allogeneic hematopoietic stem cell transplantation from alternative donors

Carmine Liberatore, Maria Teresa Lupo Stanghellini*, Francesca Lorentino, Luca Vago, Matteo Giovanni Carrabba, Raffaella Greco, Sarah Marktel, Andrea Assanelli, Francesca Farina, Consuelo Corti, Massimo Bernardi, Jacopo Peccatori, Katja Sockel, Jan Moritz Middeke, Johannes Schetelig, Anika Bergmann, Christina Rautenberg, Fabio Ciceri^D, Martin Bornhäuser, Thomas Schroeder⁺ and Friedrich Stölzel⁺

Abstract

Introduction: Azacitidine (AZA) either single-agent or with donor lymphocytes infusions (DLI) has been used as a salvage treatment for acute myeloid leukemia (AML) and myelodysplastic syndromes (MDS) relapsing after allogeneic hematopoietic stem cell transplantation (HSCT). To date, the majority of data come from patients relapsed after HSCT from full-matched donors.

Methods: We report a multicenter, collaborative, retrospective analysis of 71 patients with hematologic (n = 40, 56%) and molecular relapse (n = 31, 44%) of myeloid neoplasms after HSCT from alternative donors (mismatched unrelated, n = 39, 55%; haploidentical, n = 29, 41%) consecutively treated at three European centers with AZA ± DLI.

Results: Median time from HSCT to relapse was 9 months. Additional DLI were given to 33 patients (46%). After a median of four cycles, overall response rate (ORR) was 49% and complete response (CR) rate was 38%. CR lasted for a median of 17 months (range 5–89 months). Median follow-up in the entire cohort was 11 months (range 1–115 months). Event-free survival (EFS) and overall survival (OS) at 1 year were 26% and 53%, respectively. Treatment of molecular relapse granted higher CR rate (65% *versus* 15%; p=0.0001), 1-year EFS (43% *versus* 13%; p=0.006), and 1-year OS (79% *versus* 34%; p<0.001) compared to hematologic relapses. Addition of DLI resulted in significantly higher responses and longer 1-year EFS and OS (Mantel–Byar test, p=0.004 and p=0.002, respectively). When applied to our cohort, the APSS-R score confirmed its ability to stratify patients into distinct prognostic groups with significantly different response rates (p=0.0005) and survival (p<0.0001). Treatment was well tolerated, with the incidence of late acute and chronic graft-versus-host disease of 27% and 18%, respectively.

Conclusion: AZA \pm DLI proved feasible and effective in AML and MDS relapsing after HSCT from alternative donors. Despite modest efficacy among hematologic relapses, pre-emptive treatment with AZA \pm DLI fared better in molecular relapse. Additional DLI contributed to improving efficacy and ensuring longer survival.

Keywords: acute myeloid leukemia, allogeneic hematopoietic stem cell transplantation, alternative donor, azacitidine, myelodysplastic syndrome, post-transplantation relapse

Received: 10 January 2022; revised manuscript accepted: 14 March 2022.

journals.sagepub.com/home/tah



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

Katja Sockel Jan Moritz Middeke Johannes Schetelig Martin Bornhäuser Friedrich Stölzel Medizinische Klinik und Poliklinik I, Universitätsklinikum Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany

Anika Bergmann

Department of Hematology, Oncology and Clinical Immunology, University Hospital Duesseldorf, Medical Faculty, Heinrich Heine – University, Duesseldorf, Germany

Christina Rautenberg Thomas Schroeder Department of

Hematology, Oncology and Clinical Immunology, University Hospital Duesseldorf, Medical Faculty, Heinrich Heine – University, Duesseldorf, Germany

Department of Hematology and Stem Cell Transplantation, West German Cancer Center Essen, University Hospital Essen, Essen, Germany *first co-authors *last co-authors

Introduction

Allogeneic hematopoietic stem cell transplantation (HSCT) still represents the only potentially curative treatment option for the majority of patients with acute myeloid leukemia (AML) and high-risk myelodysplastic syndromes/myeloproliferative neoplasms (MDS/MPN). Over the last decades, the outcome of HSCT has progressively improved, due to both advances in transplantacare.1,2 tion techniques and supportive Nonetheless, the incidence of disease relapse has largely remained unchanged and still represents the major cause of treatment failure.³ Overall, about 30% of patients undergoing HSCT for AML and MDS will finally relapse, although rates up to 80% are reported depending on disease, patient, and transplantation features.³⁻⁷ The prognosis of relapsed patients remains poor. After different salvage treatments, such as intensive chemotherapy (IC), donor lymphocytes infusions (DLI), or second HSCT, complete response (CR) is obtained only in about 30% of patients and fewer than 20% achieves long-term outcomes.^{3–7} Interestingly, long-term survival appeared to rely mainly on the achievement of CR with cytoreductive therapies, followed by immunological consolidation with either DLI or subsequent HSCT.⁵⁻⁸ However, IC are burdened by severe toxicities, representing a major limit for less fit and heavily pre-treated patients, especially in the early post-transplantation period.

The hypomethylating agents (HMA), azacitidine (AZA), and decitabine either single-agent or in combination with DLI have been increasingly used in last years as salvage treatment for AML and MDS relapsing after HSCT.9-31 The rationale relies on both anti-leukemic and immunomodulating effects.³² HMA have shown to induce the upregulation of several leukemia-associated and minor-histocompatibility antigens that were previously epigenetically silenced, and of HLA class-I, HLA class-II, and costimulatory molecules, and to increase the responses of tumor-specific CD8 + T cells.³³⁻⁴³ AZA is also reported to upregulate inhibitory pathways and to induce the expansion of regulatory T cells.44-47 Despite potentially hampering tumor-specific T cells alloreactivity, immunoregulatory properties might in part prevent or attenuate graft-versus-host disease (GvHD) in the post-transplantation setting.^{48,49} HMA also showed lower toxicity compared to IC. Focusing on AZA, the majority of data come from retrospective studies and four non-randomized

prospective trials. Despite heterogeneity, the most relevant studies showed tolerability and efficacy, with response rates of 15-50% in hematologic relapses and 27-67% in case of pre-emptive therapy for molecular relapse. Overall survival (OS) at 2 years was 12-54% in the former group and 25-69% in the latter group (Table 1).^{9-26,31}

Notably, the patients analyzed till now had mainly received HSCT from fully matched sibling donors (MSD) and matched unrelated donors (MUD), while few mismatched unrelated donors (MMUD) and haploidentical donors (haplo) have been reported, without specific subgroup analysis. Therefore, less is known about AZA \pm DLI after HSCT from alternative donors, where HLA mismatches could represent a matter of concern.⁵⁰ In this setting, immunomodulating effects of both AZA and DLI could elicit, besides the expected graft-versus-leukemia effect, a potentially severe and life-threatening GvHD.

We report a multicenter, collaborative, retrospective analysis of 71 patients with AML and MDS/ MPN relapsed after HSCT from alternative donors and consecutively treated with AZA \pm DLI. The aim of the study is to evaluate efficacy and feasibility of treatment to identify factor predictive of response and survival and to compare results with the literature.

Methods

Study design

From December 2009 to July 2019, 71 patients were consecutively treated with AZA ± DLI for relapse of AML or high-risk MDS/MPN after HSCT from alternative donors. Alternative donors included MMUD (unrelated donor with maximum 1 allelic/antigenic mismatch at either HLA-A, -B, -C, -DRB1 or -DQB1 locus), family haploidentical (sibling or other relative sharing only one HLA-haplotype with recipient) and unrelated umbilical cord blood (CBU). Patients were treated at three different European centers: San Raffaele Scientific Institute in Milan (Italy), University Hospital Carl Gustav Carus in Dresden (Germany), and University Hospital in Duesseldorf (Germany).

According to institutional policies, the indication for treatment with AZA was either hematologic or molecular relapse post-HSCT not eligible to

	-					, ,					-		_							
Study	Type of study	Type of relapse	AZA Schedule	Pts (<i>n</i>)	DLI 0	Median age	MAC [%]	MSD MUD (%)	им (<i>n</i>)	Haplo (<i>n</i>)	Time to relapse (days)	ORR ([%][[S. (%)	Duration of CR days)	Median Survival (months)	2-year 0S	Acute GVH D [%]	Chronic GVHD [%]	Neutropenia grade III–IV (%)	Thrombo- cytopenia grade III-IV (%)
Graef et al.°	Retr.	Morph	100 mg/ m ² ×5dd	~	-	94	0	100	0	0	50	100	001	210	7		ou	ou		
Jabbour et al. ¹⁰	Retr	Morph	$16-40 \text{ mg/}$ $\text{m}^2 \times 5 \text{dd}$	6	0	48	78	78	0	0	240	22	33	120	6		11	ou		
Kim et al. ¹⁹	Retr	Morph	$75 \mathrm{mg/}$ m $^2 imes 7 \mathrm{dd}$	4		36		100	0	0	591	75 5	30	595	13		0	75	100	100
Lübbert et al. ²¹	^{:0} Retr	Morph	$100\mathrm{mg} imes3dd$	26	19	62	21	100	0	0	248	56	9	525	4.5	16	11	4		
Czibere et al. ²¹	¹ Retr	Morph	100 mg/ m ² × 5dd	22	18	50	91	100	0	0	103	72 2	23 4	433	5	23	33	18		
Bolaños- Meade et al. ²²	Retr	Morph	$40-75 \text{ mg/}$ m ² \times 7 dd	10	с	55	90	80	0	7	480	70	20	524	14		0	10		
Platzbecker et al. ²³	Prosp	Molec	$75 \mathrm{mg/}$ m ² $ imes$ 7 dd	20	0	58	10	75	e	2	169	80	20	231	12		0	0	80	65
Schroeder et al. ²⁴	Prosp	Morph	100 mg/ m ² ×5dd	30	22	55	13	13	7	0	175	47	23	535	4	17	37	17	65	63
Tessoulin et al. ²⁵	Retr	Morph	$75 \mathrm{mg/}$ m ² $ imes$ 7 dd	31	7	57	9				111	~	14			2J				
Schroeder et al. ²⁶	Retr	Both	100 mg/m ² × 5dd-75 mg/ m ² × 7dd	154	105	55	42	78	31	5	185	33	27			29	23	27		
		Morph		135									21			19				
		Molec		19									72			69				
Steinmann et al. ¹¹	Retr	Morph	100 mg × 3dd- 100 mg/ m² × 5dd	72	56	62	17	100				ب	` 0	1174	3.5	54	10	4		
Drozd- Sokołowska et al. ¹²	Retr	Both	75 mg/ m² × 7dd	13	9	59					180									
		Morph		6	9	57	77					33 ((231	6.8	0	11	0	67	100
		Molec		4	0	61	50										25	0	100	50
Craddock et al. ¹³	Retr	Morph	75 mg/ m²×5-7dd	181	69		25		0	4	240	29	15			12	7			
Ishikawa et al. ¹⁴	Retr	Morph	$75 \mathrm{mg/}$ m $^2 imes 7 \mathrm{dd}$	-	-	56	0	100	0	0	98	100	7 001	450			100	100		
																				(Continued)

L Carmine, LS Maria Teresa *et al.*

Therapeutic Advances in Hematology 13

Table 1. (Cor	ntinued	Ŧ																		
Study	Type of study	Type of relapse	AZA Schedule	Pts (n)	ы Г	Median age	MAC (%)	MSD MUD (%)	ими (<i>n</i>)	Haplo (<i>n</i>)	Time to relapse (days)	ORR (%)(CR [%]	Duration of CR (days)	Median Survival (months)	2-year 0S	Acute GVH D (%)	Chronic GVHD [%]	Neutropenia grade III-IV [%]	Thrombo- cytopenia grade III-IV (%)
Woo et al. ¹⁵	Prosp	Both	75 mg/ m² × 7 dd	39	-	52	30	100	0	0	< 100	49	œ			25	ω			
	Morph			2																
	Molec			34																
Platzbecker et al. ¹⁶	Prosp	Molec	75 mg/ m ² × 7 dd	24	2	59					163	70	53			62			85	9
Claiborne et al. ¹⁷	Retr	Both	$75 \mathrm{mg}/\mathrm{m}^2 imes 7 \mathrm{dd}$	28	28	57	46	60	. 	-	188		36		10	35	11	36		
		Morph		20	20								25		9.5					
		Molec		8	80								63		not reached					
Rautenberg et al. ¹⁸	Retr	Both	100 mg/ m ² × 5dd– 75 mg/ m ² × 7dd	151	105	54	36	75	34	4	147	46	41	330			42	26		
		Morph		92													41	21		
		Molec		59													42	34		
Drozd- Sokołowska et al. ³¹	Retr.	Morph.	75 mg/ m² × 7dd– 100 mg/ m² × 5dd	23	11	56	43	82	5	5	105	26	13		5.9	ω	17	4		
Liberatore	Retr	Both	32 mg/ m ² × 5dd- 75 mg/ m ² × 7dd	71	33	56	52	0	39	29	270	49	38	510	7	41	27	18	83	54
		Morph		40	17		45		19	18	300	38	15	540	9	19	20	œ	88	68
		Molec		31	16		61		20	1	240	65	65	510	15	70	35	19	75	31
		Haplo		29	6		52				300	55	31	510	7	31	14	17	83	59
		MMUD		39	24		51				240	44	44	579	8	48	38	21	92	54
AZA, azacitid regimen; MM rate; OS, over	ine; CR, IUD, mis rall surv	, complete smatched <i>v</i> ival; Prop	e response; DL unrelated don ıs, prospective;	l, dono or; Mo ; Pts, p	ir lymp lec, mo latients	hocytes i olecular; s; Retr, re	Morph trospe	ns; Gvh 1, morp ective.	HD, graft hologice	t-versus al; MSD,	s-host di matche	sease ed sibl	; Hapl ing do	lo, haploid nor; MUD	entical do , matched	nor; M/ unrela	vC, myel ted dong	oablative or; ORR, a	e conditioning overall respo	nse

clinical trials and further IC because of general condition and previous refractoriness. AZA was given as first salvage treatment; patients receiving additional anti-leukemic agents other than DLI and those who received maintenance therapy prior to relapse were excluded. Hematologic relapse was defined as either bone marrow (BM) blast count \geq 5%, appearance of blasts in peripheral blood (PB) or extramedullary relapse. Molecular relapse was defined as recurrence or progressive increase in disease markers after HSCT, without evidence of hematologic relapse. Methods for the detection of measurable residual disease (MRD) were multiparameter flow cytometry (MFC), real-time quantitative polymerase chain reaction (RT-qPCR) for known mutations (e.g. NPM1 gene), cytogenetical, and fluorescent in situ hybridization (FISH) analysis; in the absence of disease-specific markers, either a WT1 mRNA assay or a decrease in donor chimerism was adopted as a marker of impending relapse (see Supplemental data). The schedules of AZA were either 32 mg/m^2 days 1–5 every 28 days or 75 mg/m² days 1-7 every 28 days. DLI were generally given from the second cycle of AZA, every two cycles and at escalating dose starting from 1×10^5 CD3 +/Kg in haplo and from 1×10^6 CD3 + /Kg in MMUD, as per institutional guidelines. Both AZA dosage and indication to DLI relied on clinical-biological characteristics of patients, features of disease, type of HSCT, and availability of the donor. All patients gave informed consent for treatment with AZA \pm DLI. Patients were treated according to current institutional programs on written informed consent for transplantation procedures, biological sampling and use of medical records for research. Patients' date were de-identified and anonymized. The reporting of this study conforms to the STROBE statement.51

Response criteria

CR to treatment was defined as BM blasts < 5%in the absence of both circulating blasts and extramedullary disease, but hematological reconstitution was not required for CR, as factors other than underlying disease (e.g. viral infections, GvHD, or drugs) could contribute to cytopenia in the post-transplantation period. For patients in molecular relapse, disappearance of disease markers was defined as CR MRD-negative, whereas persistence was defined as CR MRD-positive. Partial response (PR) was defined as either decrease in BM blast to 5–25% or decrease in pretreatment BM blast by at least 50%. Progressive disease (PD) was defined as either increase in BM/PB blast percentage or new extramedullary disease during treatment, whereas stable disease (SD) as absence of criteria for CR, PR, and SD.⁵²

Time to response was calculated from the first day of AZA until best response. OS was defined as the time from the first day of AZA until death by any cause or last follow-up. Patients who received a subsequent HSCT were censored at that date. Event-free survival (EFS) was defined as an interval from AZA administration to either relapse or progression or death in remission (whichever came first). The intensity of conditioning regimens was defined according to Bacigalupo et al.53 HCT-comorbidity index (HCT-CI) and disease risk index (DRI) were defined according to Sorror et al.54 and Armand et al.,55 respectively. Acute and chronic GvHD were classified according to criteria by Harris et al. and NIH 2014 consensus conference, respectively.56,57 Adverse events (AE) were graded according to CTC-AE v5.0.

Statistics

Continuous variables were summarized using median (range), whereas frequency tables were used for categorical variables. For univariate comparison, cross-tabulation, Fisher's exact test, and Mann-Whitney test were used. All outcomes were measured from the time of AZA administration. Treatment-related mortality (TRM) was defined as death from any cause while in continuous remission of the primary disease.58,59 Cumulative incidences were estimated for relapse and TRM and to accommodate competing risks. Relapse or progression was a competing risk for TRM. The probabilities of OS and EFS were estimated using the Kaplan-Meyer estimator. Log-rank test was used for univariate comparisons of survival curves, while the Gray's test was conducted to compare cumulative incidences of competing risks endpoints.^{60,61} Impact of DLI as time-dependent covariate was tested using the Simon-Makuch plot and Mantel-Byar test. Statistical analyses were performed with R 4.0.4 (R Development Core Team, Vienna, Austria) software.

Results

Patients and treatment

We retrospectively analyzed 71 patients (median age, 56 years) diagnosed with AML (n=52, 73%) and MDS/MPN (n=19, 27%) who relapsed after HSCT from alternative donors: 29 patients (41%) had received haploidentical grafts and 39 patients (55%) received grafts from MMUD; furthermore, transplantation from single CBU was performed in three patients (4%). The majority of the patients had high-risk and advanced-stage disease at transplantation: DRI was high in 33 patients (46%) and very high in 11 patients (16%); 28 patients (40%) had active disease at transplantation. A myeloablative conditioning regimen was adopted in 37 patients (52%), whereas GvHD prophylaxis mainly relied on ATG-based (40 patients, 56%) and post-transplantation cyclophosphamide (PTCy)-based platforms (24 patients, 34%). Details on patients, disease, and transplantation characteristics are shown in Table S1.

Overall, 31 patients (44%) presented with molecular relapse, whereas 40 patients (56%) had an overt hematologic relapse, including 3 patients with persistence of disease after HSCT (Table 2). The median time from HSCT to relapse (TTR) in the entire cohort was 9 months (range 1-112 months). Among molecular relapse, TTR was 8 months (range 1-55 months). Methods of MRD detection were RT-qPCR in 17 patients (54%), MFC in 3 patients (10%), cytogenetical analysis in 4 patients (13%), and chimerism in unsorted BM mononuclear cells and sorted CD34 + PB cells in 4 (13%) and 3 patients (10%), respectively. Among hematologic relapses, TTR was 10 months (range 1-112 months) and median blast percentage in BM was 13% (range 5-77%). None of them presented with isolated extramedullary relapse; one patient had multiple localizations of leukemia cutis concurrent to relapse in BM. Clinical characteristics of patients at HSCT were well balanced between the types of relapse (Table S2). Compared with MMUD, a greater number of patients diagnosed with AML (p=0.012) with higher HCT-CI (p=0.047) and heavily pre-treated before HSCT more (p=0.0007), and a more common use of BM as stem cell source (p = 0.0002) and PTCy as GvHD prophylaxis (p < 0.0001) were reported among haploidentical transplantation (Table S3).

Before relapse, 30 patients (42%) were diagnosed with all-grade and 7 patients (10%) with grade III–IV acute GvHD. In the same time-period, 16 patients (23%) had all-grade and 9 patients (13%) had moderate/severe chronic GvHD. At relapse, 27 patients (38%) were still on immunosuppressive therapy (IST): 26 patients successfully discontinued IST during first AZA cycle, while 1 patient presented a reflare of acute GvHD (Table 2).

The median time between relapse and first administration of AZA was 18 days (range 0–73 days). The majority of patients (n=58, n=58)82%) received AZA 75 mg/m^2 . Any difference emerged in AZA dosage according to the type of relapse, while more patients after haplo transplantation received 32 mg/m² schedule than MMUD (n=8, 28% versus n=0, respectively;p = 0.0008). Overall, patients received a median of four AZA cycles (range 1-37 cycles), without differences among subgroups (Table 3). However, 39 patients (46%) received DLI in combination with AZA. DLI were given more commonly among MMUD than haplo (n=24, 61% versus)n=9,31%, respectively; p=0.016). Median number of DLI per patient was 2 (range 1-4) and median cumulative amount of CD3 + cells/kg per patient was 2.1×10^6 (range: $0.1 - 62.5 \times 10^6$) (Table 3). Reasons for not giving DLI in 38 patients were PD (n=17), recent or active GvHD (n=9), donor unavailability (n=4), achievement of CR with AZA alone (n=1), and leukemia relapse with HLA-loss (n=4).

Response and outcome

Overall, 27 patients (38%) achieved CR and 8 patients (11%) achieved PR, accounting for an ORR of 49%. Higher ORR (n=20, 65%) and CR (n=20, 65%) were observed in molecular relapses compared to hematologic relapses (n=15, 38%) and n=7, 15%, respectively; p=0.03 and p = 0.0001, respectively). Even considering MRD-negative CR, response rate remained higher in molecular than hematologic relapse (n=14, 45% versus n=1, 3%; p < 0.001). Any difference in ORR and CR emerged between the types of donor, although MRD-negative CR was higher in MMUD (n=10, 26%) compared to haplo (n=5, 17%; p=0.004). Median time to CR was 109 days (range 51–300 days), corresponding to four AZA cycles (range 1-8). Median duration of CR was 17 months (range 5–89 months). In addition, 34 patients (48%) had PD (Table 4).

After a median follow-up of 11 months (range 1–115 months), median EFS in the entire cohort was 6 months (range 1–90 months) and EFS at 1 year was 26% (CI 95%: 16–36%). Median OS was 7 months (range 1–90 months), with an OS at 1 year of 53% (CI 95%: 40–64%) (Figure 1(a)). Patients with molecular relapse showed higher 1-year EFS (43% *versus* 13%, p=0.006) and 1-year OS (79% *versus* 34%; p<0.001) than hematologic relapses (Figure 1(b)). No significant difference of survival emerged between haplo and MMUD. At 1-year, cumulative incidence of relapse was 73% (CI 95%: 61–82%), whereas treatment-related mortality (TRM) was 1% (CI 95%: 1–7%) (Table 4).

However, 17 out of 33 patients receiving DLI achieved CR (51%), compared to 10 out of 30 patients receiving AZA single-agent (26%; p=0.03) (Table S4). To assess the potential benefit of DLI, we plotted Simon–Makuch plots for OS and EFS of patients with or without DLI. Overall, addition of DLI correlated with both longer 1-year EFS (42% versus 17%; Mantel–Byar test, p=0.004) and 1-year OS (73% versus 31%; Mantel–Byar test, p=0.002) than AZA single-agent (Figure 2).

At data cut-off, 31 patients were alive and 13 patients who achieved CR maintained response without further treatment: 10 patients with molecular and 3 patients with hematologic relapse; 4 patients in haplo and 9 patients in MMUD group. Overall, 34 patients received further treatments at PD, including 11 patients who received a subsequent HSCT. Of those, 3 patients (27%) deceased, whereas 8 patients (67%) are alive and free of disease after a median of 15 months (range 0–100 months). At last follow-up, 40 patients had succumbed from underlying disease (n=31), infections (n=4), GvHD (1), second neoplasia (n=1), and complications of subsequent HSCT (n=1).

Predictors for response and OS

Univariate analysis identified molecular relapse, BM blasts at relapse < 13% (median at relapse) and the addition of DLI as predictors for CR. The same factors appeared as predictive for both EFS and OS at 1 year. In univariate analysis, time

Table 2.	Clinical and biological characteristics of patients at relapse.

Overall population	<i>N</i> = 71
Time from HSCT to relapse, months (range)	9 (1–112)
Type of relapse	
Hematologic	40 (56%)
Molecular	31 (44%)
Median BM blasts among hematologic relapse, % (range)	13% (5–77)
Method of MRD detection among patients with molecu	lar relapse
RT-qPCR	17 (54%)
• NPM1	8/17
• WT1	5/17
• Other	4/17
MFC	3 (10%)
Cytogenetical analysis/FISH	4 (13%)
Chimerism on unsorted mononuclear PB cells	4 (13%)
Chimerism on CD34 + sorted PB cells	3 (10%)
More than one method	5 (16%)
No. of patients with acute GvHD before relapse	30 (42%)
Grade III-IV	7 (10%)
No. of patients with chronic GvHD before relapse	16 (23%)
Moderate/severe	9 (13%)
No. of patients on IST at relapse	27 (38%)
Taper/stop	26
Reflare	1

FISH, fluorescent *in situ* hybridization; GvHD, graft-versus-host disease; HSCT, allogeneic hematopoietic stem cell transplantation; IST, immunosuppressive therapy; MRD, measurable residual disease; PB, peripheral blood; RT-qPCR, real-time quantitative polymerase chain reaction.

between HSCT and relapse < 6 months also predicted 1-year OS (Table 5).

GvHD and AE

Incidence of all-grade and grade III–IV acute GvHD was 27% (n=19) and 13% (n=9), respectively. Acute GvHD occurred after a median of two AZA cycles (range 1–8). Indeed, MMUD

	Overall population (<i>n</i> = 71)	Molecular relapse (n=31)	Morphological relapse (<i>n</i> = 40)	Haploidentical donor (<i>n</i> = 29)	MMUD (<i>n</i> = 39)
Patient age at relapse, years, median (range)	57 (20–76)	57 (26–71)	58 (20–76)	57 (20–72)	58 (32–76)
Time from HSCT to AZA, months, median (range)	10 (1–113)	9 (2–56)	10 (1–112)	10 (1–56)	10 (2–113)
Time from relapse to AZA, days, median (range)	18 (0–73)	23 (0–73)	11 (0–58)	12 (0–57)	21 (0-73)
AZA dosage					
$75\mathrm{mg/m^2}\! imes\!7\mathrm{days}$ q28d	58 (82%)	23 (74%)	35 (88%)	19 (65%)	36 (92%)
$32\mathrm{mg/m^2}\! imes\!5\mathrm{days}\mathrm{q28d}$	8 (11%)	6 (19%)	2 (5%)	8 (28%)	0 (0%)
		<i>p</i> =0.21		<i>p</i> = 0.0008	
Other	5 (7%)	2 (7%)	3 (7%)	2 (7%)	3 (8%)
AZA cycles, median (range)	4 (1–37)	6 (1–15)	4 (1–37)	4 (1–28)	5 (1–37)
		<i>p</i> =0.33		<i>p</i> =0.66	
Patients with additional DLI	33 (46%)	16 (52%)	17 (43%)	9 (31%)	24 (61%)
		<i>p</i> =0.48		<i>p</i> =0.016	
Median DLI per patient	2 (1–4)	2 [1-4]	2 (1-4)	2 (1–3)	2 (1-4)
Median cumulative amount of CD3 + /kg per patient	2.1×10 ⁶ (0.1–62.5)	1.65×10 ⁶ (0.1–62.5)	5×10 ⁶ (0.2–51)	1×10 ⁶ (0.1–16)	3.55×10 ⁶ (0.5-62.5)
		<i>p</i> =0.91		<i>p</i> =0.13	
Time from AZA to first DLI, days (range)	50 (7–463)	50 (35–303)	50 (7–463)	44 (34–247)	50 (7–463)
		<i>p</i> =0.56		<i>p</i> =0.66	

Table 3. Characteristics of salvage treatment in overall population and subgroups.

AZA, azacitidine; DLI, donor's lymphocytes infusions; HSCT, allogeneic hematopoietic stem cell transplantation; MMUD, mismatched unrelated donor. "p" statistically significant was highlighted in bold.

> suffered more from all-grade acute GvHD (n=15, 38%) compared to haplo (n=4, 14%; p=0.031). However, 10 out of 19 patients developing acute GvHD had received DLI (Table 6). The overall incidence of chronic GvHD was 18% (n=13), occurring after a median of 3.5 AZA cycles (range 1–8). Eight patients (11%) had moderate/severe chronic GvHD. No significant difference in incidence emerged among subgroups. Therefore, 5 out of 13 patients developing chronic GVHD had received AZA + DLI (Table 6).

Among 41 patients evaluable, most common grade III-IV AE during salvage treatment were

hematological: neutropenia (83%), thrombocytopenia (54%), and anemia (42%). Nonhematological grade III–IV AE were mainly infections: febrile neutropenia (32%), pneumonia (10%), invasive fungal infections (6%), and sepsis (2%).

Validation of the AZA prognostic scoring system for relapse after transplantation

In the attempt to stratify patients eligible to AZAbased salvage treatments and to predict response and survival, Rautenberg and colleagues recently developed a new prognostic tool: the AZA



Figure 1. OS and EFS after treatment with AZA \pm DLI in the entire cohort and according to the type of relapse. Figure 1(a) shows OS and EFS in the entire cohort. OS was 53% at 1 year (40–64%; CI 95%) and EFS was 26% at 1 year (16–36%; CI 95%). Figure 1(b) shows OS and EFS according to the type of relapse. In patients with molecular relapse (black curve), OS was 79% at 1 year (58–90%; CI 95%), whereas in patients with hematologic relapse (red curve), OS was 34% at 1 year (19–49%; CI 95%). In patients with molecular relapse (black curve), EFS was 43% at 1 year (26–60%; CI 95%), whereas in patients with hematologic relapse (red curve), EFS was 13% at 1 year (5–25%; CI 95%).

Prognostic Scoring System for Relapse after transplantation (APSS-R).¹⁸ In this score, 1 point was assigned for molecular relapse and 2 points for hematologic relapse; for an interval between HSCT and relapse ≥ 6 months, 0 points were assigned, while 1 point was given for an interval < 6 months. APSS-R clearly stratifies patients into three subgroups (favorable=1 point, intermediate=2 points, unfavorable=3 points), with CR rates of 71%, 39%, and 29%, respectively,

Therapeutic Advances in Hematology 13

Table 4.	Response to	treatment	and c	outcomes in	overall	pop	ulation	and	subgrou	ps.

	Overall population (n=71)	Molecular relapse (n=31)	Morphological relapse (n=40)	Haploidentical donor (<i>n</i> = 29)	MMUD (<i>n</i> =39)
Disease response to AZA					
ORR (CR + PR)	35 (49%)	20 (65%)	15 (38%)	16 (55%)	17 (44%)
		p=0.03		p=0.46	
CR	27 (38%)	20 (65%)	7 (15%)	9 (31%)	17 (44%)
		p < 0.001		p=0.32	
CR MRD-negative	15 (21%)	14 (45%)	1 (3%)	5 (17%)	10 (26%)
		p<0.001		p=0.004	
PR	8 (11%)	0 (0%)	8 (20%)	7 (24%)	0 (0%)
SD	2 (3%)	1 (3%)	1 (2%)	2 (7%)	0 (0%)
PD	34 (48%)	10 (32%)	24 (60%)	11 (38%)	22 (56%)
Median time to CR, days (range)	109 (51–300)	110.5 (62–300)	109 (51–274)	103 (65–196)	125 (51–300)
Median AZA cycles to CR (range)	4 (1–8)	4 (1–8)	3 (2–7)	4 (2–7)	4 [1-8]
• Pts already in CR before DLI	13/27	9/20	4/7	8/9	5/17
Median duration of CR, months	17 (5–89)	17 (5–89)	18 (7–90)	17 (5–64)	19 (5–90)
		p=0.85		p=0.48	
Median EFS, months	6 (1 – 90)	7 (1–90)	4 (1–90)	5 (1–64)	6 (1-90)
Median OS, months	7 (1-90)	15 (2–90)	6 (1-90)	7 (1–64)	8 (1-90)
Median follow-up, months	11 (1–115)	23 (2–115)	7 (1-90)	12 (1–115)	10 (1–90)
		p=0.98		p=0.16	
1-year OS (95% CI)	53% (40-64)	79% (58–90)	34% (19–49)	53% (33–70)	55% (37–70)
		p < 0.001		p=0.60	
2-year OS (95% CI)	41% (29–53)	70% (49–84)	19% (8–33)	31% (14–50)	48% (31–64)
		p < 0.001		p=0.60	
1-year EFS (95% CI)	26% (16–36)	43% (26–60)	13% (5–25)	21% (8–37)	32% (18–47)
		p=0.006		p = 0.39	
2-year EFS (95% CI)	18% (10–28)	32% (16–49)	8% (2–18)	13% (4–28)	23% (11–38)
		p=0.006		p=0.39	
1-year TRM (95% CI)	1% (1–7)	0%	3% [1–12]	3% (0–16)	0%
		p=0.35		p=0.94	
1-year relapse incidence (95% CI)	73% (61–82)	57% (36–72)	85% (69–93)	76% (55–88)	68% (50-81)
		p=0.008		p = 0.46	

AZA, azacitidine; CR, complete response; DLI, donor lymphocytes infusions; EFS, event-free survival; MMUD, mismatched unrelated donor; MRD, measurable residual disease; ORR, overall response rate; OS, overall survival; PD, progressive disease; PR, partial response; SD, stable disease; TRM, treatment-related mortality. "p" statistically significant was highlighted in bold.



Figure 2. Simon–Makuch plots of OS and EFS in patients receiving AZA + DLI compared to AZA. Figure 2 shows Simon–Makuch plots of OS and EFS in patients receiving AZA + DLI compared to AZA. On the left, patients receiving AZA + DLI (red curve) presented longer 1-year OS than those (black curve) receiving AZA single-agent (73% *versus* 31%, respectively; Mantel–Byar test, p = 0.002). On the right, patients receiving AZA + DLI (red curve) presented longer 1-year EFS than those (black curve) receiving AZA single-agent (42% *versus* 17%, respectively; Mantel–Byar test, p = 0.004).

and 2-year OS rates of 64%, 38%, and 27%, respectively.¹⁸ When applied to our cohort, we observed CR rates of 76%, 31%, and 13%, whereas OS was 94%, 46%, and 25% at 1-year, respectively. The model confirmed its ability to stratify patients into distinct prognostic groups with significantly different response rates (p=0.0005) and survival (p<0.0001) (Table S5) (Figure 3).

Discussion

This study represents, to the best of our knowledge, the first report analyzing in details AZA \pm DLI as salvage treatments for AML and MDS/MPN relapsed after HSCT from alternative donors.

When using treatments able to modulate graft alloreactivity, as both AZA and DLI, HSCT from alternative donors represents a particular condition, because of the potentially increased risk of life-threatening GvHD due to donor-recipient HLA disparity. Moreover, HSCT from alternative donors are often performed in advancedstage disease and burdened by greater toxicities, thus representing a matter of concern for subsequent salvage treatments. Indeed, this is the case of our cohort, characterized by heavily pre-treated patients, high-risk disease, and early recurrence post-HSCT. Nonetheless, administration of AZA 75 mg/m^2 proved feasible and tolerance. As expected in the early post-transplantation period, most frequent AE were hematological toxicities and infectious complications, directly related to severe neutropenia induced by treatment. Anyway, TRM remained low.

In our series, we observed an ORR of 49% and CR rate of 38%, EFS at 1- and 2-year of 26% and 18%, respectively, and OS at 1- and 2-year of 53% and 41%, respectively. These data compared favorably with studies on 'standard' salvage treatments, reporting CR rate of about 30% and 2-year OS between 10% and 20%.⁵⁻⁷ Our results also resembled those available in literature on post-transplantation AZA \pm DLI, characterized by CR rates of 15–67% and a 2-year OS of 12–69% (Table 1).⁹⁻³¹

The burden of leukemia at relapse appears to profoundly influence both response and outcomes. Two retrospective studies found that pre-emptive intervention at MRD reoccurrence leads to higher response and longer survival than treatment at hematologic relapse.^{18,26} Furthermore, the prospective phase II trial RELAZA-2 reported 53% of CR MRD-negative and a 2-year OS of 62%

Table 5	Univariate anal	veic of rick	factors for r	hac against	outcomes
Table 5.	Univariate anat	YSIS ULLISK	Idcluisiui I	esponse anu	outcomes.

	CR	1-year EFS (95% CI)	1-year OS (95% CI)
Type of relapse			
Molecular $(n=31)$	20	44% (26–60)	79% (58–90)
Morphological $(n = 40)$	7	13% (5–25)	34% (19–49)
<i>p</i> -value	0.0001	0.006	<0.0001
Type of donor			
Haplo (<i>n</i> =29)	9	21% (8–37)	53% (33–70)
MMUD (n=39)	17	32% (18–47)	55% (37–70)
<i>p</i> -value	0.32	0.39	0.60
Blast at relapse ^a			
<13% (<i>n</i> =48)	23	36% (23–50)	66% (50–78)
≥13% (<i>n</i> =22)	4	5% (0–19)	25% (8–46)
<i>p</i> -value	0.02	0.0003	0.002
Blast at relapse			
<20% (<i>n</i> =57)	26	32% (20-44)	65% (50–77)
≥20% (<i>n</i> = 13)	1	0	0%
<i>p</i> -value	0.01	<0.0001	<0.0001
Median time from HSCT to relapse			
<9 months (<i>n</i> = 34)	11	21% (9–36)	43% (25–59)
\geq 9 months (<i>n</i> =37)	16	30% (16–45)	62% (44–76)
<i>p</i> -value	0.34	0.44	0.06
Median time from HSCT to relapse			
<6 months (<i>n</i> = 28)	8	18% (6–35)	36% (18–55)
\geq 6 months (<i>n</i> = 43)	19	30% (17–44)	63% (46–76)
<i>p</i> -value	0.21	0.31	0.05
DLI			
Yes (<i>n</i> =33)	17	41% (24–57)	72% (53–84)
No (n=38)	10	12% (4–25)	34% (19–51)
<i>p</i> -value	0.03	0.004	0.009

CR, complete response; DLI, donor lymphocytes infusions; EFS, event-free survival; HSCT, allogeneic hematopoietic stem cell transplantation; MMUD, mismatched unrelated donor; OS, overall survival. ^aThreshold of 13% was chosen as it represents the median BM blast percentage at morphological relapse.

 Table 6.
 Incidence of acute and chronic GvHD in overall population and subgroups.

	Overall population (<i>n</i> = 71)	Molecular relapse (n=31)	Morphological relapse (n=40)	Haploidentical donor (<i>n</i> = 29)	MMUD (<i>n</i> = 39)
Patients with all-grade aGvHD during AZA, <i>n</i> (%)	19 (27%)	11 (35%)	8 (20%)	4 (14%)	15 (38%)
		<i>p</i> =0.18		<i>p</i> =0.031	
Patients with grade III–IV aGvHD during AZA, <i>n</i> (%)	9 (13%)	6 (19%)	3 (8%)	1 (4%)	7 (18%)
AZA cycle of aGvHD onset, median	2 (1–8)	2 (1–8)	2.5 (1–8)	1.5 (1–8)	4 (1–8)
Patients who received DLI before aGvHD onset, <i>n</i> (%)	10/19 (53%)	6/11 (55%)	4/8 (50%)	1/4 (25%)	9/15 (60%)
CR of aGvHD to steroids, <i>n</i> (%)	12/19 (63%)	7/11 (64%)	6/8 (75%)	3/4 (75%)	9/15 (60%)
Patients with previous signs of acute/ chronic GvHD who delevoped aGvHD during AZA, n (%)	6/34 (18%)				
Patients with cGvHD during AZA, n (%)	13 (18%)	8 (26%)	5 (13%)	5 (17%)	8 (21%)
		<i>p</i> =0.22		<i>p</i> = 1	
Patients with NIH moderate-severe cGvHD during AZA, <i>n</i> (%)	8 (11%)	6 (19%)	2 (5%)	4 (13%)	4 (10%)
AZA cycle of cGvHD onset, median	3.5 (1–8)	3 (1–8)	3.5 (1–7)	2.5 (1–4)	5 (1–8)
Patients who received DLI before cGvHD onset, <i>n</i> (%)	5/13 (38%)	3/8 (38%)	2/5 (40%)	0	5/8 (63%)
Patients with previous signs of acute/ chronic GvHD who developed cGvHD during AZA \pm DLI, n (%)	9/34 (26%)				

aGvHD, acute graft-versus-host disease; AZA, azacitidine; DLI, donor lymphocytes infusions; cGvHD, chronic graft-versus-host disease; MMUD, mismatched unrelated donors.

"p" statistically significant was highlighted in bold.

among patients relapsed post-HSCT, comparably higher than historical controls.¹⁶ In our cohort, univariate analysis identified patients treated at molecular relapse as having significantly higher response rate, EFS, and OS than hematologic relapses. No difference emerged in the number of AZA cycle administered. Pre-emptive treatment with AZA \pm DLI led to durable remissions in a considerable proportion of patients, as reflected by the 2-year EFS of 32%. As the duration of CR was particularly long-lasting (median 17 months), even for those patients experiencing PD, preemptive therapy might represent a valid intervention, able to delay overt hematologic relapse and to postpone the need for more intensive treatments, allowing recovery from transplant-related toxicities. Therefore, despite prospective, randomized trials are warranted, our results reinforce the value of close post-transplantation MRD monitoring and rapid pre-emptive intervention in case of MRD positivity. On the contrary, $AZA \pm DLI$ showed modest efficacy in case of hematologic relapse, probably due to the high proliferative rate of the disease and the insufficient anti-leukemic effect. Nonetheless, trying to identify patients who could benefit from AZA, univariate analysis identified burden of disease in BM as predictive of both response and outcomes. Whereas in the setting of hematologic relapse, IC could still play a role in disease control, in patients with targetable molecular lesions in the combination of either IC or HMA with novel target drugs,



Figure 3. OS in the entire cohort according to APSS-R. Figure 3 shows OS in the entire cohort according to APSS-R. OS at 1 year was 94% (63–99%; Cl 95%) in favorable-risk patients (1 point), 46% (28–62%; Cl 95%) in intermediate-risk patients (2 points), and 25% (8–57%; Cl 95%) in high-risk patients (3 points) (p < 0.0001).

such as FLT-3 and IDH1-2 inhibitors, showed promising results.^{62–67} Moreover, in those patients without targetable mutations, recent results from the combinations of HMA with BCL-2 inhibitor Venetoclax appeared promising.^{68–70} HMA are also employed as maintenance strategies after HSCT in high-risk patients. Beside the risk of overtreatment, conflicting results emerged from initial studies.^{32,71,72} Nonetheless, post-HSCT maintenance could still be an option for high-risk patients without disease-specific markers and drug-targetable molecular features. Following encouraging results of oral AZA as maintenance in AML,⁷³ results are awaited from ongoing trial in the setting of HSCT.^{74,75}

Focusing on type of donor, $AZA \pm DLI$ appeared equally effective in both MMUD and haploidentical setting. The lower rate of MRD-negative CR observed among haplo might be related to a selection bias of more aggressive and heavily pre-treated diseases in our cohort. Considering the burden of previous toxicities, haplo also more often received AZA 32 mg/m².

In previous studies, the most frequently reported predictive factors of response and survival were molecular-only relapse, burden of disease at relapse, and TTR.^{13,17,18,26} In our cohort, we confirmed these factors, together with the combination of DLI to AZA. Notably, molecular-only relapse emerged as a strong predictor of both response and outcomes in univariate analysis. Many authors tried to develop prognostic scores that are able to identify patients who more capable to benefit from AZA \pm DLI. A first attempt was made by Craddock et al. with the 'AZA Relapse Prognostic Score' (ARPS), considering burden of disease in BM and TTR.13 Despite its validity, the ARPS was tested only in patients with hematologic relapse. This appears as a possible limit, considering both increasing evidence and real-life practice favoring MRD-driven pre-emptive interventions. Recently, Rautenberg et al. proposed a novel scoring system, the APSS-R, developed from a single-center retrospective analysis and based on type of relapse and TTR.18 When applied to our cohort, the APSS-R was able to clearly stratify patients even in the setting of alternative donors. Therefore, APSS-R might become a useful tool to predict outcomes in all patients receiving AZA post-transplantation.

The effective role of DLI is still a matter of debate. As DLI appeared necessary in consolidating responses to IC in post-transplantation relapses,^{5,6,8} their contribution when given together with AZA is less clear. A large retrospective study by Schroeder et al. initially reported on a possible benefit.²⁶ Conversely, Craddock et al.¹³ found no impact of DLI both on response and survival. Notably, this study included only hematologic relapses and the majority of patients received DLI in the absence of a clinical response. A prospective study on pre-emptive AZA without DLI also showed consistent but not durable responses.²³ Recently, Rautenberg et al.¹⁸ reported higher response rates and significant survival advantage in patients receiving DLI for both molecular and hematologic relapses. In our experience, we confirmed that the addition of DLI to AZA significantly improved response rates and correlated with longer EFS and OS compared to AZA single-agent.

In our population, the incidence of both acute and chronic GvHD before relapse was consistent with historical data on alternative donors.^{50,76–78} Despite HLA mismatches, during and shortly after treatment with AZA \pm DLI, we observed an incidence of all-grade acute and chronic GvHD consistent with previous studies (Table 1).9,10-26 Besides the limit of small numbers, a previous diagnosis of GvHD and administration of DLI did not impact on the incidence of GvHD during salvage treatment. The incidence of GvHD in our study compared favorably also with that reported by haploidentical DLI.79-81 Indeed, the different incidence of acute GvHD between the types of donors appeared to mainly rely on the relatively low incidence observed among haplo. Possible explanations could be a greater use of BM as HSC source and PTCy as GvHD prophylaxis in haplo. Notably, sirolimus was also adopted in combination with PTCy in many haploidentical HSCTs in our cohort, a platform of GvHD prophylaxis reported to favor the expansion of regulatory T cells.82,83

Limitations of this study include retrospective non-randomized nature, limited numbers only allowing univariate analysis, the possibly biasedselected population and heterogeneity of treatments. Multicentricity might limit the single-center effect in the management of patients.

Conclusion

AZA either single-agent or in combination with DLI proved feasible and effective in treating patients with AML and MDS/MPN relapsing after HSCT from alternative donors. The treatment was well tolerated, with an acceptable burden of toxicities and GvHD. In case of overt hematologic relapses, the salvage treatment with $AZA \pm DLI$ showed a modest efficacy. In this setting, combinational strategies of either IC or HMA with novel target drugs and Venetoclax might play a role. Conversely, AZA fared better in the setting of MRD. Pre-emptive treatment of molecular relapse granted higher response rate and longer outcomes, leading to durable remissions in a considerable proportion of patients. Even for those patients finally experiencing PD, pre-emptive therapy might represent a valid intervention, able to delay overt hematologic relapse, and to postpone the need for more intensive treatments. Our results reinforce the need of close post-transplantation MRD monitoring and the

value of early pre-emptive interventions. When applied to our cohort, APSS-R clearly stratified patients in distinct prognostic subgroups, thus appearing as a useful tool to predict outcomes even for alternative donors. Finally, we confirmed the benefits of DLI in combination with AZA, as their role appeared essential in obtaining better responses and ensuring longer survival.

Author contribution(s)

Carmine Liberatore: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – original draft; Writing – review & editing.

Maria Teresa Lupo-Stanghellini: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Francesca Lorentino: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Luca Vago: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Matteo Giovanni Carrabba: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Raffaella Greco: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Sarah Marktel: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Andrea Assanelli: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Francesca Farina: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Consuelo Corti: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Massimo Bernardi: Conceptualization; Data curation; Formal analysis; Methodology;

Supervision; Validation; Writing – review & editing.

Jacopo Peccatori: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Katja Sockel: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Jan Moritz Middeke: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Johannes Schetelig: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Anika Bergmann: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Christina Rautenberg: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Fabio Ciceri: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Martin Bornhäuser: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Thomas Schroeder: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Friedrich Stölzel: Conceptualization; Data curation; Formal analysis; Methodology; Supervision; Validation; Writing – review & editing.

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

Conflict of interest statement

The authors declared the following potential conflicts of interest with respect to the research,

authorship, and/or publication of this article: S.F. received honoraria from Jazz Pharmaceuticals, served on advisory boards for Medac GmbH and Novartis. B.M. received honoraria from Celgene/ BMS, Alexion, Novartis, MSD; travel grants from Celgene/MSD; served on advisory boards for BMS and Jazz Pharmaceuticals. C.M.G. declared consultancy for Janssen, Novartis, and Abbvie. S.T. declared research funding and consultancy for Celgene/BMS. R.C. received travel grants from Jazz Pharmaceuticals and Celgene/ BMS; lecturing from Abbvie, Celgene/BMS, and Jazz Pharmaceuticals. MTLS has received travel grants from Neovii Biotech; honoraria from Incyte; served on advisory boards for Novartis and Mallinckrodt Pharmaceuticals. L.C., L.F., V.L., G.R., M.S., A.A., F.F., C.C., B.M., P.J., S.K., M.J.M., S.J., B.A., and C.F. declare no competing financial interests.

Ethics approval, consent to participate

Six patients in Duesseldorf cohort were treated within the prospective phase Π trial (NCT00795548). Data have been reported previously²⁴ but were updated for this analysis. All patients gave informed consent for treatment with AZA \pm DLI. Patients were treated according to current institutional programs on written informed consent for transplantation procedures, biological sampling, and use of medical records for research (Ethical approvals: ALMON study approved by San Raffaele Institutional Ethical Committee on date 19/10/2007; Düsseldorf ethic Approval No. 4138/5309; TU Dresden Ethical Approval No. EK132042018). Patients' date was de-identified and anonymized.

ORCID iD

Fabio Ciceri D https://orcid.org/0000-0003-0873-0123

Availability of data and materials

All data produced in the present study are available upon reasonable request to the authors.

Supplemental material

Supplemental material for this article is available online.

References

- 1. Gooley TA, Chien JW, Pergam SA, et al.
 - Reduced mortality after allogeneic hematopoietic-

cell transplantation. New Engl J Med 2010; 363: 2091–2101.

- D'Souza A, Lee S, Zhu X, et al. Current use and trends in hematopoietic cell transplantation in the United States. *Biol Blood Marrow Transplant* 2017; 23: 1417–1421.
- Horowitz M, Schreiber H, Elder A, et al. Epidemiology and biology of relapse after stem cell transplantation. *Bone Marrow Transplantation* 2018; 53: 1379–1389.
- 4. Craddock C, Hoelzer D and Komanduri K. Current status and future clinical directions in the prevention and treatment of relapse following hematopoietic transplantation for acute myeloid and lymphoblastic leukemia. *Bone Marrow Transplant* 2019; 54(1): 6–16.
- Schmid C, Labopin M, Nagler A, et al. Treatment, risk factors, and outcome of adults with relapsed AML after reduced intensity conditioning for allogeneic stem cell transplantation. *Blood* 2012; 119: 1599–1606.
- 6. Bejanyan N, Weisdorf DJ, Logan BR, *et al.* Survival of patients with acute myeloid leukemia relapsing after allogeneic hematopoietic cell transplantation: a center for international blood and marrow transplant research study. *Biol Blood Marrow Transplant* 2015; 21(3): 454–459.
- Bazarbachi A, Schmid C, Labopin M, et al. Evaluation of trends and prognosis over time in patients with AML relapsing after allogeneic hematopoeitic cell transplant reveals improved survival for young patients in recent years. *Clin Cancer Res* 2020; 26: 6475–6482.
- Schmid C, Labopin M, Nagler A, et al. Donor lymphocyte infusion in the treatment of first hematological relapse after allogeneic stem-cell transplantation in adults with acute myeloid leukemia: a retrospective risk factors analysis and comparison with other strategies by the EBMT acute leukem. J Clin Oncol 2007; 25: 4938–4945.
- Graef T, Kuendgen A, Fenk R, et al. Successful treatment of relapsed AML after allogeneic stem cell transplantation with azacitidine. *Leuk Res* 2007; 31(2): 257–259.
- Jabbour E, Giralt S, Kantarjian H, et al. Lowdose azacitidine after allogeneic stem cell transplantation for acute leukemia. *Cancer* 2009; 115: 1899–1905.
- Steinmann J, Bertz H, Wäsch R, et al.
 5-Azacytidine and DLI can induce long-term remissions in AML patients relapsed after allograft. Bone Marrow Transplant 2015; 50(5): 690–695.

- Drozd-Sokołowska J, Gil L, Waszczuk-Gajda A, et al. Azacitidine use after allogeneic stem cell transplantation – results from the Polish adult leukemia group. *Transplant Proc* 2016; 48(5): 1802–1805.
- Craddock C, Labopin M, Robin M, et al. Clinical activity of azacitidine in patients who relapse after allogeneic stem cell transplantation for acute myeloid leukemia. *Haematologica* 2016; 101(7): 879–883.
- Ishikawa T, Fujii N, Imada M, et al. Graftversus-leukemia effect with a WT1-specific T-cell response induced by azacitidine and donor lymphocyte infusions after allogeneic hematopoietic stem cell transplantation. *Cytotherapy* 2017; 19(4): 514–520.
- Woo J, Deeg HJ, Storer B, *et al.* Factors determining responses to azacitidine in patients with myelodysplastic syndromes and acute myeloid leukemia with early post-transplantation relapse: a prospective trial. *Biol Blood Marrow Transplant* 2017; 23(1): 176–179.
- Platzbecker U, Middeke JM, Sockel K, et al. Measurable residual disease-guided treatment with azacitidine to prevent haematological relapse in patients with myelodysplastic syndrome and acute myeloid leukaemia (RELAZA2): an openlabel, multicentre, phase 2 trial. *Lancet Oncol* 2018; 19(12): 1668–1679.
- Claiborne J, Bandyopathyay D, Roberts C, et al. Managing post allograft relapse of myeloid neoplasms: azacitidine and donor lymphocyte infusions as salvage therapy. *Leuk Lymphoma* 2019; 60(11): 2733–2743.
- Rautenberg C, Bergmann A, Germing U, et al. Prediction of response and survival following treatment with azacitidine for relapse of acute myeloid leukemia and myelodysplastic syndromes after allogeneic hematopoietic stem cell transplantation. *Cancers* 2020; 12: 2255.
- Kim SY, Cho SG, Cho BS, et al. Azacytidine treatment after discontinuation of immunosuppressants in patients with myelodysplastic syndrome and relapse after allo-SCT at a single center. Bone Marrow Transplant 2010; 45(8): 1375–1376.
- Lübbert M, Bertz H, Wäsch R, et al. Efficacy of a 3-day, low-dose treatment with 5-azacytidine followed by donor lymphocyte infusions in older patients with acute myeloid leukemia or chronic myelomonocytic leukemia relapsed after allografting. Bone Marrow Transplant 2010; 45(4): 627–632.

- Czibere A, Bruns I, Kröger N, et al. 5-Azacytidine for the treatment of patients with acute myeloid leukemia or myelodysplastic syndrome who relapse after allo-SCT: a retrospective analysis. *Bone Marrow Transplantation* 2010; 45: 872–876.
- Bolaños-Meade J, Smith BD, Gore SD, et al.
 5-Azacytidine as Salvage treatment in relapsed myeloid tumors after allogeneic bone marrow transplantation. *Biol Blood Marrow Transplant* 2011; 17(5): 754–758.
- Platzbecker U, Wermke M, Radke J, et al. Azacitidine for treatment of imminent relapse in MDS or AML patients after allogeneic HSCT: results of the RELAZA trial. *Leukemia* 2012; 26(3): 381–389.
- Schroeder T, Czibere A, Platzbecker U, et al. Azacitidine and donor lymphocyte infusions as first salvage therapy for relapse of AML or MDS after allogeneic stem cell transplantation. Leukemia 2013; 1229–1235.
- Tessoulin B, Delaunay J, Chevallier P, et al. Azacitidine salvage therapy for relapse of myeloid malignancies following allogeneic hematopoietic SCT. Bone Marrow Transplant 2014; 49(4): 567–571.
- 26. Schroeder T, Rachlis E, Bug G, et al. Treatment of acute myeloid leukemia or myelodysplastic syndrome relapse after allogeneic stem cell transplantation with azacitidine and donor lymphocyte infusions – a retrospective multicenter analysis from the German cooperative transplant study group. *Biol Blood Marrow Transplant* 2015; 21: 653–660.
- Ganguly S, Amin M, Divine C, *et al.* Decitabine in patients with relapsed acute myeloid leukemia (AML) after allogeneic stem cell transplantation (allo-SCT). *Ann Hematol* 2013; 92(4): 549–550.
- Singh SN, Cao Q, Gojo I, et al. Durable complete remission after single agent decitabine in AML relapsing in extramedullary sites after allo-SCT. Bone Marrow Transplant 2012; 47(7): 1008–1009.
- Liu XL, Zhao X, Wang C, *et al.* Decitabine treatment for acute myeloid leukemia relapse after allogeneic hematopoietic stem cell transplantation. *J Biol Regul Homeost Agents* 2017; 31(1): 171–175.
- Wang B, Jin X, Wang Q, et al. Decitabine+ CAG +DLI in relapsed acute myeloid leukemia after allogeneic stem cell transplantation. *J BUON* 2016; 21(1): 280–281.
- Drozd-Sokołowska J, Karakulska-Prystupiuk E, Biecek P, *et al.* Azacitidine for relapse of acute myeloid leukemia or myelodysplastic

syndrome after allogeneic hematopoietic stem cell transplantation, multicenter PALG analysis. *European Journal of Haematology* 2021; 107: 129–136.

- 32. Schroeder T, Rautenberg C, Haas R, *et al.* Hypomethylating agents for treatment and prevention of relapse after allogeneic blood stem cell transplantation. *Int J Hematol* 2018; 107(2): 138–150.
- Atanackovic D, Luetkens T, Kloth B, et al. Cancer-testis antigen expression and its epigenetic modulation in acute myeloid leukemia. Am J Hematol 2011; 86(11): 918–922.
- 34. Goodyear O, Agathanggelou A, Novitzky-Basso I, et al. Induction of a CD8+ T-cell response to the MAGE cancer testis antigen by combined treatment with azacitidine and sodium valproate in patients with acute myeloid leukemia and myelodysplasia. *Blood* 2010; 116: 1908–1918.
- 35. Almstedt M, Blagitko-Dorfs N, Duque-Afonso J, et al. The DNA demethylating agent 5-aza-2'-deoxycytidine induces expression of NY-ESO-1 and other cancer/testis antigens in myeloid leukemia cells. Leuk Res 2010; 34(7): 899–905.
- Hambach L, Ling KW, Pool J, et al. Hypomethylating drugs convert HA-1 negative solid tumors into targets for stem cell based immunotherapy. Blood 2009; 113: 2715–2722.
- Pinto A, Maio M, Attadia V, *et al.* Modulation of HLA-DR antigens expression in human myeloid leukaemia cells by cytarabine and 5-AZA-2'deoxycytidine. *Lancet* 1984; 2: 867–868.
- Fonsatti E, Nicolay HJM, Sigalotti L, et al. Functional up-regulation of human leukocyte antigen class I antigens expression by 5-aza-2 -deoxycytidine in cutaneous melanoma: immunotherapeutic implications. Clin Cancer Res 2007; 13: 3333–3338.
- Chiappinelli KB, Strissel PL, Desrichard A, et al. Inhibiting DNA methylation causes an interferon response in cancer via dsRNA including endogenous retroviruses. *Cell* 2015; 162: 974– 986.
- 40. Roulois D, Yau HL, Pugh TJ, *et al.* DNAdemethylating agents target colorectal cancer cells by inducing viral mimicry by endogenous transcripts article DNA-demethylating agents target colorectal cancer cells by inducing viral mimicry by endogenous transcripts. *Cell* 2015; 162: 961–973.
- 41. Parker BS, Rautela J and Hertzog PJ. Antitumour actions of interferons: implications for cancer therapy. *Nat Rev Cancer* 2016; 16(3): 131–144.

- Santourlidis S, Trompeter H-I, Weinhold S, et al. Crucial role of DNA methylation in determination of clonally distributed killer cell Ig-like receptor expression patterns in NK cells. J Immunol 2002; 169: 4253–4261.
- Bhagat TD, Chen S, Bartenstein M, et al. Epigenetically aberrant stroma in MDS propagates disease via Wnt/β-catenin activation. *Cancer Res* 2017; 77: 4846–4857.
- 44. Yang H, Bueso-Ramos C, DiNardo C, *et al.* Expression of PD-L1, PD-L2, PD-1 and CTLA4 in myelodysplastic syndromes is enhanced by treatment with hypomethylating agents. *Leukemia* 2014; 28(6): 1280–1288.
- 45. Ørskov AD, Treppendahl MB, Skovbo A, et al. Hypomethylation and up-regulation of PD-1 in T cells by azacytidine in MDS/AML patients: a rationale for combined targeting of PD-1 and DNA methylation. Oncotarget 2015; 6: 9612– 9626.
- Goodyear OC, Dennis M, Jilani NY, et al. Azacitidine augments expansion of regulatory T cells after allogeneic stem cell transplantation in patients with acute myeloid leukemia (AML). Blood 2012; 119: 3361–3369.
- Schroeder T, Fröbel J, Cadeddu RP, et al. Salvage therapy with azacitidine increases regulatory T cells in peripheral blood of patients with AML or MDS and early relapse after allogeneic blood stem cell transplantation. *Leukemia* 2013; 27(9): 1910–1913.
- Choi J, Ritchey J, Prior JL, et al. In vivo administration of hypomethylating agents mitigate graft-versus-host disease without sacrificing graft-versus-leukemia. Blood 2010; 116: 129–139.
- Sánchez-Abarca LI, Gutierrez-Cosio S, Santamaría C, *et al.* Immunomodulatory effect of 5-azacytidine (5-azaC): potential role in the transplantation setting. *Blood* 2010; 115: 107– 121.
- 50. Shouval R, Fein JA, Labopin M, et al. Outcomes of allogeneic haematopoietic stem cell transplantation from HLA-matched and alternative donors: a European Society for Blood and Marrow Transplantation registry retrospective analysis. *Lancet Haematol* 2019; 6(11): e573–e584.
- 51. von Elm E, Altman DG, Egger M, et al. The strengthening the reporting of observational studies in epidemiology (STROBE) statement: guidelines for reporting observational studies. *J Clin Epidemiol* 2008; 61: 344–349.

- 52. Döhner H, Estey E, Grimwade D, *et al.* Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood* 2017; 129: 424–447.
- Bacigalupo A, Ballen K, Rizzo D, et al. Defining the intensity of conditioning regimens: working definitions. *Biol Blood Marrow Transplant* 2009; 15(12): 1628–1633.
- 54. Sorror ML, Storb RF, Sandmaier BM, et al. Comorbidity-age index: a clinical measure of biologic age before allogeneic hematopoietic cell transplantation. J Clin Oncol 2014; 32: 3249– 3256.
- Armand P, Kim HT, Logan BR, et al. Validation and refinement of the Disease Risk Index for allogeneic stem cell transplantation. *Blood* 2014; 123: 3664–3671.
- 56. Harris AC, Young R, Devine S, et al. International, multicenter standardization of acute graft-versus-host disease clinical data collection: a report from the Mount Sinai acute GVHD international consortium. *Biol Blood Marrow Transplant* 2016; 22(1): 4–10.
- 57. Jagasia MH, Greinix HT, Arora M, et al. National institutes of health consensus development project on criteria for clinical trials in chronic graft-versus-host disease: I. The 2014 diagnosis and staging working group report. *Biol Blood Marrow Transplant* 2015; 21: 389–401.e1.
- Gooley TA, Leisenring W, Crowley J, et al. Estimation of failure probabilities in the presence of competing risks: new representations of old estimators. Stat Med 1999; 18: 695–706.
- Kaplan EL and Meier P. Nonparametric estimation from incomplete samples. J Am Stat Assoc 1958; 53: 457–481.
- Mantel N. Evaluation of survival data and two new rank order statistics arising in its consideration. *Cancer Chemother Rep* 1966; 50: 163–170.
- 61. Gray RJ. A class of K-sample tests for comparing the cumulative incidence of a competing risk. *Ann Stat* 1988; 16: 1141–1154.
- Bazarbachi AH, al Hamed R, Malard F, et al. Allogeneic transplant for FLT3 – ITD mutated AML: a focus on FLT3 inhibitors before, during, and after transplant. *Ther Adv Hematol* 2019; 10: 204062071988266.
- 63. Bazarbachi A, Labopin M, Battipaglia G, *et al.* Sorafenib improves survival of FLT3-mutated acute myeloid leukemia in relapse after allogeneic stem cell transplantation: a report of the EBMT

acute leukemia working party. *Haematologica* 2019; 104: E398–E401.

- Rautenberg C, Nachtkamp K, Dienst A, et al. Sorafenib and azacitidine as salvage therapy for relapse of FLT3-ITD mutated AML after allo-SCT. Eur J Haematol 2017; 98: 348–354.
- Perl AE, Martinelli G, Cortes JE, et al. Gilteritinib or chemotherapy for relapsed or refractory FLT3 -mutated AML. New Engl J Med 2019; 381: 1728–1740.
- Stein EM, DiNardo CD, Pollyea DA, et al. Enasidenib in mutant IDH2 relapsed or refractory acute myeloid leukemia. *Blood* 2017; 130: 722–731.
- 67. DiNardo CD, Stein EM, de Botton S, et al. Durable remissions with ivosidenib in IDH1 -mutated relapsed or refractory AML. New Engl J Med 2018; 378: 2386–2398.
- Byrne M, Danielson N, Sengsayadeth S, et al. The use of venetoclax-based salvage therapy for post-hematopoietic cell transplantation relapse of acute myeloid leukemia. Am J Hematol 2020; 95(9): 1006–1014.
- 69. Joshi M, Cook J, McCullough K, *et al.* Salvage use of venetoclax-based therapy for relapsed AML post allogeneic hematopoietic cell transplantation. *Blood Cancer Journal* 2021; 11: 49.
- Schuler E, Wagner-Drouet EM, Ajib S, et al. Treatment of myeloid malignancies relapsing after allogeneic hematopoietic stem cell transplantation with venetoclax and hypomethylating agents – a retrospective multicenter analysis on behalf of the German Cooperative Transplant Study Group. Ann Hematol 2021; 100: 959–968.
- 71. de Lima M, Giralt S, Thall PF, et al. Maintenance therapy with low-dose azacitidine after allogeneic hematopoietic stem cell transplantation for recurrent acute myelogenous leukemia or myelodysplastic syndrome: a dose and schedule finding study. *Cancer* 2010; 116: 5420–5431.
- 72. Oran B, de Lima M, Garcia-Manero G, et al. Maintenance with 5-azacytidine for acute myeloid leukemia and myelodysplastic syndrome patients. Blood 2018; 132: 971.
- 73. Wei AH, Döhner H, Pocock C, *et al*. Oral azacitidine maintenance therapy for acute

myeloid leukemia in first remission. *New Engl J Med* 2020; 383: 2526–2537.

- 74. AMADEUS: randomized study of oral azacitidine vs placebo maintenance in AML or MDS patients after allo-SCT. NCT04173533, ClinicalTrials. gov Identifier, https://clinicaltrials.gov/ct2/show/ NCT04173533
- 75. de Lima M, Roboz GJ, Platzbecker U, *et al.* AML and the art of remission maintenance. *Blood Rev* 2021; 49: 100829.
- 76. Flowers MED, Inamoto Y, Carpenter PA, et al. Comparative analysis of risk factors for acute graft-versus-host disease and for chronic graft-versus-host disease according to National Institutes of Health consensus criteria. *Blood* 2011; 117: 3214–3219.
- 77. Zeiser R and Blazar BR. Acute graft-versushost disease – biologic process, prevention, and therapy. *New Engl J Med* 2017; 377: 2167–2179.
- Zeiser R and Blazar BR. Pathophysiology of chronic graft-versus-host disease and therapeutic targets. *New Engl J Med* 2017; 377: 2565–2579.
- 79. Zeidan AM, Forde PM, Symons H, et al. HLA-Haploidentical donor lymphocyte infusions for patients with relapsed hematologic malignancies after related HLA-haploidentical bone marrow transplantation. *Biol Blood Marrow Transplant* 2014; 20(3): 314–318.
- Ghiso A, Raiola AM, Gualandi F, et al. DLI after haploidentical BMT with post-transplant CY. Bone Marrow Transplant 2015; 50(1): 56–61.
- Goldsmith SR, Slade M, DiPersio JF, et al. Donor-lymphocyte infusion following haploidentical hematopoietic cell transplantation with peripheral blood stem cell grafts and PTCy. *Bone Marrow Transplant* 2017; 52(12): 1623– 1628.
- 82. Peccatori J, Forcina A, Clerici D, et al. Sirolimusbased graft-versus-host disease prophylaxis promotes the in vivo expansion of regulatory T cells and permits peripheral blood stem cell transplantation from haploidentical donors. *Leukemia* 2015; 29(2): 396–405.
- Greco R, Lorentino F, Albanese S, et al. Posttransplantation cyclophosphamide- and sirolimus-based graft-versus-host-disease prophylaxis in allogeneic stem cell transplant. *Transplant Cell Ther* 2021; 27: 776.e1–776.e13.

Visit SAGE journals online journals.sagepub.com/ home/tah

SAGE journals