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# Searching for Mechanisms Underlying the Assembly of Calcium Entry Units: The Role of Temperature and pH

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**Abstract:** Store-operated  $\text{Ca}^{2+}$  entry (SOCE) is a mechanism that allows muscle fibers to recover external  $\text{Ca}^{2+}$ , which first enters the cytoplasm and then, via SERCA pump, also refills the depleted intracellular stores (i.e., the sarcoplasmic reticulum, SR). We recently discovered that SOCE is mediated by Calcium Entry Units (CEUs), intracellular junctions formed by: (i) SR stacks containing STIM1; and (ii) I-band extensions of the transverse tubule (TT) containing Orai1. The number and size of CEUs increase during prolonged muscle activity, though the mechanisms underlying exercise-dependent formation of new CEUs remain to be elucidated. Here, we first subjected isolated extensor digitorum longus (EDL) muscles from wild type mice to an ex vivo exercise protocol and verified that functional CEUs can assemble also in the absence of blood supply and innervation. Then, we evaluated whether parameters that are influenced by exercise, such as temperature and pH, may influence the assembly of CEUs. Results collected indicate that higher temperature (36 °C vs. 25 °C) and lower pH (7.2 vs. 7.4) increase the percentage of fibers containing SR stacks, the n. of SR stacks/area, and the elongation of TTs at the I band. Functionally, assembly of CEUs at higher temperature (36 °C) or at lower pH (7.2) correlates with increased fatigue resistance of EDL muscles in the presence of extracellular  $\text{Ca}^{2+}$ . Taken together, these results indicate that CEUs can assemble in isolated EDL muscles and that temperature and pH are two of the possible regulators of CEU formation.

**Keywords:** exercise; excitation-contraction (EC) coupling; skeletal muscle; store-operated  $\text{Ca}^{2+}$  entry (SOCE)



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## 1. Introduction

The role of external  $\text{Ca}^{2+}$  in skeletal muscle contractility has been overlooked for many years, as excitation-contraction (EC) coupling, the mechanism that allows activation of muscle contraction by transducing the action potential into release of  $\text{Ca}^{2+}$  from the sarcoplasmic reticulum (SR), is mechanical in skeletal muscle and does not depend on  $\text{Ca}^{2+}$  entry as in cardiac and smooth muscle [1–6]. In mechanical coupling, the  $\alpha$ -1s subunit of DHPRs, voltage-gated L-type  $\text{Ca}^{2+}$  channels (also known as Cav1.1) of transverse tubules (TTs) act as a voltage sensor [5,7], which directly activates the SR  $\text{Ca}^{2+}$  release from RyR type-1 (RyR1) channels [8–10]. The intracellular sites that allow mechanical interaction between DHPR and RyR1 are called  $\text{Ca}^{2+}$  Release Units (CRUs) or triads, intracellular junctions formed by the association of two SR terminal cisternae with a central TT [11,12].

Recently the role that  $\text{Ca}^{2+}$  entry may play in skeletal muscle function has attracted new attention. A pathway known as excitation-coupled  $\text{Ca}^{2+}$  entry (ECCE), independent of SR store depletion and not required for EC coupling itself, was identified as a slow  $\text{Ca}^{2+}$  entry

pathway through DHPs based on a retrograde signal from RyR1 to Cav1.1 [13–16]. ECCE causes  $\text{Ca}^{2+}$  influx in response to physiological triggers [17,18]. In addition, a mechanism triggered by depletion of intracellular  $\text{Ca}^{2+}$  stores (endoplasmic/sarcoplasmic reticulum, ER and SR), known as store-operated  $\text{Ca}^{2+}$  entry (SOCE) [19–22], was also reported in skeletal muscle [23]. SOCE is a pathway mainly mediated by the interaction between (a) stromal interaction molecule-1 (STIM1), a protein placed in the ER/SR membrane, which has an intra-luminal domain that acts as  $\text{Ca}^{2+}$  sensor; and (b) Orai1, a protein that mediates  $\text{Ca}^{2+}$  release-activated (CRAC) current and is placed in external membranes or TTs [24–34].

The mechanism of activation of SOCE was first studied in non-excitable cells:  $\text{Ca}^{2+}$  depletion of internal ER stores induces dimerization of STIM1 and its translocation towards the plasma membrane (PM), enabling in this way STIM1 to interact and activate Orai1  $\text{Ca}^{2+}$  channels [34,35]. In skeletal muscle, SOCE also relies on the concerted activity of STIM1 and Orai1 [25,30,36–38], as supported by the finding that SOCE is abolished in mice lacking STIM1 [39] and in muscle fibers from dominant-negative and muscle-specific Orai1-knockout mice [40,41]. The exact stoichiometry of the active SOCE complex is currently unclear [42,43]. Several studies hypothesized a 1:1 STIM1:Orai1 stoichiometry, with a STIM1s dimer that interacts with a pair of Orai1 C-terminal subunits [44–47]. It has also been reported that STIM1-Orai1 complexes form as a result of the interaction of one STIM1 dimer with one Orai1 channel [22].

The idea that impairment in SOCE can be behind muscle pathologies characterized by increased fatigability and reduced SR  $\text{Ca}^{2+}$  release [48] has recently increased the interest in this mechanism in the muscle system. Indeed, aberrant SOCE function was associated with several forms of muscle dysfunction: weakness in aging [49–51], oversensitivity to heat in malignant hyperthermia [52,53], and muscular dystrophy in mdx mice [54–57]. In addition, mutations in STIM1 and Orai1 have been linked to tubular aggregate myopathy (TAM), a rare condition characterized by muscle pain, cramping, weakness, and the presence of peculiar remodeling of the SR [58–65].

Some authors reported that SOCE in skeletal muscle fibers has evolved in a different mechanism that is activated in two different modes: (i) rapidly activated SOCE (within milliseconds) of limited amplitude (phasic SOCE); and (ii) a slowly activated, large-scale SOCE or chronic SOCE [64,66–70]. Phasic SOCE in muscle fibers is activated significantly faster than in non-excitable cells, where the process from ER store depletion to Orai1 channel activation takes tens of seconds [35,67,69–73]. Rapid activation of SOCE in skeletal fibers could be explained by STIM1 and Orai1 being pre-assembled in specific sites. For this reason, initially it was proposed that SOCE would occur in triads, the intracellular junctions deputed to EC coupling (see above). This hypothesis was based on the assumption that in triads SR and TT membranes, which contain respectively STIM1 and Orai1, are already associated and would easily allow rapid activation of SOCE. A STIM1 splice variant highly expressed in skeletal muscle (STIM1-long) was proposed to account for rapid SOCE activation [74,75]. Recently, some authors also suggested that slow and fast SOCE may use different pools of Orai1 channels [76].

While there was always general agreement about the important role that SOCE plays in limiting muscle fatigue [40,41,50,51,77], exactly how and where STIM1/Orai1 interact to allow  $\text{Ca}^{2+}$  entry in skeletal muscle fibers has been debated for several years. Recent experimental evidence collected in our laboratory indicated that intracellular junctions, named  $\text{Ca}^{2+}$  Entry Units (CEUs), formed by the association of SR and TT at the I band, are the most likely sites of interaction between STIM1 and Orai1 during SOCE [77–83]. CEUs are few and small in control conditions (the main reason that did not allow their prompt identification), but increase in number and size during exercise [77] to then disassemble following recovery [81]. CEU assembly during exercise requires: (i) remodeling of SR membranes at the I band to form stacks of flat cisternae (i.e., SR stacks); and (ii) elongation of TTs from the triad toward the Z line. This remodeling promotes enhanced STIM1 and Orai1 colocalization and increased fatigue resistance in the presence of extracellular

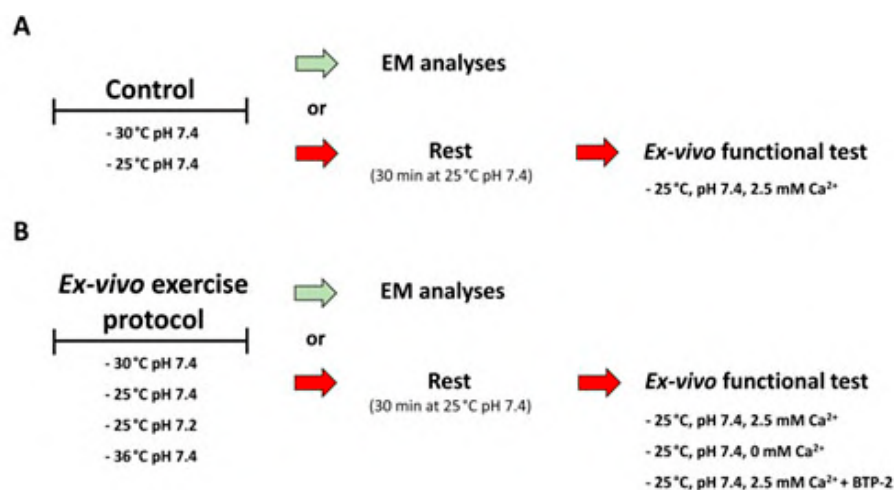
$\text{Ca}^{2+}$  [77]. After proposing that these junctions provide a structure to recover  $\text{Ca}^{2+}$  ions via SOCE from the extracellular space [77,78], the conclusive evidence of CEUs being a site for SOCE came with the demonstration that: (i) these junctions promote increased rate on  $\text{Mn}^{2+}$  quench, i.e., the gold-standard technique used to assess entry of divalent cations from the extracellular space [77,78,81,82]; and (ii) exercise-dependent enhancement of SOCE, due to assembly of additional CEUs, does not occur in muscle fibers lacking Orai1 [81]. Finally, as proof of principle, we also found that CEUs are constitutively assembled in muscle fibers of mice lacking CASQ1 [82], which develop quickly to SR depletion (i.e., the putative trigger of STIM1 aggregation and SOCE activation) during repetitive stimulation [84].

Together, these findings indicate that CEUs are dynamic intracellular junctions between SR and TTs that form when muscle fibers need to use external  $\text{Ca}^{2+}$  to replenish depleted SR stores, hence limiting muscle fatigue. Nevertheless, the mechanisms that promote the formation of SOCE-sites (i.e., CEUs) during exercise and disassembly during post-training recovery are still obscure. In this work, we first: (i) investigated if CEUs can assemble in the absence of blood supply and innervation in isolated extensor digitorum longus (EDL) muscles stimulated ex vivo; and then (ii) studied the influence of temperature and extracellular pH in CEU assembly, two parameters that change physiologically during exercise in vivo [85–89].

## 2. Results

### 2.1. Functional Calcium Entry Units Can Assemble Ex Vivo

We recently demonstrated that acute exercise in vivo promotes the assembly of CEUs, intracellular junctions located at the I band that mediate SOCE. CEUs are formed by the association of two distinct components: SR stacks and I band extensions of TTs [77,78,81]. To verify whether exercise-dependent assembly of CEUs can take place in the absence of blood supply and innervation, we subjected isolated EDL muscles of adult wild type (WT) mice to an ex vivo exercise protocol at 30 °C (see Section 4. Materials and Methods and scheme in Figure 1 for additional detail).

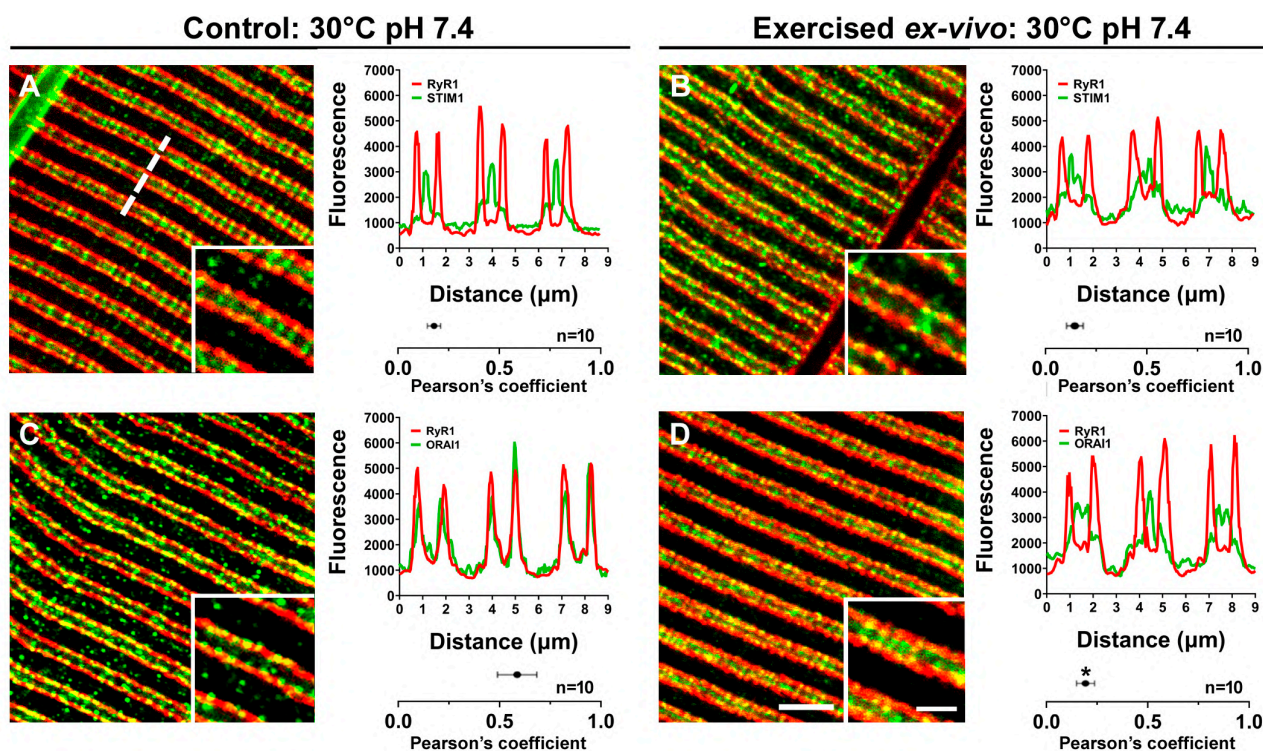


**Figure 1.** Schematic representation of the ex vivo experimental procedures. (A) Control EDL muscles were kept at rest in KH solution for 45 min (the same duration of the ex vivo exercise protocol) and then either immediately fixed for EM analysis (green arrow) or kept in a rest condition for 30 min (25 °C, pH 7.4) and then subjected to the ex vivo functional test (red arrows). (B) EDL muscles exercised ex vivo were either immediately fixed for EM analysis or kept in rest condition for 30 min (25 °C, pH 7.4) and then subjected to the ex vivo functional test.

Samples from two different groups were fixed for immunofluorescence and electron microscopy (EM): (a) EDL muscles that were not electrically stimulated (controls), but simply kept in the solution at 30 °C for 45 min (the same duration of the ex vivo exercise protocol); (b) EDL muscles subjected to the ex vivo exercise protocol. To determine the



subcellular co-localization between STIM1 and Orai1 under control conditions and to verify if STIM1-Orai1 colocalization increases following the ex vivo exercise protocol (as in vivo during acute exercise [77]), small bundles of EDL fibers were double-labeled in immunofluorescence experiments (Figure 2) as follows: RyR1 vs. STIM1 (Figure 2A,B) and RyR1 vs. Orai1 (Figure 2C,D).

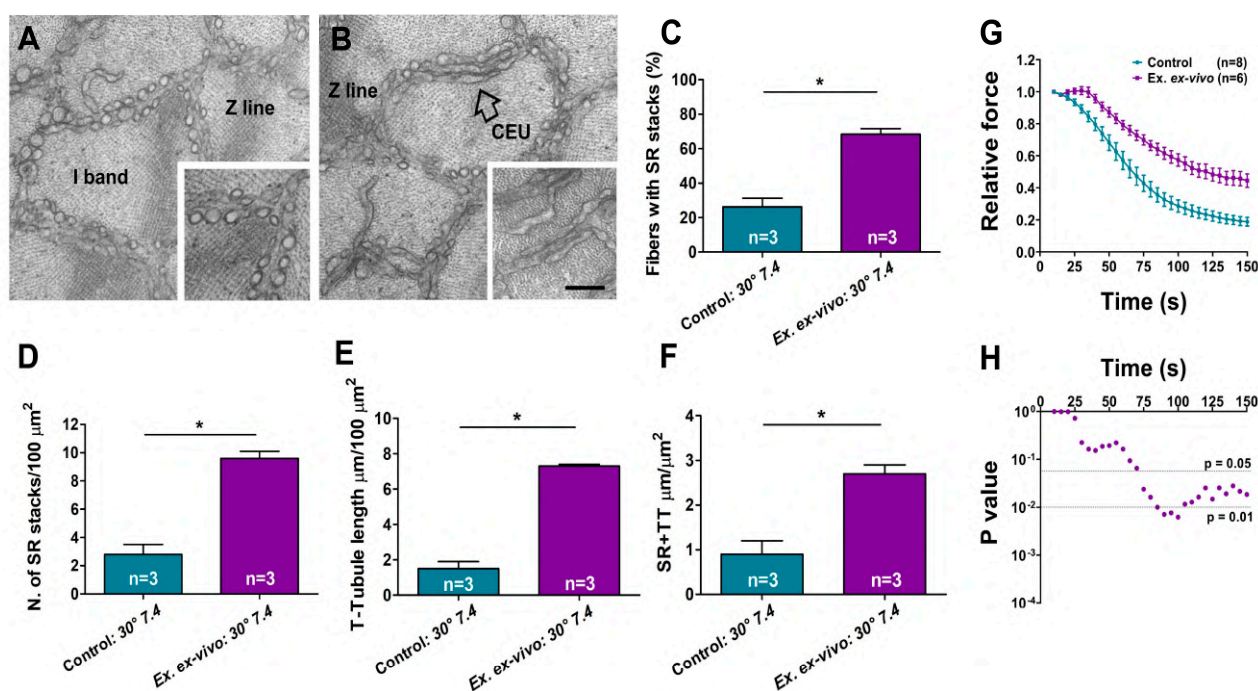


**Figure 2.** Sarcomeric localization of STIM1 and Orai1 before and after an ex vivo protocol performed at 30 °C and pH 7.4. Representative immunofluorescence images of EDL fibers showing RyR1 and STIM1 (A,B) and RyR1-Orai1 (C,D) double-staining. Each panel contains also a fluorescence intensity profile along three sarcomeres (see dashed line in (A)) and the Pearson's correlation coefficient value, i.e., a method of measuring the covariance of pixel intensities, given as the mean  $\pm$  SEM. \*  $p < 0.01$ , compared to fibers from control mice;  $n$  = number of images analyzed. Scale bar: 2.5  $\mu$ m (insets: 1  $\mu$ m).

In Boncompagni et al., 2017 [77], we showed that in resting muscle STIM1 is primarily localized throughout the entire I band region, while Orai1 has a preferential localization in TTs at the triads. Results collected in experiments performed for the present work confirmed previous findings (Figure 2): the level of Orai1 co-localization with RyR1 in the resting condition is high, as shown by Pearson's correlation coefficient value (Figure 2C). On the other hand, in muscles exercised ex vivo (Figure 2D) a fraction of the Orai1 signal shifted toward the I band, decreasing in this way RyR1-Orai1 co-localization, as shown by peaks of fluorescence (traces in Figure 2D) and by the decreased Pearson's correlation coefficient value (compared with Pearson's correlation coefficient values in Figure 2C;  $p < 0.01$ ). As the subcellular localization of STIM1 does not change following exercise (i.e., STIM1 is localized primarily throughout the I band region, also after the ex vivo exercise protocol; Figure 2B), the increased presence of Orai1 at the I band (Figure 2D) is consistent with an increased co-localization with STIM1.

Muscles fixed and processed for EM were used to verify the presence of CEUs (Figure 3A,B). The presence of fully assembled CEUs was assessed by quantitative analysis of their two main components, i.e., SR stacks and TT extensions at the I band. We performed quantitative analysis of four different parameters that are indicative of the structural assembly of CEUs: (i) the percentage of fibers containing SR stacks (Figure 3C); (ii) the number of

SR stacks/100  $\mu\text{m}^2$  of cross-section (Figure 3D); (iii) the extension of TTs at the I band in 100  $\mu\text{m}^2$  of cross-section (Figure 3E); and finally (iv) the extension of SR-TT contacts at the I band (Figure 3F). The data collected in Figure 3 clearly indicate that CEUs can assemble ex vivo following an exercise protocol at 30 °C and pH 7.4, as all the parameters quantified in Figure 3C–F are increased following the ex vivo exercise protocol (see also Table S1). The increased inferred colocalization of STIM1 and Orai1 at the I band (data in Figure 2) is supported by the elongation of T-tubules at the I band (Figure 3E), which underlines the translocation of Orai1 at the I band.

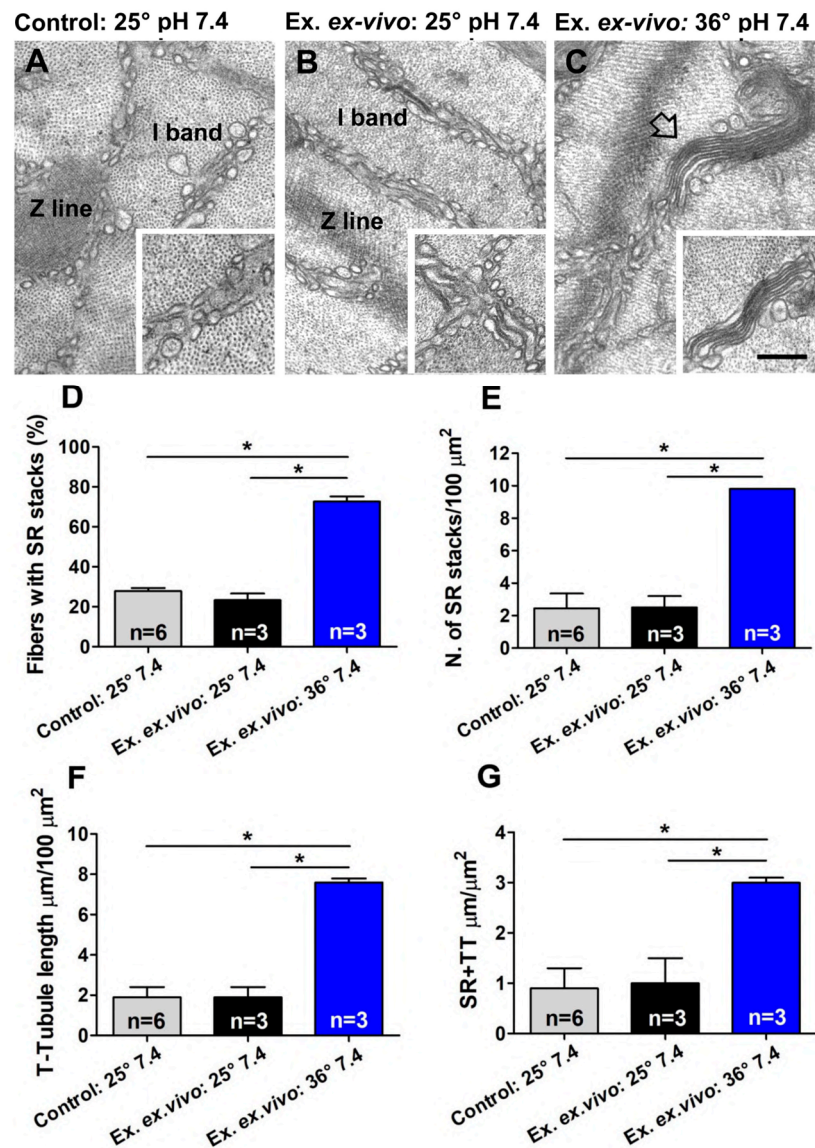


**Figure 3. Quantitative and functional analysis of  $\text{Ca}^{2+}$  entry units (CEUs) assembled ex vivo at 30 °C and pH 7.4.** (A,B) EM images of fibers from isolated EDLs that: (i) have rested at 30 °C and pH 7.4 for 45 min (panel (A)); and (ii) have been subjected to an ex vivo exercise protocol performed at 30 °C and pH 7.4 (panel (B)). The inset in panel A shows SR appearance at the I band; the empty arrow and the inset in panel (B) shows the remodeling of SR in stacks. (C,D) Percentage of fibers containing SR stacks and number of SR stacks/100  $\mu\text{m}^2$  of section. (E,F) Extension of TTs at the I band in 100  $\mu\text{m}^2$  of section and analysis of SR-TT contact length. (G) Ex vivo functional test showing time course of average relative force decay in EDL muscles previously exercised ex vivo and of controls (experiments performed at 30 °C and pH 7.4). Arrow points to the bump-phase, previously characterized in two papers by Michelucci and colleagues as the phase of activation of SOCE [81,82]. (H) Semi-log plot showing results of multiple unpaired *t* test followed by Tukey post hoc test. Data are shown as mean  $\pm$  SEM (\* *p* < 0.05). *n* = number of EDL analyzed. Scale bar: (A,B) = 0.1  $\mu\text{m}$ ; insets = 0.2  $\mu\text{m}$ .

We previously demonstrated that the exercise-dependent assembly of CEUs increases the resistance to fatigue of EDL muscles in the presence of extracellular  $\text{Ca}^{2+}$  [77]. Here, we verified if muscles containing a greater number of CEUs (pre-exercised ex vivo) are also more resistant to fatigue than controls (Figure 3G). EDL muscles were subjected to an ex vivo functional test (see Section 4. Materials and Methods and scheme in Figure 1 for additional detail): as shown in Figure 3G, pre-exercised muscles display greater resistance to fatigue than control muscles during a fatigue protocol in which muscles were stimulated at 60 Hz, for 1 sec every 5 sec (in the presence of extracellular  $\text{Ca}^{2+}$ ).

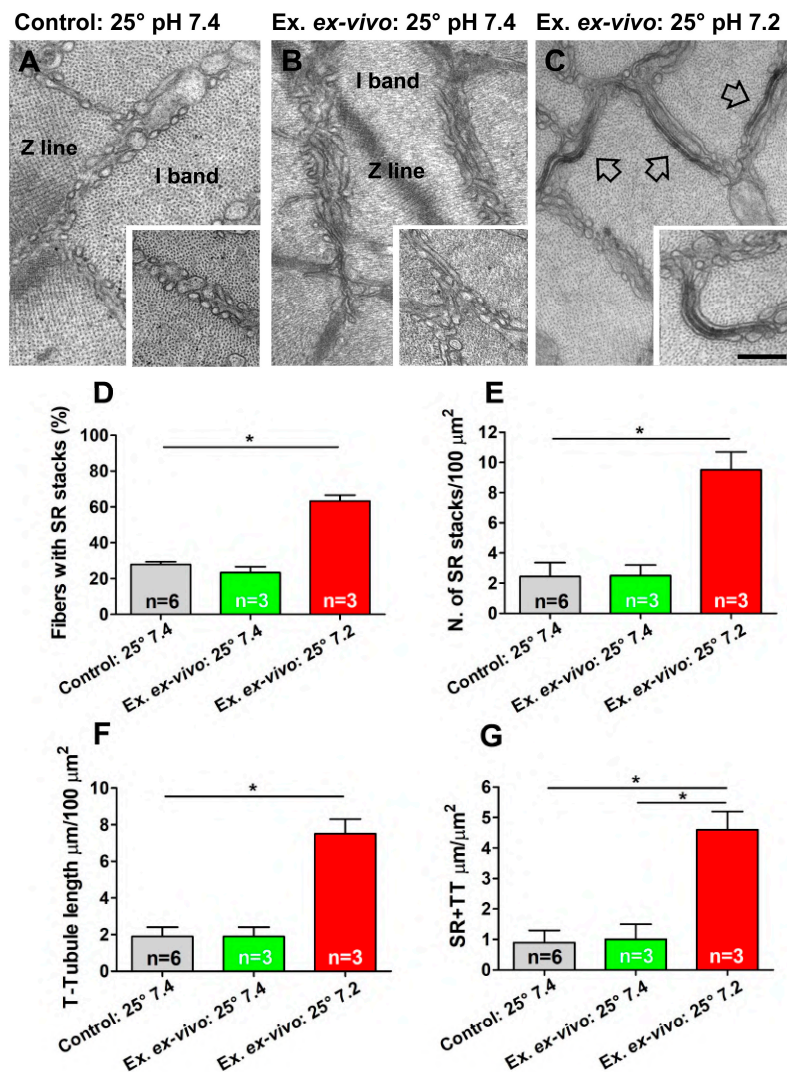
## 2.2. The Assembly of CEUs Is Affected by Temperature and Extracellular pH

CEUs assembled *in vivo* during a fatigue protocol in which mice ran on treadmill for about 1 h [77]. Nevertheless, the mechanism underlying the SR and TT remodeling necessary to increase number and size of CEUs during exercise is unknown. Here, we tested the effect of two physiological parameters that change during exercise (Figures 4 and 5), i.e., body temperature rises, and extracellular pH becomes more acidic due to the combined effect of increased CO<sub>2</sub> and lactic acid production [86–89].



**Figure 4.** Electron-micrographs and quantitative analysis of assembly of CEUs at 25 vs. 36 °C (pH 7.4). (A–C) EM images of fibers in isolated EDL muscles that: (i) have rested at 25 °C and pH 7.4 for 45 min (panel (A)); (ii) have been subjected to the ex vivo exercise protocol performed at 25 °C and pH 7.4 (panel (B)); and (iii) have been subjected to the ex vivo exercise protocol performed at 36 °C and pH 7.4 (panel (C)). The insets in panels (A,B) show SR appearance at the I band; the empty arrow and the inset in panel (C) show the remodeling of SR in stacks. (D,E) Percentage of fibers containing SR stacks and number of SR stacks/100 μm<sup>2</sup> of section. (F,G) Extension of TTs at the I band in 100 μm<sup>2</sup> of section and analysis of SR-TT contact length. Data are shown as mean ± SEM (\* *p* < 0.05). n = number of EDL analyzed. Scale bar: (A–C) = 0.1 μm; insets = 0.2 μm.



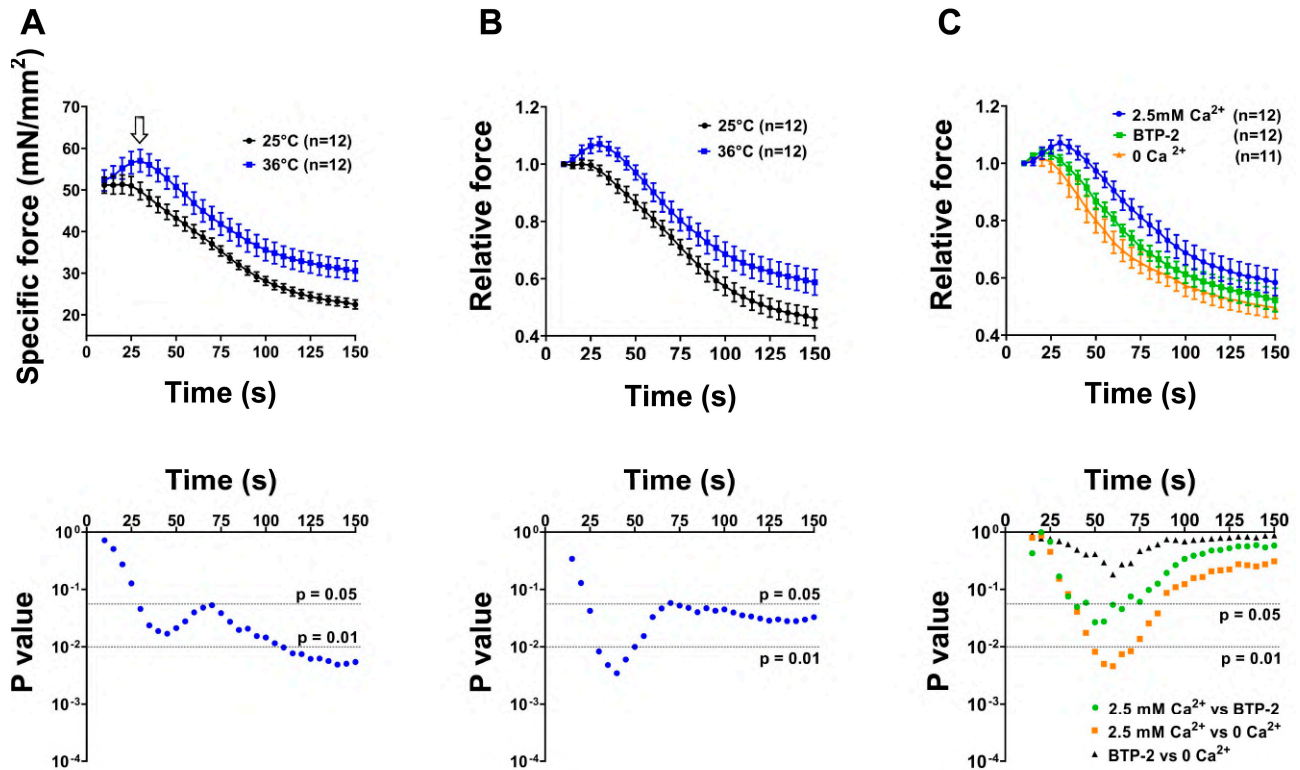


**Figure 5.** Electron-micrographs and quantitative analysis of assembly of CEUs at pH 7.4 vs. 7.2 (25 °C). (A–C) EM images in isolated EDL muscles that: (i) have rested at 25 °C and pH 7.4 (panel (A)); (ii) have been subjected to the ex vivo exercise protocol performed at 25 °C and pH 7.4 (panel (B)); and (iii) have been subjected to the ex vivo exercise protocol performed at 25 °C and pH 7.2 (panel (C)). The insets in panel (A,B) show SR appearance at the I band; the empty arrows and the inset in panel (C) show the remodeling of SR in stacks. (D,E) Percentage of fibers containing SR stacks and number of SR stacks/100 μm<sup>2</sup> of section. (F,G) Extension of TTs at the I band in 100 μm<sup>2</sup> of section and analysis of SR-TT contact length. Data are shown as mean ± SEM (\*  $p < 0.05$ ). n = number of EDL analyzed. Scale bar: (A–C) = 0.1 μm; insets = 0.2 μm.

### 2.3. CEUs Assembled Ex Vivo (i.e., at Higher Temperature and Lower Extracellular pH) Promote Enhanced Resistance to Fatigue in Presence of External Ca<sup>2+</sup>

Following EM analysis shown in Figures 4 and 5, we performed ex vivo functional experiments to assess the contractility of EDL muscles in the presence of either external Ca<sup>2+</sup> or blocking Ca<sup>2+</sup> entry by removing external Ca<sup>2+</sup> (replaced by an equimolar concentration of Mg<sup>2+</sup>) or supplementing the solution with a compound that is frequently used to block SOCE, i.e., BTP-2 [90] (Figures 6 and 7). We analyzed resistance to fatigue using a short protocol of the duration of 2.5 min (see ex vivo functional test in Section 4. Materials and Methods), in EDL muscles previously subjected to the ex vivo exercise protocol at the different conditions reported in Figures 4 and 5. Specifically, we compared muscles fatigued ex vivo at 25 °C at pH 7.4 vs. 36 °C at pH 7.4 (experiments designed to test the effect of temperature on CEU assembly; structural analysis in Figure 4) and 25 °C at

pH 7.4 vs. 25 °C at pH 7.2 (experiments designed to test the effect of pH; structural analysis in Figure 5). Control muscles not subjected to the ex vivo exercise protocol (analyzed with EM in Figures 4 and 5) were excluded from these experiments. Results of these experiments are shown in Figures 6 and 7 and summarized in the following section.

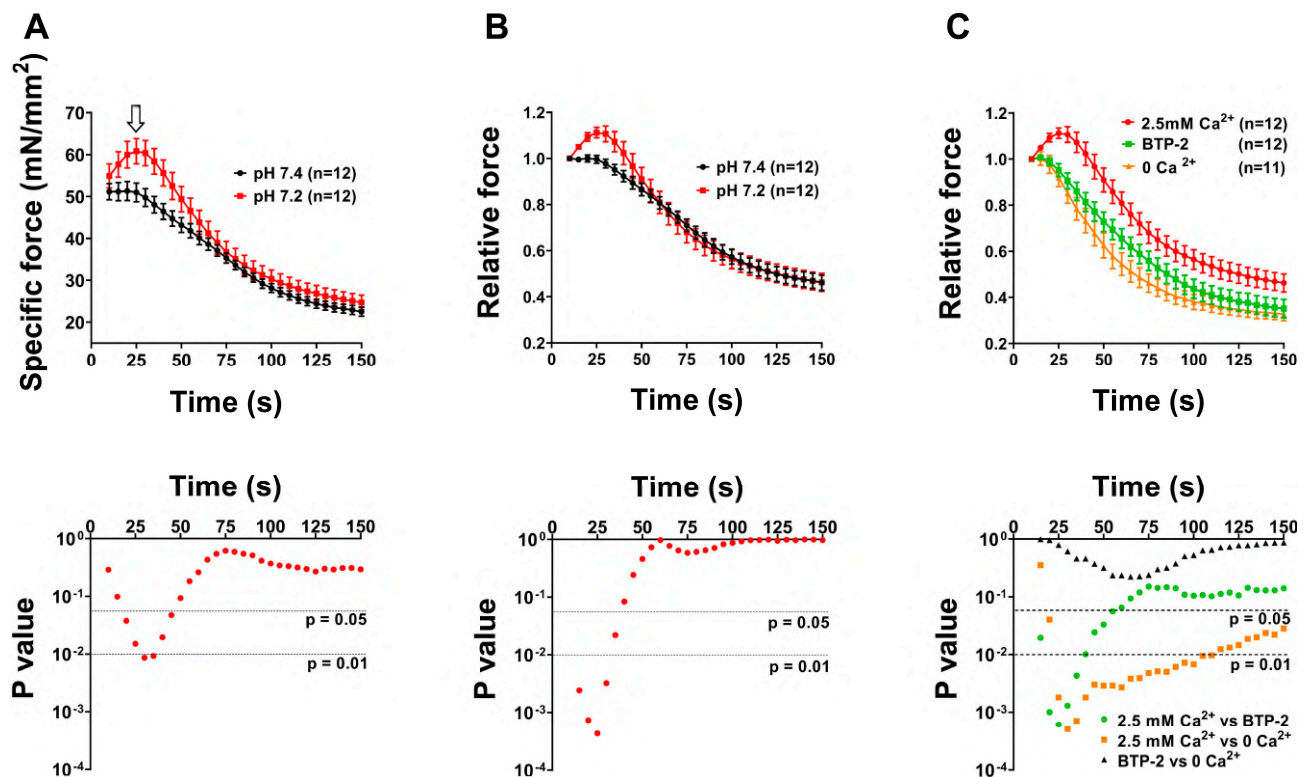


**Figure 6.** Contractile force of EDL muscles after the ex vivo exercise protocol at 25 vs. 36 °C (pH 7.4). (A) Time course of average specific force decay during 30 consecutive frequency stimulus trains (60 Hz, 1 s, every 5 s), normalized to CSA, in EDL muscles previously ex vivo exercised at 25 °C or at 36 °C. Arrow points to the bump-phase, previously characterized in two papers by Michelucci and colleagues, as the phase of activation of SOCE [81,82]. (B) Data represented as relative force (normalized to the first stimulus train). (C) Time course of average relative force decay (in presence or absence of 2.5 mM extracellular Ca<sup>2+</sup> and in a solution supplemented with 10 μM BTP-2) in EDL muscles previously subjected to the ex vivo protocol at 36 °C. Data are shown as mean ± SEM. Bottom panels represent semi-log plot showing results of multiple unpaired *t* test (first two panels) or two-way repeated measures ANOVA followed by Tukey post hoc test (right panel). *n* = number of EDL muscles analyzed for each condition.

EDL muscles stimulated at 36 °C and pH 7.2 exhibited an increased ability to maintain contractile force during a high frequency fatigue protocol in comparison to those stimulated at 25 °C and pH 7.4, shown by the reduced decay of muscle force along the protocol (Figure 6A,B and Figure 7A,B). Note the bump-phase (indicated by arrows in Figures 6A and 7A), which was previously characterized in two papers by Michelucci and colleagues as the phase of activation of SOCE [81,82]. To demonstrate whether the increased fatigue resistance registered in EDL muscles exercised ex vivo at 36 °C (pH 7.4) and at 7.2 (25 °C) was due to increased Ca<sup>2+</sup> entry via SOCE, we performed parallel experiments conducted using either: (i) a Ca<sup>2+</sup>-free solution, in which Ca<sup>2+</sup> was replaced by an equimolar concentration of Mg<sup>2+</sup>; or (ii) a Ca<sup>2+</sup>-containing solution supplemented with 10 μM BTP-2 (Figures 6C and 7C), an established inhibitor of SOCE [77,79,90–92]. These experiments indicated that the enhanced fatigue resistance of EDL muscles exercised ex vivo at 36 °C (pH 7.4) and at pH 7.2 (25 °C) containing ex vivo assembled CEUs (Figures 4 and 5) was effectively due to



the entry of extracellular  $\text{Ca}^{2+}$  via SOCE, as inhibition of  $\text{Ca}^{2+}$  entry results in faster decay of contractile force (Figures 6C and 7C).



**Figure 7.** Contractile force of EDL muscle after the ex vivo exercise protocol at pH 7.4 vs. 7.2 (25 °C). (A) Time course of average specific force decay during 30 consecutive frequency stimulus trains (60 Hz, 1 s, every 5 s), normalized to CSA, in EDL muscles previously ex vivo exercised at pH 7.4 or at pH 7.2. Arrow points to the bump-phase, previously characterized in two papers by Michelucci and colleagues as the phase of activation of SOCE [81,82]. (B) Data represented as relative force (normalized to the first stimulus train). (C) Time course of average relative force decay (in presence or absence of 2.5 mM extracellular  $\text{Ca}^{2+}$  and in a solution supplemented with 10  $\mu\text{M}$  BTP-2) in EDL muscles previously subjected to the ex vivo protocol at pH 7.2. Data are shown as mean  $\pm$  SEM. Bottom panels represent semi-log plot showing results of multiple unpaired *t* test (first two panels) or two-way repeated measures ANOVA followed by Tukey post hoc test (right panel). *n* = number of EDL muscles analyzed for each condition.

### 3. Discussion

#### 3.1. Background

We recently found that acute exercise induces a striking remodeling of SR and TTs at the I band of sarcomeres, which leads to increased STIM1/Orai1 colocalization, enhanced  $\text{Ca}^{2+}$  entry via SOCE, and improved fatigue resistance [77–79,81–83]. These new junctions were named  $\text{Ca}^{2+}$  entry units (CEUs), dynamic entities that form during exercise to promote recovery of external  $\text{Ca}^{2+}$  to then disassemble during recovery; the mechanisms underlying their assembly are still obscure. Here, we first investigated whether CEUs can assemble in isolated muscles, in the absence of nerve and blood supply, applying an ex vivo incremental protocol to mimic the treadmill protocol previously used in vivo [77]. Then, we tested if temperature and pH, physiological parameters that change in muscle during exercise [86–89], may affect the exercise-mediated assembly of CEUs, comparing different physiological conditions: (a) 36 vs. 25 °C at pH 7.4; (b) pH 7.2 vs. 7.4 at 25 °C. After these protocols, we performed a combination of confocal and electron microscopy and ex vivo contractile experiments (in the presence or absence of external  $\text{Ca}^{2+}$ ) to quantify the assembly of functional CEUs in the different experimental conditions.

### 3.2. Main Findings of the Study

Data collected in the first part of this work (Figures 2 and 3) suggest that CEUs can also assemble in isolated muscles during an ex vivo exercise protocol (in which temperature and extracellular pH were respectively kept at 30 °C and 7.4) without innervation and blood supply. The presence of fully assembled CEUs were verified by qualitative and quantitative EM analysis of EDL muscles excised immediately after the ex vivo exercise protocol (see Figure 1 for a scheme of the experimental procedures). These structures were analyzed in their two main components (i.e., SR stacks and elongated TTs). Increased presence of Orai1 at the I band following the ex vivo protocol (Figure 2) in immunofluorescence experiments, which implies increased co-localization with STIM1, is the consequence of TTs being more extended at the I band following the exercise protocol (Figure 3 and Table S1). Note that the I band is the sarcomeric region where CEUs assemble also in vivo [77]. The increased length of contacts between SR stacks and TTs would provide the structural framework for increased STIM1-Orai1 colocalization and for the functional interaction between them. Indeed, during the fatigue protocol in Figure 3, the force generation in the presence of external  $\text{Ca}^{2+}$  is increased in those muscles that have increased presence of CEUs.

In the second part of this study, we tested the effect of temperature and of extracellular pH on the assembly of CEUs (Figures 4–7). Results collected indicate that the remodeling of SR in stacks and the elongation of TTs at the I band are both temperature and pH sensitive. Indeed, the EM analysis shows how number of SR stacks, elongation of TTs, and finally, the establishment of contact between SR stacks and TTs are structural parameters all influenced both by temperature and extracellular pH (Figures 4 and 5 and Tables S2 and S3). The classic features of sarcotubular remodeling that emerged from the quantitative analysis were also clearly visible in EM images of cross-sections of EDL muscle fibers stimulated at 36 °C (pH 7.4) and pH 7.2 (25 °C), where the insets highlight the typical flattening of SR into stacks (EM images in Figures 4 and 5).

It is also important to underline two other aspects of these findings:

- a. Both higher temperature (36 °C) and lower pH (7.2) promote CEU assembly independently, as: (i) in experiments comparing 25 °C vs. 36 °C, the extracellular pH was kept in both at 7.4 (Figure 4 and Table S2); (ii) when we tested pH 7.2 vs. 7.4, temperature was kept in both at 25 °C (Figure 5 and Table S3). Yet in both conditions, formation of CEUs was greatly improved only changing one of the two parameters (Figures 4 and 5 and Tables S2 and S3).
- b. Numeric values collected in quantitative analysis of EDL muscles stimulated at 25 °C (pH 7.4) (Figures 4 and 5 and Tables S2 and S3) were not different from those of control muscles, i.e., EDL muscles held at 25 °C (pH 7.4) for 45 min and not subjected to the ex vivo exercise protocol (Figures 4 and 5 and Tables S2 and S3).

Structural assembly of CEUs assessed in Figures 4 and 5 by EM was followed by functional protocols aiming to determine fatigue resistance in different conditions: (i) in the presence of external  $\text{Ca}^{2+}$ , (ii) in nominally  $\text{Ca}^{2+}$  free solution, and (iii) in the presence of external  $\text{Ca}^{2+}$ , but in a solution supplemented with a SOCE inhibitor (BTP-2) (Figures 6 and 7). The results collected indicate that muscles containing more CEUs display a greater fatigue resistance (Figures 6A,B and 7A,B), which is reduced in conditions that limit  $\text{Ca}^{2+}$  entry from the extracellular space (Figures 6C and 7C).

One aspect of our findings deserves additional attention: when comparing structural assembly of CEUs by EM at 30 °C and 36 °C (Figure 3 vs. Figure 4; see also Tables S1 and S2) surprisingly, we found no significant quantitative difference in the formation of SR stacks and elongation of TTs. Though, functional analysis suggests that activation of SOCE at 30 °C is not as prompt as at 36 °C, as shown by the slightly less pronounced bump-phase at the lower temperature (compare Figure 3G with Figure 6). The reason for this difference is still unclear and deserves a more in-depth investigation. Though we could speculate that while SR stacks and TT elongation are fully assembled already at 30 °C, at 36 °C either

(i) part of STIM1 and Orai1 are already pre-assembled or (ii) they respond to electrical stimulation more promptly at higher temperature.

### 3.3. Our Findings in the Context of Existing Knowledge

It is well known that skeletal muscle tissue is capable of great plasticity and shows a wide spectrum of adaptations in response to mechanical and metabolic stress induced by physical exercise.

- *Changes in temperature.* STIM1-mediated SOCE has been proposed to play a critical role in the development, contraction, fatigue resistance, and remodeling of skeletal muscle cells [39,40]. Xiao and colleagues demonstrated that heating cells induce STIM1 clustering and activation of STIM1/Orai1-mediated  $\text{Ca}^{2+}$  influx [93]. Moreover, the same authors demonstrated that temperature sensitivity of STIM1-dependent  $\text{Ca}^{2+}$  signaling can impact gene expression in immune cells in response to heat alone, suggesting that STIM1 acts as a temperature sensor. Muscle activity generates heat: the temperature of human skeletal muscle can increase from 33 °C up to 39 °C during exercise [94], raising the possibility that heat-induced  $\text{Ca}^{2+}$  influx via SOCE could play a role in skeletal muscle physiology. We have recently shown that exercise-dependent assembly of CEUs could contribute to exertional heat stroke when exercise is performed in adverse environmental conditions [92]. In the present study, the assembly of functional CEUs was favored by physiological temperature, when compared to 25 °C. In addition, functional activation of SOCE (shown by the bump-phase, pointed by arrows in Figures 6 and 7) was more pronounced at 36 °C than at 25 or 30 °C.
- *Changes in pH.* Cytosolic pH in cells is tightly regulated [95], because dramatic differences in protein function and cell behavior are driven by relatively small changes in pH. Under normal physiological conditions extracellular pH in healthy tissues is maintained within a narrow range between 7.3 and 7.4, while intracellular pH is kept between 7.1 and 7.2. For example, increases in pH are permissive for growth factor-induced cell proliferation [96], cell cycle progression [97,98], and differentiation [99,100]. Regarding pH regulation of  $\text{Ca}^{2+}$  entry, SOCE-mediated platelet aggregation is dependent on extra-platelet pH [101]. In exercising muscle, both extracellular and intracellular pH can drop as low as 6.9 and 6.7, respectively [102]. Results regarding pH regulation on STIM1-Orai1 interaction are controversial, and reduction of pH has not always been associated with gain of function on SOCE. Tsujikawa and colleagues demonstrated that the Orai1/STIM1 channel is regulated by changes of both intracellular and extracellular pH [103]. Authors showed that acidic internal and external pH reduce STIM1/Orai1 interaction, whereas alkaline intracellular and extracellular pH enhance SOCE activity. Mancarella and colleagues showed that intracellular low pH caused by oxidative stress induces uncoupling of Orai1 and STIM1, thereby inhibiting ICRAC, and that intracellular high pH causes store depletion, thereby activating ICRAC [104]. Though, in our results, a change of extracellular pH from 7.4 to 7.2 greatly improved CEU assembly and SOCE activation (Figures 5 and 7).

### 3.4. Final Remarks

The present work aimed to investigate which conditions could influence the formation of functional CEUs. Our structural and functional analyses demonstrate that CEU assembly (i) is a process controlled by the muscle itself, as it does not require blood supply and innervation, as demonstrated by the fact that they do form also when neural and vascular components are removed away from the skeletal muscle; and (ii) is favored by physiological temperatures and acidification of extracellular pH, as demonstrated by manipulating the extracellular milieu surrounding the isolated muscle during ex vivo contractility experiments.

Many other parameters that deserve investigation may influence and modulate the remodeling of SR and TT during exercise. For example, STIM1 was shown to be directly activated via S-glutathionylation under conditions of oxidative stress, suggesting



that STIM1 is a redox sensor. Hence, STIM1 could function as a polymodal sensor of temperature, ER  $\text{Ca}^{2+}$ , and oxidative stress [105]. Several reactions occurring in muscle during exercise change the intracellular micro-environment, as (a) decrease in ATP/ADP ratio and increase in levels of inorganic phosphate; (b) repetitive activation-relaxation cycles of contractile machinery, which activate mechano-sensors in contractile filaments and cytoskeleton [106,107].

These events may also play a role in membrane remodeling during CEU assembly, even though we have not yet tested this hypothesis. See also Protasi et al., 2021 [83], for additional discussion of the role that proteins involved in TT biogenesis and membrane-bending may play [108–111].

## 4. Materials and Methods

### 4.1. Animals

All experiments were conducted according to the Directive of the European Union 2010/63/UE and were approved by the Animal Ethical Committee of the University of Chieti-Pescara and by Italian Ministry of Health (n. 1202/2020-PR).

Wild type (WT) C57bl/6 male mice were housed in microisolator cages at 20 °C in a 12 h light/dark cycle and provided free access to standard food and water. All animals were sacrificed by cervical dislocation at 4 months of age, as approved by the Italian D. lgs. n.26/2014.

### 4.2. In Vitro Experiments

Extensor digitorum longus (EDL) muscles were excised from euthanized 4-month-old male WT mice and subjected to *in vitro* contraction experiments using the Aurora Muscle Physiology System (1200A: Isolated Muscle System, Aurora Scientific, ON, Canada). Intact excised EDL muscles were attached to a servo motor and force transducer (model 1200A, Aurora Scientific, ON, Canada) and stimulated using two platinum electrodes in a chamber continuously perfused with Ringer's solution as previously described [77]. Before starting each experiment, optimal stimulation and muscle length (L0) were determined using a series of 1 Hz twitch stimulation trains while stretching the muscle to a length that generated maximal force (F0). After establishing L0, muscles were first equilibrated using three tetani (0.5 s, 150 Hz) given at 1 min intervals and then *ex vivo* exercised to induce CEU assembly, as follows.

- *Ex vivo* exercise protocol (*Ex. ex vivo*). In order to evaluate the role of temperature and pH in the *ex vivo* assembly of CEUs induced by the incremental fatigue protocol, the above-mentioned protocol was carried out using Ringer's solution at different temperatures (25 °C, 30 °C, or 36 °C) and two different pH levels (7.4 or 7.2). The experimental protocol consisted of 4 steps of tetanic stimulus train: (i) 25 consecutive 0.5 s stimulus trains at 80 Hz of frequency applied every 25 s; (ii) 35 consecutive 0.5 s stimulus trains at 80 Hz of frequency applied every 20 s; (iii) 45 consecutive 0.5 s stimulus trains at 80 Hz of frequency applied every 15 s; (iv) 55 consecutive 0.5 s stimulus trains at 80 Hz of frequency applied every 10 s. EDL muscles were then directly intended for electron microscopy (EM) or subjected to an *ex vivo* functional test.
- *Ex vivo* functional test. After the *ex vivo* exercise protocol, muscles were kept in Ringer's solution at 25 °C, pH 7.4, for 30 min to allow muscle recovery. For a schematic representation of the *ex vivo* procedures, see Figure 1. Then, EDL muscles were subjected to an *ex vivo* functional test consisting of 30 consecutive stimulus trains at 60 Hz of frequency (each pulse having a duration of 1 s) applied every 5 s. To determine the relative contribution of extracellular  $\text{Ca}^{2+}$  entry, experiments were conducted also in the presence of 10  $\mu\text{M}$  BTP-2, an established inhibitor of SOCE [90], or in the absence of external  $\text{Ca}^{2+}$ . Specific force ( $\text{mN}/\text{mm}^2$ ) was calculated by normalizing the absolute force (mN) to the cross-sectional area (CSA,  $\text{mm}^2$ ) obtained as the following formula: muscle wet weight (mg)/L0 (mm)  $\times$  1.06 ( $\text{mg}/\text{mm}^3$ ).

#### 4.3. Immunofluorescence Labeling and Confocal Microscopy (CM)

EDL muscles were dissected from sacrificed mice, mounted in the above described system for ex vivo experiments, and subjected to the ex vivo exercise protocol or not (control muscles) at 30 °C. EDLs were then fixed in 2% paraformaldehyde in phosphate buffered saline (PBS) for 20 min at room temperature (RT). Small bundles of fixed fibers were (a) permeabilized for 30 min in 10% goat serum and 0.5% Triton X-100 PBS/BSA solution; (b) blocked for 1 h in PBS containing 10% goat serum; (c) washed 3 times for 10 min in PBS/BSA 1% solution; (d) incubated overnight at 4 °C in primary antibody diluted in PBS/BSA 1%; washed 3 times in PBS; (e) incubated with the secondary antibody for 1 h at RT; and (f) washed 3 times in PBS/BSA 1% solution before being mounted on coverslips with anti-bleach media. Primary antibodies used (a) mouse monoclonal anti-RyR1/RyR3 (34C antibody, 1:30, Developmental Studies Hybridoma Bank, IA, USA); (b) rabbit polyclonal anti-stromal-interacting molecule-1 (STIM1) (1:100, Sigma Aldrich, St. Louis, OH, USA); and (c) rabbit polyclonal anti-Orai1, (1:20, Thermo Scientific, Waltham, MA, USA). Secondary antibodies used (a) Cy5-labeled goat anti-mouse IgG (1:50); or (b) Cy3-labeled goat anti-rabbit (1:200) [112]. All secondary antibodies were from Jackson ImmunoResearch Laboratories (West Grove, PA, USA). Specimens were viewed and imaged using a scanning laser confocal microscope (LSM 800 Carl Zeiss, Germany) interfaced with an inverted Zeiss Axio Observer microscope. Fluorescence image profiles and co-localization were obtained from ZEN blue image analysis software (Carl Zeiss, Germany).

#### 4.4. Preparation of Samples for EM

EDL muscles for ultrastructural analysis were prepared for EM after being subjected to invitroexperiments (see above). Muscles were pinned on Sylgard dishes and fixed at RT in 3.5% glutaraldehyde in 0.1 M sodium cacodylate (NaCaCO) buffer (pH 7.2) and stored in the fixative solution at 4 °C until the embedding procedure. Fixed muscles were then post-fixed, stained en-block, and embedded in epoxy resin as previously described [112,113]. Briefly, for standard EM analysis, fixed muscle samples were post-fixed for 1–2 h in 2% OsO<sub>4</sub>. For transverse tubule (TT) staining in EM, specimens were post-fixed in a mixture of 2% OsO<sub>4</sub> and 0.8% potassium ferrocyanide (K<sub>3</sub>Fe(CN)<sub>6</sub>) for 1–2 h followed by a rinse with 0.1 M NaCaCO buffer with 75 mM CaCl<sub>2</sub> and then further processed. Potassium ferrocyanide precipitate within the TT network is visualized as an electron-dense dark precipitate in EM images [77].

EM ultra-thin sections (~50 nm of thickness) were cut from embedded samples (either with standard protocol or with TT staining) using a Leica Ultracut R microtome (Leica Microsystem, Wien, Austria) with a 45° Diatome Ultra diamond knife (Diatome, Biel, Switzerland) and stained with uranyl acetate replacement and lead citrate. Sections were viewed at 60 kV using a FP 505 Morgagni series 268D transmission electron microscope (FEI Company, Brno, Czech Republic), equipped with a Megaview III digital camera (Olympus Soft Imaging Solutions, Munster, Germany) and Soft Imaging System.

#### 4.5. Quantitative Analysis by EM

For all quantitative EM analyses, micrographs of non-overlapping regions were randomly collected from transverse sections of internal areas of fast-twitch EDL muscle fibers, as described previously [77]. The following ultrastructural parameters were evaluated:

- *SR stacks*. Incidence of fibers presenting SR stacks (expressed as percentage) and number of SR stacks (per 100 μm<sup>2</sup> of section) were determined in micrographs collected from EDL muscle fibers in transverse sections. In each specimen, 15–20 fibers were analyzed, and in each fiber, 5 micrographs were taken at 28,000× magnification.
- *Non-triadic TT network at the I band*. We determined both (i) the extension of the SR in close association with the TT, and (ii) the total network of the TT at the I band of sarcomere. The non-triadic TT network was evaluated in micrographs collected from EDL muscle fibers either stained or not with ferrocyanide in transverse sections and

reported as average number per area of section ( $100 \mu\text{m}^2$ ). In each specimen, 15–20 fibers were analyzed, and in each fiber, 5 micrographs were taken at  $28,000\times$  magnification.

#### 4.6. Statistical Analyses

Statistical analyses were determined using PRISM 9 (GraphPad Software, San Diego, CA, USA) and Microsoft Excel (Microsoft Office, Redmond, WA, USA). Significance was evaluated using Chi-square, ANOVA, and t-tests for EM analysis. The significance of ex vivo experiments was evaluated using two-way ANOVA followed by Tukey's post hoc test for pairwise comparisons of more than two groups or multiple *t* tests followed by Tukey's post hoc test. In all cases, data are shown as mean  $\pm$  SEM and differences are considered statistically significant at  $p < 0.05$ .

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms24065328/s1>.

**Author Contributions:** F.P. and L.P. conceived and directed the study. A.D.F. prepared, embedded and sectioned samples for EM. B.G., M.S. and C.P. performed the experimental work and data analysis. In detail: (A) B.G. and M.S. performed qualitative and quantitative analysis of immunofluorescence in Figure 2; (B) L.P. supervised the EM work of B.G., who performed qualitative and quantitative analysis of Figures 3–5; (C) C.P. supervised the work of M.S., who performed functional ex vivo experiments and analysis of Figures 2, 6 and 7. Finally, B.G., M.S. and L.P. contributed to writing and editing the manuscript, under the final supervision of F.P. All authors have read and agreed to the published version of the manuscript.

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**Institutional Review Board Statement:** The animal study protocol was conducted according to the Directive of the European Union 2010/63/UE and was approved by the Animal Ethical Committee of the Univ. G. d'Annunzio of Chieti-Pescara and by Italian Ministry of Health (n. 1202/2020-PR).

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

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#### Abbreviations

CEU:  $\text{Ca}^{2+}$  Entry Unit; EC coupling: excitation-contraction coupling; EDL: extensor digitorum longus; EM: electron microscopy; SR: sarcoplasmic reticulum; STIM1: stromal interaction molecule-1; SOCE: store-operated  $\text{Ca}^{2+}$  entry; TT: transverse tubule; WT: wild type.

#### References

1. Bers, D.M. Cardiac excitation-contraction coupling. *Nature* **2002**, *415*, 198–205. [[CrossRef](#)] [[PubMed](#)]
2. Calderon, J.C.; Bolanos, P.; Caputo, C. The excitation-contraction coupling mechanism in skeletal muscle. *Biophys. Rev.* **2014**, *6*, 133–160. [[CrossRef](#)] [[PubMed](#)]
3. Fabiato, A. Calcium-induced release of calcium from the cardiac sarcoplasmic reticulum. *Am. J. Physiol.* **1983**, *245*, C1–C14. [[CrossRef](#)] [[PubMed](#)]
4. Rios, E.; Pizarro, G. Voltage sensor of excitation-contraction coupling in skeletal muscle. *Physiol. Rev.* **1991**, *71*, 849–908. [[CrossRef](#)] [[PubMed](#)]
5. Schneider, M.F. Control of calcium release in functioning skeletal muscle fibers. *Annu. Rev. Physiol.* **1994**, *56*, 463–484. [[CrossRef](#)]
6. Schneider, M.F.; Chandler, W.K. Voltage dependent charge movement of skeletal muscle: A possible step in excitation-contraction coupling. *Nature* **1973**, *242*, 244–246. [[CrossRef](#)]
7. Rios, E.; Brum, G. Involvement of dihydropyridine receptors in excitation-contraction coupling in skeletal muscle. *Nature* **1987**, *325*, 717–720. [[CrossRef](#)]
8. Franzini-Armstrong, C.; Protasi, F. Ryanodine receptors of striated muscles: A complex channel capable of multiple interactions. *Physiol. Rev.* **1997**, *77*, 699–729. [[CrossRef](#)]
9. Sorrentino, V. The ryanodine receptor family of intracellular calcium release channels. *Adv. Pharmacol.* **1995**, *33*, 67–90.
10. Sorrentino, V.; Volpe, P. Ryanodine receptors: How many, where and why? *Trends Pharm. Sci.* **1993**, *14*, 98–103. [[CrossRef](#)]



11. Franzini-Armstrong, C. The sarcoplasmic reticulum and the control of muscle contraction. *FASEB J.* **1999**, *13* (Suppl. S2), S266–S270. [[CrossRef](#)]
12. Franzini-Armstrong, C.; Jorgensen, A.O. Structure and development of E-C coupling units in skeletal muscle. *Annu. Rev. Physiol.* **1994**, *56*, 509–534. [[CrossRef](#)]
13. Cherednichenko, G.; Ward, C.W.; Feng, W.; Cabrales, E.; Michaelson, L.; Samsó, M.; Lopez, J.R.; Allen, P.D.; Pessah, I.N. Enhanced excitation-coupled calcium entry in myotubes expressing malignant hyperthermia mutation R163C is attenuated by dantrolene. *Mol. Pharmacol.* **2008**, *73*, 1203–1212. [[CrossRef](#)] [[PubMed](#)]
14. Cherednichenko, G.; Zima, A.V.; Feng, W.; Schaefer, S.; Blatter, L.A.; Pessah, I.N. NADH oxidase activity of rat cardiac sarcoplasmic reticulum regulates calcium-induced calcium release. *Circ. Res.* **2004**, *94*, 478–486. [[CrossRef](#)] [[PubMed](#)]
15. Hurne, A.M.; O'Brien, J.J.; Wingrove, D.; Cherednichenko, G.; Allen, P.D.; Beam, K.G.; Pessah, I.N. Ryanodine receptor type 1 (RyR1) mutations C4958S and C4961S reveal excitation-coupled calcium entry (ECCE) is independent of sarcoplasmic reticulum store depletion. *J. Biol. Chem.* **2005**, *280*, 36994–37004. [[CrossRef](#)] [[PubMed](#)]
16. Yang, T.; Allen, P.D.; Pessah, I.N.; Lopez, J.R. Enhanced excitation-coupled calcium entry in myotubes is associated with expression of RyR1 malignant hyperthermia mutations. *J. Biol. Chem.* **2007**, *282*, 37471–37478. [[CrossRef](#)]
17. Bannister, R.A.; Pessah, I.N.; Beam, K.G. The skeletal L-type  $\text{Ca}^{2+}$  current is a major contributor to excitation-coupled  $\text{Ca}^{2+}$  entry. *J. Gen. Physiol.* **2009**, *133*, 79–91. [[CrossRef](#)]
18. Cherednichenko, G.; Hurne, A.M.; Fessenden, J.D.; Lee, E.H.; Allen, P.D.; Beam, K.G.; Pessah, I.N. Conformational activation of  $\text{Ca}^{2+}$  entry by depolarization of skeletal myotubes. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 15793–15798. [[CrossRef](#)]
19. Lu, T.; Zhang, Y.; Su, Y.; Zhou, D.; Xu, Q. Role of store-operated  $\text{Ca}^{2+}$  entry in cardiovascular disease. *Cell Commun. Signal.* **2022**, *20*, 33. [[CrossRef](#)]
20. Parekh, A.B.; Penner, R. Store depletion and calcium influx. *Physiol. Rev.* **1997**, *77*, 901–930. [[CrossRef](#)]
21. Putney, J.W., Jr. A model for receptor-regulated calcium entry. *Cell Calcium* **1986**, *7*, 1–12. [[CrossRef](#)] [[PubMed](#)]
22. Shen, Y.; Thillaiappan, N.B.; Taylor, C.W. The store-operated  $\text{Ca}^{2+}$  entry complex comprises a small cluster of STIM1 associated with one Orai1 channel. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2010789118. [[CrossRef](#)]
23. Kurebayashi, N.; Ogawa, Y. Depletion of  $\text{Ca}^{2+}$  in the sarcoplasmic reticulum stimulates  $\text{Ca}^{2+}$  entry into mouse skeletal muscle fibres. *J. Physiol.* **2001**, *533*, 185–199. [[CrossRef](#)]
24. Feske, S.; Gwack, Y.; Prakriya, M.; Srikanth, S.; Puppel, S.H.; Tanasa, B.; Hogan, P.G.; Lewis, R.S.; Daly, M.; Rao, A. A mutation in Orai1 causes immune deficiency by abrogating CRAC channel function. *Nature* **2006**, *441*, 179–185. [[CrossRef](#)]
25. Dirksen, R.T. Checking your SOCCs and feet: The molecular mechanisms of  $\text{Ca}^{2+}$  entry in skeletal muscle. *J. Physiol.* **2009**, *587*, 3139–3147. [[CrossRef](#)]
26. Lewis, R.S. The molecular choreography of a store-operated calcium channel. *Nature* **2007**, *446*, 284–287. [[CrossRef](#)]
27. Liou, J.; Fivaz, M.; Inoue, T.; Meyer, T. Live-cell imaging reveals sequential oligomerization and local plasma membrane targeting of stromal interaction molecule 1 after  $\text{Ca}^{2+}$  store depletion. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 9301–9306. [[CrossRef](#)]
28. Liou, J.; Kim, M.L.; Heo, W.D.; Jones, J.T.; Myers, J.W.; Ferrell, J.E., Jr.; Meyer, T. STIM is a  $\text{Ca}^{2+}$  sensor essential for  $\text{Ca}^{2+}$ -store-depletion-triggered  $\text{Ca}^{2+}$  influx. *Curr. Biol.* **2005**, *15*, 1235–1241. [[CrossRef](#)]
29. Luik, R.M.; Wang, B.; Prakriya, M.; Wu, M.M.; Lewis, R.S. Oligomerization of STIM1 couples ER calcium depletion to CRAC channel activation. *Nature* **2008**, *454*, 538–542. [[CrossRef](#)]
30. Lyfenko, A.D.; Dirksen, R.T. Differential dependence of store-operated and excitation-coupled  $\text{Ca}^{2+}$  entry in skeletal muscle on STIM1 and Orai1. *J. Physiol.* **2008**, *586*, 4815–4824. [[CrossRef](#)]
31. Roos, J.; DiGregorio, P.J.; Yeromin, A.V.; Ohlsen, K.; Lioudyno, M.; Zhang, S.; Safrina, O.; Kozak, J.A.; Wagner, S.L.; Cahalan, M.D.; et al. STIM1, an essential and conserved component of store-operated  $\text{Ca}^{2+}$  channel function. *J. Cell Biol.* **2005**, *169*, 435–445. [[CrossRef](#)]
32. Vig, M.; Peinelt, C.; Beck, A.; Koomoa, D.L.; Rabah, D.; Koblan-Huberson, M.; Kraft, S.; Turner, H.; Fleig, A.; Penner, R.; et al. CRACM1 is a plasma membrane protein essential for store-operated  $\text{Ca}^{2+}$  entry. *Science* **2006**, *312*, 1220–1223. [[CrossRef](#)]
33. Wu, M.M.; Buchanan, J.; Luik, R.M.; Lewis, R.S.  $\text{Ca}^{2+}$  store depletion causes STIM1 to accumulate in ER regions closely associated with the plasma membrane. *J. Cell Biol.* **2006**, *174*, 803–813. [[CrossRef](#)] [[PubMed](#)]
34. Zhang, S.L.; Yu, Y.; Roos, J.; Kozak, J.A.; Deerinck, T.J.; Ellisman, M.H.; Stauderman, K.A.; Cahalan, M.D. STIM1 is a  $\text{Ca}^{2+}$  sensor that activates CRAC channels and migrates from the  $\text{Ca}^{2+}$  store to the plasma membrane. *Nature* **2005**, *437*, 902–905. [[CrossRef](#)] [[PubMed](#)]
35. Wu, M.M.; Covington, E.D.; Lewis, R.S. Single-molecule analysis of diffusion and trapping of STIM1 and Orai1 at endoplasmic reticulum-plasma membrane junctions. *Mol. Biol. Cell* **2014**, *25*, 3672–3685. [[CrossRef](#)]
36. Luik, R.M.; Wu, M.M.; Buchanan, J.; Lewis, R.S. The elementary unit of store-operated  $\text{Ca}^{2+}$  entry: Local activation of CRAC channels by STIM1 at ER-plasma membrane junctions. *J. Cell Biol.* **2006**, *174*, 815–825. [[CrossRef](#)]
37. Zhang, H.; Bryson, V.G.; Wang, C.; Li, T.; Kerr, J.P.; Wilson, R.; Muoio, D.M.; Bloch, R.J.; Ward, C.; Rosenberg, P.B. Desmin interacts with STIM1 and coordinates  $\text{Ca}^{2+}$  signaling in skeletal muscle. *JCI Insight* **2021**, *6*, e143472. [[CrossRef](#)]
38. Zhou, Y.; Cai, X.; Nwokonko, R.M.; Loktionova, N.A.; Wang, Y.; Gill, D.L. The STIM-Orai coupling interface and gating of the Orai1 channel. *Cell Calcium* **2017**, *63*, 8–13. [[CrossRef](#)]
39. Stiber, J.; Hawkins, A.; Zhang, Z.S.; Wang, S.; Burch, J.; Graham, V.; Ward, C.C.; Seth, M.; Finch, E.; Malouf, N.; et al. STIM1 signalling controls store-operated calcium entry required for development and contractile function in skeletal muscle. *Nat. Cell Biol.* **2008**, *10*, 688–697. [[CrossRef](#)]

40. Carrell, E.M.; Coppola, A.R.; McBride, H.J.; Dirksen, R.T. Orai1 enhances muscle endurance by promoting fatigue-resistant type I fiber content but not through acute store-operated  $\text{Ca}^{2+}$  entry. *FASEB J.* **2016**, *30*, 4109–4119. [[CrossRef](#)]
41. Wei-Lapierre, L.; Carrell, E.M.; Boncompagni, S.; Protasi, F.; Dirksen, R.T. Orai1-dependent calcium entry promotes skeletal muscle growth and limits fatigue. *Nat. Commun.* **2013**, *4*, 2805. [[CrossRef](#)] [[PubMed](#)]
42. Conte, E.; Imbrici, P.; Mantuano, P.; Coppola, M.A.; Camerino, G.M.; De Luca, A.; Liantonio, A. Alteration of STIM1/Orai1-Mediated SOCE in Skeletal Muscle: Impact in Genetic Muscle Diseases and Beyond. *Cells* **2021**, *10*, 2722. [[CrossRef](#)]
43. Yen, M.; Lewis, R.S. Numbers count: How STIM and Orai stoichiometry affect store-operated calcium entry. *Cell Calcium* **2019**, *79*, 35–43. [[CrossRef](#)] [[PubMed](#)]
44. Hoover, P.J.; Lewis, R.S. Stoichiometric requirements for trapping and gating of  $\text{Ca}^{2+}$  release-activated  $\text{Ca}^{2+}$  (CRAC) channels by stromal interaction molecule 1 (STIM1). *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 13299–13304. [[CrossRef](#)]
45. Stathopoulos, P.B.; Schindl, R.; Fahrner, M.; Zheng, L.; Gasmi-Seabrook, G.M.; Muik, M.; Romanin, C.; Ikura, M. STIM1/Orai1 coiled-coil interplay in the regulation of store-operated calcium entry. *Nat. Commun.* **2013**, *4*, 2963. [[CrossRef](#)]
46. Zhou, Y.; Wang, X.; Loktionova, N.A.; Cai, X.; Nwokonko, R.M.; Vrana, E.; Wang, Y.; Rothberg, B.S.; Gill, D.L. STIM1 dimers undergo unimolecular coupling to activate Orai1 channels. *Nat. Commun.* **2015**, *6*, 8395. [[CrossRef](#)]
47. Zhou, Y.; Nwokonko, R.M.; Cai, X.; Loktionova, N.A.; Abdulqadir, R.; Xin, P.; Niemeyer, B.A.; Wang, Y.; Trebak, M.; Gill, D.L. Cross-linking of Orai1 channels by STIM proteins. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E3398–E3407. [[CrossRef](#)]
48. Sztretze, M.; Geyer, N.; Vincze, J.; Al-Gaadi, D.; Olah, T.; Szentesi, P.; Kis, G.; Antal, M.; Balatoni, I.; Csernoch, L.; et al. SOCE Is Important for Maintaining Sarcoplasmic Calcium Content and Release in Skeletal Muscle Fibers. *Biophys. J.* **2017**, *113*, 2496–2507. [[CrossRef](#)]
49. Brotto, M. Aging, sarcopenia and store-operated calcium entry: A common link? *Cell Cycle* **2011**, *10*, 4201–4202. [[CrossRef](#)]
50. Pan, Z.; Brotto, M.; Ma, J. Store-operated  $\text{Ca}^{2+}$  entry in muscle physiology and diseases. *BMB Rep.* **2014**, *47*, 69–79. [[CrossRef](#)]
51. Thornton, A.M.; Zhao, X.; Weisleder, N.; Brotto, L.S.; Bougoin, S.; Nosek, T.M.; Reid, M.; Hardin, B.; Pan, Z.; Ma, J.; et al. Store-operated  $\text{Ca}^{2+}$  entry (SOCE) contributes to normal skeletal muscle contractility in young but not in aged skeletal muscle. *Aging* **2011**, *3*, 621–634. [[CrossRef](#)] [[PubMed](#)]
52. Yarotsky, V.; Dirksen, R.T. Temperature and RyR1 regulate the activation rate of store-operated  $\text{Ca}^{2+}$  entry current in myotubes. *Biophys. J.* **2012**, *103*, 202–211. [[CrossRef](#)]
53. Yarotsky, V.; Protasi, F.; Dirksen, R.T. Accelerated activation of SOCE current in myotubes from two mouse models of anesthetic- and heat-induced sudden death. *PLoS ONE* **2013**, *8*, e77633. [[CrossRef](#)]
54. Garcia-Castaneda, M.; Michelucci, A.; Zhao, N.; Malik, S.; Dirksen, R.T. Postdevelopmental knockout of Orai1 improves muscle pathology in a mouse model of Duchenne muscular dystrophy. *J. Gen. Physiol.* **2022**, *154*, e202213081. [[CrossRef](#)]
55. Goonasekera, S.A.; Davis, J.; Kwong, J.Q.; Accornero, F.; Wei-LaPierre, L.; Sargent, M.A.; Dirksen, R.T.; Molkenin, J.D. Enhanced  $\text{Ca}^{2+}$  influx from STIM1-Orai1 induces muscle pathology in mouse models of muscular dystrophy. *Hum. Mol. Genet.* **2014**, *23*, 3706–3715. [[CrossRef](#)]
56. Onopiuk, M.; Brutkowsky, W.; Young, C.; Krasowska, E.; Rog, J.; Ritso, M.; Wojciechowska, S.; Arkle, S.; Zablocki, K.; Gorecki, D.C. Store-operated calcium entry contributes to abnormal  $\text{Ca}^{2+}$  signalling in dystrophic mdx mouse myoblasts. *Arch. Biochem. Biophys.* **2015**, *569*, 1–9. [[CrossRef](#)]
57. Zhao, X.; Moloughney, J.G.; Zhang, S.; Komazaki, S.; Weisleder, N. Orai1 mediates exacerbated  $\text{Ca}^{2+}$  entry in dystrophic skeletal muscle. *PLoS ONE* **2012**, *7*, e49862. [[CrossRef](#)]
58. Bohm, J.; Bulla, M.; Urquhart, J.E.; Malfatti, E.; Williams, S.G.; O'Sullivan, J.; Szlauer, A.; Koch, C.; Baranello, G.; Mora, M.; et al. ORAI1 Mutations with Distinct Channel Gating Defects in Tubular Aggregate Myopathy. *Hum. Mutat.* **2017**, *38*, 426–438. [[CrossRef](#)]
59. Bohm, J.; Chevessier, F.; Maues De Paula, A.; Koch, C.; Attarian, S.; Feger, C.; Hantai, D.; Laforet, P.; Ghorab, K.; Vallat, J.M.; et al. Constitutive activation of the calcium sensor STIM1 causes tubular-aggregate myopathy. *Am. J. Hum. Genet.* **2013**, *92*, 271–278. [[CrossRef](#)]
60. Bohm, J.; Laporte, J. Gain-of-function mutations in STIM1 and ORAI1 causing tubular aggregate myopathy and Stormorken syndrome. *Cell Calcium* **2018**, *76*, 1–9. [[CrossRef](#)]
61. Endo, Y.; Noguchi, S.; Hara, Y.; Hayashi, Y.K.; Motomura, K.; Miyatake, S.; Murakami, N.; Tanaka, S.; Yamashita, S.; Kizu, R.; et al. Dominant mutations in ORAI1 cause tubular aggregate myopathy with hypocalcemia via constitutive activation of store-operated  $\text{Ca}^{2+}$  channels. *Hum. Mol. Genet.* **2015**, *24*, 637–648. [[CrossRef](#)] [[PubMed](#)]
62. Garibaldi, M.; Fattori, F.; Riva, B.; Labasse, C.; Brochier, G.; Ottaviani, P.; Sacconi, S.; Vizzaccaro, E.; Laschena, F.; Romero, N.B.; et al. A novel gain-of-function mutation in ORAI1 causes late-onset tubular aggregate myopathy and congenital miosis. *Clin. Genet.* **2017**, *91*, 780–786. [[CrossRef](#)] [[PubMed](#)]
63. Nesin, V.; Wiley, G.; Kousi, M.; Ong, E.C.; Lehmann, T.; Nicholl, D.J.; Suri, M.; Shahrizaila, N.; Katsanis, N.; Gaffney, P.M.; et al. Activating mutations in STIM1 and ORAI1 cause overlapping syndromes of tubular myopathy and congenital miosis. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 4197–4202. [[CrossRef](#)] [[PubMed](#)]
64. Protasi, F.; Girolami, B.; Roccabianca, S.; Rossi, D. Store-operated calcium entry: From physiology to tubular aggregate myopathy. *Curr. Opin. Pharm.* **2023**, *68*, 102347. [[CrossRef](#)] [[PubMed](#)]
65. Sallinger, M.; Tiffner, A.; Schmidt, T.; Bonhenry, D.; Waldherr, L.; Frischauf, I.; Lunz, V.; Derler, I.; Schober, R.; Schindl, R. Luminal STIM1 Mutants that Cause Tubular Aggregate Myopathy Promote Autophagic Processes. *Int. J. Mol. Sci.* **2020**, *21*, 4410. [[CrossRef](#)]

66. Cully, T.R.; Edwards, J.N.; Murphy, R.M.; Launikonis, B.S. A quantitative description of tubular system Ca(2+) handling in fast- and slow-twitch muscle fibres. *J. Physiol.* **2016**, *594*, 2795–2810. [[CrossRef](#)]
67. Koenig, X.; Choi, R.H.; Launikonis, B.S. Store-operated Ca(2+) entry is activated by every action potential in skeletal muscle. *Commun. Biol.* **2018**, *1*, 31. [[CrossRef](#)]
68. Pearce, L.; Meizoso-Huesca, A.; Seng, C.; Lamboley, C.R.; Singh, D.P.; Launikonis, B.S. Ryanodine receptor activity and store-operated Ca(2+) entry: Critical regulators of Ca(2+) content and function in skeletal muscle. *J. Physiol.* **2022**. [[CrossRef](#)]
69. Lilliu, E.; Hilber, K.; Launikonis, B.S.; Koenig, X. Phasic Store-Operated Ca(2+) Entry During Excitation-Contraction Coupling in Skeletal Muscle Fibers From Exercised Mice. *Front. Physiol.* **2020**, *11*, 597647. [[CrossRef](#)]
70. Koenig, X.; Choi, R.H.; Schicker, K.; Singh, D.P.; Hilber, K.; Launikonis, B.S. Mechanistic insights into store-operated Ca(2+) entry during excitation-contraction coupling in skeletal muscle. *Biochim. Biophys. Acta Mol. Cell Res.* **2019**, *1866*, 1239–1248. [[CrossRef](#)]
71. Edwards, J.N.; Blackmore, D.G.; Gilbert, D.F.; Murphy, R.M.; Launikonis, B.S. Store-operated calcium entry remains fully functional in aged mouse skeletal muscle despite a decline in STIM1 protein expression. *Aging Cell* **2011**, *10*, 675–685. [[CrossRef](#)]
72. Launikonis, B.S.; Rios, E. Store-operated Ca2+ entry during intracellular Ca2+ release in mammalian skeletal muscle. *J. Physiol.* **2007**, *583*, 81–97. [[CrossRef](#)]
73. Launikonis, B.S.; Stephenson, D.G.; Friedrich, O. Rapid Ca2+ flux through the transverse tubular membrane, activated by individual action potentials in mammalian skeletal muscle. *J. Physiol.* **2009**, *587 Pt 10*, 2299–2312. [[CrossRef](#)]
74. Darbellay, B.; Arnaudeau, S.; Bader, C.R.; Konig, S.; Bernheim, L. STIM1L is a new actin-binding splice variant involved in fast repetitive Ca2+ release. *J. Cell Biol.* **2011**, *194*, 335–346. [[CrossRef](#)]
75. Darbellay, B.; Arnaudeau, S.; Konig, S.; Jousset, H.; Bader, C.; Demaurex, N.; Bernheim, L. STIM1- and Orai1-dependent store-operated calcium entry regulates human myoblast differentiation. *J. Biol. Chem.* **2009**, *284*, 5370–5380. [[CrossRef](#)]
76. Sztretye, M.; Singlar, Z.; Balogh, N.; Kis, G.; Szentesi, P.; Angyal, A.; Balatoni, I.; Csernoch, L.; Dienes, B. The Role of Orai1 in Regulating Sarcoplasmic Calcium Release, Mitochondrial Morphology and Function in Myostatin Deficient Skeletal Muscle. *Front. Physiol.* **2020**, *11*, 601090. [[CrossRef](#)]
77. Boncompagni, S.; Michelucci, A.; Pietrangelo, L.; Dirksen, R.T.; Protasi, F. Exercise-dependent formation of new junctions that promote STIM1-Orai1 assembly in skeletal muscle. *Sci. Rep.* **2017**, *7*, 14286. [[CrossRef](#)]
78. Boncompagni, S.; Michelucci, A.; Pietrangelo, L.; Dirksen, R.T.; Protasi, F. Addendum: Exercise-dependent formation of new junctions that promote STIM1-Orai1 assembly in skeletal muscle. *Sci. Rep.* **2018**, *8*, 17463. [[CrossRef](#)]
79. Boncompagni, S.; Pecorai, C.; Michelucci, A.; Pietrangelo, L.; Protasi, F. Long-Term Exercise Reduces Formation of Tubular Aggregates and Promotes Maintenance of Ca(2+) Entry Units in Aged Muscle. *Front. Physiol.* **2021**, *11*, 601057. [[CrossRef](#)]
80. Melzer, W. ECC meets CEU-New focus on the backdoor for calcium ions in skeletal muscle cells. *J. Gen. Physiol.* **2020**, *152*, e202012679. [[CrossRef](#)]
81. Michelucci, A.; Boncompagni, S.; Pietrangelo, L.; Garcia-Castaneda, M.; Takano, T.; Malik, S.; Dirksen, R.T.; Protasi, F. Transverse tubule remodeling enhances Orai1-dependent Ca(2+) entry in skeletal muscle. *Elife* **2019**, *8*, e47576. [[CrossRef](#)] [[PubMed](#)]
82. Michelucci, A.; Boncompagni, S.; Pietrangelo, L.; Takano, T.; Protasi, F.; Dirksen, R.T. Pre-assembled Ca2+ entry units and constitutively active Ca2+ entry in skeletal muscle of calsequestrin-1 knockout mice. *J. Gen. Physiol.* **2020**, *152*, e202012617. [[CrossRef](#)] [[PubMed](#)]
83. Protasi, F.; Pietrangelo, L.; Boncompagni, S. Calcium entry units (CEUs): Perspectives in skeletal muscle function and disease. *J. Muscle Res. Cell Motil.* **2020**, *42*, 233–249. [[CrossRef](#)] [[PubMed](#)]
84. Canato, M.; Scorzeto, M.; Giacomello, M.; Protasi, F.; Reggiani, C.; Stienen, G.J. Massive alterations of sarcoplasmic reticulum free calcium in skeletal muscle fibers lacking calsequestrin revealed by a genetically encoded probe. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 22326–22331. [[CrossRef](#)]
85. Allen, D.G.; Lamb, G.D.; Westerblad, H. Skeletal muscle fatigue: Cellular mechanisms. *Physiol Rev.* **2008**, *88*, 287–332. [[CrossRef](#)]
86. Fitts, R.H. Cellular mechanisms of muscle fatigue. *Physiol. Rev.* **1994**, *74*, 49–94. [[CrossRef](#)]
87. Lamb, G.D. Excitation-contraction coupling and fatigue mechanisms in skeletal muscle: Studies with mechanically skinned fibres. *J. Muscle Res. Cell Motil.* **2002**, *23*, 81–91. [[CrossRef](#)]
88. Lamb, G.D. Mechanisms of excitation-contraction uncoupling relevant to activity-induced muscle fatigue. *Appl. Physiol. Nutr. Metab.* **2009**, *34*, 368–372. [[CrossRef](#)]
89. Westerblad, H.; Allen, D.G. Emerging roles of ROS/RNS in muscle function and fatigue. *Antioxid. Redox Signal.* **2011**, *15*, 2487–2499. [[CrossRef](#)]
90. Zitt, C.; Strauss, B.; Schwarz, E.C.; Spaeth, N.; Rast, G.; Hatzelmann, A.; Hoth, M. Potent inhibition of Ca2+ release-activated Ca2+ channels and T-lymphocyte activation by the pyrazole derivative BTP2. *J. Biol. Chem.* **2004**, *279*, 12427–12437. [[CrossRef](#)]
91. Bootman, M.D.; Collins, T.J.; Mackenzie, L.; Roderick, H.L.; Berridge, M.J.; Peppiatt, C.M. 2-aminoethoxydiphenyl borate (2-APB) is a reliable blocker of store-operated Ca2+ entry but an inconsistent inhibitor of InsP3-induced Ca2+ release. *FASEB J.* **2002**, *16*, 1145–1150. [[CrossRef](#)]
92. Girolami, B.; Serano, M.; Michelucci, A.; Pietrangelo, L.; Protasi, F. Store-Operated Ca(2+) Entry in Skeletal Muscle Contributes to the Increase in Body Temperature during Exertional Stress. *Int. J. Mol. Sci.* **2022**, *23*, 3772. [[CrossRef](#)]
93. Xiao, B.; Coste, B.; Mathur, J.; Patapoutian, A. Temperature-dependent STIM1 activation induces Ca(2+) influx and modulates gene expression. *Nat. Chem. Biol.* **2011**, *7*, 351–358. [[CrossRef](#)]



94. Kenny, G.P.; Reardon, F.D.; Zaleski, W.; Reardon, M.L.; Haman, F.; Ducharme, M.B. Muscle temperature transients before, during, and after exercise measured using an intramuscular multisensor probe. *J. Appl. Physiol.* **1985**, *94*, 2350–2357. [[CrossRef](#)]
95. Boron, W.F. Regulation of intracellular pH. *Adv. Physiol. Educ.* **2004**, *28*, 160–179. [[CrossRef](#)]
96. Denker, S.P.; Huang, D.C.; Orłowski, J.; Furthmayr, H.; Barber, D.L. Direct binding of the Na–H exchanger NHE1 to ERM proteins regulates the cortical cytoskeleton and cell shape independently of H<sup>(+)</sup> translocation. *Mol. Cell* **2000**, *6*, 1425–1436. [[CrossRef](#)]
97. Putney, L.K.; Barber, D.L. Na-H exchange-dependent increase in intracellular pH times G2/M entry and transition. *J. Biol. Chem.* **2003**, *278*, 44645–44649. [[CrossRef](#)]
98. Turchi, L.; Loubat, A.; Rochet, N.; Rossi, B.; Ponzio, G. Evidence for a direct correlation between c-Jun NH2 terminal kinase 1 activation, cyclin D2 expression, and G(1.1)/S phase transition in the murine hybridoma 7TD1 cells. *Exp. Cell Res.* **2000**, *261*, 220–228. [[CrossRef](#)]
99. Boussouf, A.; Gaillard, S. Intracellular pH changes during oligodendrocyte differentiation in primary culture. *J. Neurosci. Res.* **2000**, *59*, 731–739. [[CrossRef](#)]
100. Uzman, J.A.; Patil, S.; Uzgare, A.R.; Sater, A.K. The role of intracellular alkalinization in the establishment of anterior neural fate in *Xenopus*. *Dev. Biol.* **1998**, *193*, 10–20. [[CrossRef](#)]
101. Marumo, M.; Suehiro, A.; Kakishita, E.; Groschner, K.; Wakabayashi, I. Extracellular pH affects platelet aggregation associated with modulation of store-operated Ca<sup>(2+)</sup> entry. *Thromb. Res.* **2001**, *104*, 353–360. [[CrossRef](#)] [[PubMed](#)]
102. Street, D.; Bangsbo, J.; Juel, C. Interstitial pH in human skeletal muscle during and after dynamic graded exercise. *J. Physiol.* **2001**, *537*, 993–998. [[CrossRef](#)] [[PubMed](#)]
103. Tsujikawa, H.; Yu, A.S.; Xie, J.; Yue, Z.; Yang, W.; He, Y.; Yue, L. Identification of key amino acid residues responsible for internal and external pH sensitivity of Orai1/STIM1 channels. *Sci. Rep.* **2015**, *5*, 16747. [[CrossRef](#)] [[PubMed](#)]
104. Mancarella, S.; Wang, Y.; Deng, X.; Landesberg, G.; Scalia, R.; Panettieri, R.A.; Mallilankaraman, K.; Tang, X.D.; Madesh, M.; Gill, D.L. Hypoxia-induced acidosis uncouples the STIM-Orai calcium signaling complex. *J. Biol. Chem.* **2011**, *286*, 44788–44798. [[CrossRef](#)]
105. Hawkins, B.J.; Irrinki, K.M.; Mallilankaraman, K.; Lien, Y.C.; Wang, Y.; Bhanumathy, C.D.; Subbiah, R.; Ritchie, M.F.; Soboloff, J.; Baba, Y.; et al. S-glutathionylation activates STIM1 and alters mitochondrial homeostasis. *J. Cell Biol.* **2010**, *190*, 391–405. [[CrossRef](#)]
106. Piazzesi, G.; Caremani, M.; Linari, M.; Reconditi, M.; Lombardi, V. Thick Filament Mechano-Sensing in Skeletal and Cardiac Muscles: A Common Mechanism Able to Adapt the Energetic Cost of the Contraction to the Task. *Front. Physiol.* **2018**, *9*, 736. [[CrossRef](#)]
107. Ward, C.W.; Prosser, B.L.; Lederer, W.J. Mechanical stretch-induced activation of ROS/RNS signaling in striated muscle. *Antioxid. Redox Signal.* **2014**, *20*, 929–936. [[CrossRef](#)]
108. Al-Qusairi, L.; Laporte, J. T-tubule biogenesis and triad formation in skeletal muscle and implication in human diseases. *Skelet. Muscle* **2011**, *1*, 26. [[CrossRef](#)]
109. Dowling, J.J.; Vreede, A.P.; Low, S.E.; Gibbs, E.M.; Kuwada, J.Y.; Bonnemann, C.G.; Feldman, E.L. Loss of myotubularin function results in T-tubule disorganization in zebrafish and human myotubular myopathy. *PLoS Genet.* **2009**, *5*, e1000372. [[CrossRef](#)]
110. Lee, E.; Marcucci, M.; Daniell, L.; Pypaert, M.; Weisz, O.A.; Ochoa, G.C.; Farsad, K.; Wenk, M.R.; De Camilli, P. Amphiphysin 2 (Bin1) and T-tubule biogenesis in muscle. *Science* **2002**, *297*, 1193–1196. [[CrossRef](#)]
111. Takeshima, H.; Hoshijima, M.; Song, L.S. Ca<sup>(2+)</sup> microdomains organized by junctophilins. *Cell Calcium* **2015**, *58*, 349–356. [[CrossRef](#)]
112. Pietrangelo, L.; D’Incecco, A.; Ainbinder, A.; Michelucci, A.; Kern, H.; Dirksen, R.T.; Boncompagni, S.; Protasi, F. Age-dependent uncoupling of mitochondria from Ca<sup>(2+)</sup> release units in skeletal muscle. *Oncotarget* **2015**, *6*, 35358–35371. [[CrossRef](#)]
113. Pietrangelo, L.; Michelucci, A.; Ambrogini, P.; Sartini, S.; Guarnier, F.A.; Fusella, A.; Zamparo, I.; Mammucari, C.; Protasi, F.; Boncompagni, S. Muscle activity prevents the uncoupling of mitochondria from Ca<sup>(2+)</sup> Release Units induced by ageing and disuse. *Arch. Biochem. Biophys.* **2019**, *663*, 22–33. [[CrossRef](#)]

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