



Dual targeting of cancer-related human matrix metalloproteinases and carbonic anhydrases by chiral N-(biarylsulfonyl)-phosphonic acids

Grazia Luisi, Guido Angelini, Carla Gasbarri, Antonio Laghezza, Mariangela Agamennone, Fulvio Liodice, Claudiu T. Supuran, Cristina Campestre & Paolo Tortorella

To cite this article: Grazia Luisi, Guido Angelini, Carla Gasbarri, Antonio Laghezza, Mariangela Agamennone, Fulvio Liodice, Claudiu T. Supuran, Cristina Campestre & Paolo Tortorella (2017) Dual targeting of cancer-related human matrix metalloproteinases and carbonic anhydrases by chiral N-(biarylsulfonyl)-phosphonic acids, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 32:1, 1260-1264, DOI: [10.1080/14756366.2017.1378192](https://doi.org/10.1080/14756366.2017.1378192)

To link to this article: <http://dx.doi.org/10.1080/14756366.2017.1378192>



© 2017 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Published online: 26 Sep 2017.



Submit your article to this journal [↗](#)



View related articles [↗](#)




View Crossmark data [↗](#)

RESEARCH PAPER



Dual targeting of cancer-related human matrix metalloproteinases and carbonic anhydrases by chiral *N*-(biarylsulfonyl)-phosphonic acids

Grazia Luisi^a, Guido Angelini^a, Carla Gasbarri^a, Antonio Laghezza^b , Mariangela Agamennone^a, Fulvio Loiodice^b, Claudiu T. Supuran^c, Cristina Campestre^a and Paolo Tortorella^b

^aDepartment of Pharmacy, “G. d’Annunzio” University of Chieti-Pescara, Chieti, Italy; ^bDepartment of Pharmacy and Pharmaceutical Sciences, “A. Moro” University of Bari, Bari, Italy; ^cNeurofarba Department, Section of Pharmaceutical and Nutraceutical Sciences, University of Florence, Sesto Fiorentino (Florence), Italy

ABSTRACT

A series of nanomolar phosphonate matrix metalloproteinase (MPP) inhibitors was tested for inhibitory activity against a panel of selected human carbonic anhydrase (CA, EC 4.2.1.1) isozymes, covering the cancer-associated CA IX and XII. None of the reported sulfonyl and sulfonylamino-derivatives sensitively affected the catalytic activity of the cytosolic isoforms CA I and II, which are considered off-target isoforms in view of their physiological role. The most active inhibitors were in the series of chiral *N*-(sulfonyl)phosphoalane derivatives, which showed good to excellent inhibitory activity over target CAs, with compound **15** presenting the best isoform-selectivity toward CA IX. We suggest here that the phosphonates have the potential as dual inhibitors of MMPs and CAs, both involved in tumor formation, invasion and metastasis.

ARTICLE HISTORY

Received 27 July 2017
Revised 6 September 2017
Accepted 7 September 2017

KEYWORDS

Carbonic anhydrases; matrix metalloproteinases; phosphonic acids; sulfonamides; zinc-binding group

Introduction





Carbonic anhydrases (CAs, carbonate hydro-lyases, EC 4.2.1.1) are a large family of metalloenzymes that primarily act as efficient catalyzers of the reversible hydration of carbon dioxide to bicarbonate ions and protons^{1–3}. This biochemical reaction has been recognized a crucial role in regulating CO₂/electrolyte interchange at the respiratory and photosynthetic surfaces in eukaryotic as well prokaryotic organisms⁴, with a corollary of diverse physiological processes depending on this activity, including pH homeostasis, ion transport, bone dynamics, and energetic metabolism⁴.

It is widely accepted that a deregulation in CA function and expression is involved in the pathogenesis of important human diseases, among all glaucoma, epilepsy, osteoporosis, and cancer^{5–7}. A growing attention is focused on both CA IX and CA XII, since these trans-membrane isoenzymes have been reported to be overexpressed in many tumors, and associated with cancer progression and metastasis. In order to survive to the hypoxic stress, tumor cells execute an aberrant metabolic adjustment known as the Warburg effect⁸, which leads to acidification of the extracellular *milieu*; this harsh micro-environment creates selective survival conditions for solid tumors over normal tissues, by facilitating cancer invasiveness. The need to maintain the intracellular pH near physiological levels requires the activation of complex molecular mechanisms, such as the up-regulation of CA IX and CA XII. Therefore, these isoforms have become attractive targets for anti-cancer therapeutic purposes^{9,10}. It is worthy to note that, unlike CA XII, CA IX is minimally expressed in normal tissues. Furthermore, compared with isoform XII, CA IX is more prevalent in solid and metastatic tumors, the ones that more frequently

develop resistance to radiation and/or chemotherapy, probably because this isoform is able to retain its catalytic activity even in the acidic conditions of the tumor habitat.

Thus, the inhibition of this latter enzyme represents one of the most opportune strategy to sensitize highly aggressive tumors to the effects of antineoplastic drugs. From a general point of view, however, the single target approach in drug discovery often results in a poorer therapeutic impact if compared to the synergic effects obtainable with the modulation of multiple systems; thus, we were intrigued in the development of dual-acting ligands for cancer-related enzymes that, although belonging to different families, and despite the different mechanism of action, share significant commonalities in terms of molecular recognition. Over the years, we focused our attention on matrix metalloproteinases (MMPs), a family of zinc-dependent isoenzymes involved in bone homeostasis, extracellular matrix remodeling, and inflammation¹¹. Dysregulated MMPs, particularly MMP-2, have long been recognized to have a crucial role in tumor genesis and aggressiveness¹², giving rise to the challenging approach of inhibiting chosen MMPs for cancer treatment^{13,14}.

Recently, we capitalized on the observation that both MMP and α -class CA proteins depend upon the presence of a zinc ion in the catalytic domain to process substrates, as starting point to establish if the two enzymatic families could be inhibited by similar compounds. We reported a variety of potent and selective MMP inhibitors (MMPIs) based on the presence of a zinc-binding group (ZBG) coupled to a variable backbone able to fit into the S1' specificity pocket of the individual MMP active site^{15–17}. Due to its bidentate nature, the hydroxamic acid is one of the most strong chelating group for catalytic metal ions, and indeed the

CONTACT Claudiu T. Supuran  claudiu.supuran@unifi.it  Neurofarba Department, Section of Pharmaceutical and Nutraceutical Sciences, University of Florence, Via U. Schiff 6, Sesto Fiorentino (Florence), Italy; Cristina Campestre  cristina.campestre@unich.it  Department of Pharmacy, “G. d’Annunzio” University of Chieti-Pescara, Via dei Vestini 31, 66100 Chieti, Italy

© 2017 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

hydroxamate-based MMP inhibitors have been found the most active *in vitro*; however all attempts to obtain clinically successful drugs from inhibitors featuring this moiety have failed owing to common drawbacks, such as low MMP subtype selectivity, activity on a broad range of metal ions, including Fe(III), *in vivo* instability, dose limiting side effects and low oral availability^{18,19}. To address the main limitations of hydroxamates, alternative monodentate ZBGs such as carboxylate, thiolate, and phosphonate, that coordinate rather than chelate the zinc ion, may be used in metalloenzyme inhibitor design to obtain a gain in inhibitor selectivity; owing to the formation of less stable metal coordination complexes, in fact, a major contribute to the binding between inhibitor and protein is played by the distinctive substituents on the molecule scaffold²⁰.

This approach has been successfully exploited in our laboratories, leading to a series of (*R*)- α -biarylsulfonylamino-2-methyl-propyl phosphonates as MMPis characterized by high potency and enantioselectivity towards a panel of target isoforms;^{16,17} they represent by far the most powerful inhibitors containing the phosphonate moiety as ZBG, exhibiting IC₅₀ values in the nanomolar or even sub-nanomolar range. Our MMP-8 selective inhibitor (*R*)-1-[3'-methyl-biphenyl-4-sulfonylamino]-2-methylpropyl phosphonic acid has been shown to play an *in vivo* protective effect in experimental models of multiple sclerosis, systemic inflammatory response syndrome, and ventilator-induced lung injury, thus corroborating the link between MMP-8 up-regulation and systemic inflammatory diseases²¹⁻²³.

For the purpose to obtain further insights on this topic, we then investigated a small set of analogs of the reported α -arylsulfonylamino-methyl phosphonates, characterized by the replacement of the sulfonylamino with a sulfonyl group in alpha to the ZBG. Although most of the new α -arylsulfonyl-methyl phosphonates were found good to excellent inhibitors of selected MMPs, they resulted less active in comparison with their sulfonylamino counterparts; however, these phosphonates are notably more selective for MMP-2 over the anti-target isoforms MMP-3 and MMP-9²⁴.

We were then intrigued in evaluating a panel of compounds from these two chemical entities as CA inhibitors, with the aim to disclose a possible dual activity against MMPs and CAs. To the best of our knowledge, no data on dual MMP/CA inhibitors containing either sulfonylamino-methyl- or sulfonylmethyl-groups directly bound to a single phosphonate ZBG have been reported to date; in this context only related studies, disclosing sulfonylamino-methyl- or arylaminomethyl- bisphosphonates,^{25,26} and sulfonylated-aminoacyl-hydroxamates²⁷ as dual inhibitors, have been published by our team.

The α -arylsulfonylamino-methyl phosphonates under scrutiny (compounds **12-15**) contain a secondary sulfonamide moiety (-SO₂NH-), a distinctive feature of classical CAIs which bind as monodentate anions to the zinc ion in the active site, displacing the Zn²⁺-bound solvent molecule. Although multiple alternative inhibition mechanisms have been evidenced, as thoroughly detailed in our expertise,^{28,29} the sulfonamide-based CA inhibitors remain between the most successful drug candidates. The α -arylsulfonyl-methyl phosphonates we tested in the present study (compounds **1, 4-11**) exhibit a sulfonyl (-SO₂-) unit, and are therefore devoid of the possible second coordinating group. For comparison, the α -arylsulfonyl-methyl carboxylate and hydroxamate analogs (compounds **2** and **3**, respectively) have been also investigated.

It should be mentioned that compounds **12-15** can be considered under all respects as *N*-(biphenylsulfonyl)-phosphoalanyl derivatives, in that they contain the phosphonic counterpart of

aminoacid valine. By testing chiral **12** and **15**, which incorporate a residue of (*R*)-phosphoalanyl, we intended to evaluate the impact of isopropyl side-chain orientation on CA inhibitory activity and selectivity.

Experimental

The synthetic protocol for compounds **1-15** has been previously reported^{16,17,24}.

CA inhibition

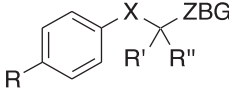
An applied photophysics stopped-flow instrument has been used for assaying the CA catalyzed CO₂ hydration activity. Phenol red (at a concentration of 0.2 mM) has been used as indicator, working at the absorbance maximum of 557 nm, with 20 mM Hepes, pH 7.5, as buffer, and 20 mM NaClO₄ (for maintaining constant the ionic strength), following the initial rates of the CA-catalyzed CO₂ hydration reaction for a period of 10-100 s. The CO₂ concentrations ranged from 1.7 to 17 mM for the determination of the kinetic parameters and inhibition constants. For each inhibitor at least six traces of the initial 5-10% of the reaction have been used for determining the initial velocity. The uncatalyzed rates were determined in the same manner and subtracted from the total observed rates. Stock solutions of inhibitor (10-50 mM) were prepared in distilled-deionized water and dilutions up to 0.1 nM were done thereafter with the assay buffer. Inhibitor and enzyme solutions were preincubated together for 15 min at room temperature prior to assay, in order to allow for the formation of the E-I complex or for the eventual active site mediated hydrolysis of the inhibitor. The IC₅₀ values were obtained by non-linear least-squares methods using PRISM 3 and represent the molarity of inhibitor at which the enzyme activity was halved (mean from at least three different determinations). Human CA I, II, IX, XII, and XIV were recombinant proteins obtained in-house as described earlier³⁰⁻³⁷.

Results and discussion

The inhibitory properties of the previously identified MMP inhibitors **1-15** have been investigated on five CA members, and more specifically against the cytosolic human CA I and II, which are widespread throughout the human body and represent drug targets for clinically used diuretics, antiglaucoma drugs, and anticonvulsants, as well as on the trans-membrane human isoforms CA IX and XII, that are validated antitumor targets, and XIV, which has been described as a possible objective for the treatment of epilepsy, some retinopathies, and skin tumors³¹. The standard, clinically used, CA inhibitor acetazolamide (AAZ) has been used as reference compound.

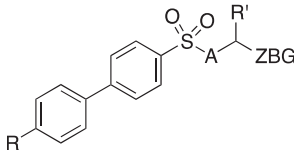
The initial investigation of compounds **1-15** on CA I and II meets the purpose to assay the novel inhibitory activity of this set of phosphonate-based MMPis against cytoplasmic CAs, and to evaluate the most active compounds against trans-membrane CAs, in order to obtain a more complete inhibitory profile. This is congruent, in view of the high degree of sequence and structural homology at the sites relevant to catalysis in all mammalian CA isoforms.

The *in vitro* inhibition effects of derivatives **1-15** on CA isoforms I and II are outlined in Table 1. All compounds resulted inactive on the ubiquitous CA I, with only the hydroxamic derivative **3** showing a very weak activity against this isozyme (IC₅₀ = 8.3 μ M). A better inhibitory profile was observed on CA II,

Table 1. Inhibition data for compounds 1–15 against CAs.


Compound	R	R'	R''	X	ZBG	IC ₅₀ /μM ^a	
						CA I	CA II
AAZ						0.299	0.023
1	C ₆ H ₅	H	H	SO ₂	PO ₃ H ₂	>10	>10
2	C ₆ H ₅	H	H	SO ₂	COOH	>10	>10
3	C ₆ H ₅	H	H	SO ₂	CONHOH	8.299	3.660
4	4-CH ₃ O-C ₆ H ₄	H	H	SO ₂	PO ₃ H ₂	>10	>10
5	4-CF ₃ -C ₆ H ₄	H	H	SO ₂	PO ₃ H ₂	>10	>10
6	3-Cl-C ₆ H ₄	H	H	SO ₂	PO ₃ H ₂	>10	>10
7	2-thienyl	H	H	SO ₂	PO ₃ H ₂	>10	>10
8	4-CH ₃ O-C ₆ H ₄ O	H	H	SO ₂	PO ₃ H ₂	>10	>10
9	C ₆ H ₅	H	(CH ₃) ₂ CHCH ₂	SO ₂	PO ₃ H ₂	>10	>10
10	4-CH ₃ O-C ₆ H ₄	H	C ₆ H ₅ CH ₂	SO ₂	PO ₃ H ₂	>10	>10
11	C ₆ H ₅	CH ₃	CH ₃	SO ₂	PO ₃ H ₂	>10	>10
(R)-12	4-CH ₃ -C ₆ H ₄	H	(CH ₃) ₂ CH	SO ₂ NH	PO ₃ H ₂	>10	4.140
(R)-13	3-CH ₃ -C ₆ H ₄	H	(CH ₃) ₂ CH	SO ₂ NH	PO ₃ H ₂	>10	>10
14	4-Br-	H	(CH ₃) ₂ CH	SO ₂ NH	PO ₃ H ₂	>10	>10
(R)-15	4-CH ₃ O-C ₆ H ₄	H	(CH ₃) ₂ CH	SO ₂ NH	PO ₃ H ₂	>10	3.777

^aErrors in the range of 5–10% of the reported value (from three different assays).

Table 2. Inhibition data for compounds 3, 10, 12, and 15 against CAs and MMPs.


Compound	R	R''	A	ZBG	IC ₅₀ /μM ^a						
					CA I	CA II	CA IX	CA XII	CA XIV	MMP-2	MMP-8
AAZ					0.299	0.023	0.903	0.088	0.224	ND	ND
3	H	H	–	CONHOH	8.299	3.660	0.527	8.862	8.525	0.0060	0.024
10	4-CH ₃ O	C ₆ H ₅ CH ₂	–	PO ₃ H ₂	>10	21.102	0.837	8.483	7.472	8.2	ND
(R)-12	4-CH ₃	(CH ₃) ₂ CH	NH	PO ₃ H ₂	>10	4.140 ^b	0.093	0.475	0.369	0.0023	0.0004
(R)-15	4-CH ₃ O	(CH ₃) ₂ CH	NH	PO ₃ H ₂	>10	3.777	0.084	6.220	1.521	0.0015	0.0014

^aErrors in the range of 5–10% of the reported value (from three different assays).

^bCA II from Sigma. pedices have been corrected.

but only for compounds **3**, **12**, and **15**, which resulted micromolar inhibitors of the enzyme, although with an almost 100-fold decrease in activity compared to the standard AAZ.

The most active compounds **3**, **12**, **15** were selected for subsequent evaluation on CA IX, XII and XIV. Sulfonyl derivative **10**, sharing the SAR-controlling 4'-CH₃O- substituent on the biphenyl group with the sulphonylammino-phosphonate **15**,^{17,24} was investigated as well on the cited isoforms, although essentially inactive on CA I and II. **Table 2** collects the observed inhibitory activities of such compounds on the complete group of CAs, together with the existing data on MMP-2 and MMP-8^{16,17,24}.

The potent MMP inhibitor of the hydroxamate-type **3** displayed only a one-digit micromolar inhibition on cytosolic isoforms CA I and II and membrane-bounds CA XII and XIV, but achieved its maximal potency on CA IX, with a sub-micromolar IC₅₀ that resulted almost half the value of the reference compound AAZ on the same isoform (0.5 vs. 0.9 μM, respectively).

Compound **10** resulted less active than the hydroxamic derivative **3** against MMP-2 and CAs; however, this sulfonyl derivative, with an IC₅₀ against CA IX in the high nanomolar range, that is almost 10 or even 20 (see CA II) times lower than the corresponding values for the remaining CAs, manifested a certain selectivity towards this isoform.

Sulfonylamino-derivatives **12** and **15**, while exhibiting an activity comparable to that of hydroxamate **3** on the cytosolic isoforms, presented a quite interesting inhibition profile against CA IX, with corresponding IC₅₀ values in the low nanomolar range. Furthermore **12** was able to inhibit, albeit with a 4-fold decrease in potency in comparison to CA IX, the other two isoforms CA XII (IC₅₀ = 0.47 μM) and CA XIV (IC₅₀ = 0.37 μM), thus resulting the most complete CA/MMP inhibitor. Compared to **12**, its (*R*)-*N*-(4'-methoxy) analog **15**, differing only for the CH₃O-substituent in the 4' position of the biphenyl unit instead of the CH₃ group, manifested weaker potencies on CA XII and XIV, but shared the low nanomolar inhibitory activity on CA IX, thus showing selectivity towards this latter isoform.

With respect to the inhibitory profiles of tested compounds **1–15**, the following observations can be pointed out: with the exception of the α -biphenylsulfonyl-methyl hydroxamate **3**, which inhibited CA I and II in the low micromolar range, the α -biphenylsulfonylmethyl carboxylic acid **2** and all the other substituted α -biarylsulfonylmethyl phosphonates (compounds **1**, **4–11**) were completely devoid of activity towards these off-target isoforms, regardless of the nature of the substituents. In spite of this, racemic 1-(4'-methoxybiphenyl-4-sulfonyl)-2-phenylethylphosphonic acid (**10**) was considered for inhibition of the three

membrane-bound CAs, resulting in the more selective CA IX phosphonate inhibitor of the series.

These experimental results could be tentatively analyzed in light of the assumption that none of the two oxygen atoms of the sulfonyl moiety is able to establish hydrogen bond interactions so productive as to reinforce the Zn²⁺-coordination network from the dissociated phosphonic (compounds **1**, **4–11**) or carboxylic acid group (compound **2**) inside the CA I and II active sites. As far as the α -biphenylsulfonyl-methyl hydroxamate **3** is concerned, we postulated that the stronger binding properties of the bidentate -CONHOH chelator may account for the appearance of the moderate to good inhibition observed towards the full panel of selected CAs. The expected lack of selectivity and the lipophilic character of hydroxamate **3**, on the other hand, were the main limitations we tried to deal with by introducing alternative, but clearly weaker, ZBGs in the sulfonyl series.

The nanomolar MMPs (*R*)-*N*-(4'-methylbiphenylsulfonyl)-phosphoalane (**12**) and (*R*)-*N*-(4'-methoxybiphenylsulfonyl)-phosphoalane (**15**) manifested excellent activities also on target-CAs, and particularly on CA IX. They both feature a secondary -SO₂NHR fragment positioned next to the ZBG, which possess good metal-coordinating properties; so we expect that both the sulfonamide and the phosphonate contribute to ion binding in a bidentate mode, somewhat in analogy to the model we proposed for the sulfonylaminoacyl-hydroxamate dual-inhibitors of the same enzyme families²⁷. With an inhibitory profile ranging from the sub-nanomolar IC₅₀ value against MMP-8 to the two/three digit nanomolar activities towards target CAs, compound **12** resulted the most powerful inhibitor in this set, thus presenting a potential in view of its cross-reactivity on cancer-related MMP/CA members.

Conclusions

Dysregulated expression of membrane-bound CA IX and XII results in decrease of extracellular pH, an event which is strongly associated with cancer cell survival and malignant progression. Thus specifically targeting these tumor-associated isoforms over the cytosolic CA I and II, which have a physiological relevance, is considered to be a promising strategy in cancer therapy. In recent years, multitarget approaches directed toward inhibition of CAs and enzymes of different families have received increasing attention. The MMPs are a family of metallo-endopeptidases that play a central role in many important physiological and physiopathological events, including tumor invasion and metastasis. Taking advantage of the fact that MMPs and CAs are both zinc-dependent enzymes, the approach of developing molecules with the ability to inhibit selected members of both families is particularly challenging.

In this article, we describe the inhibitory properties of a series of potent phosphonate-based MMP inhibitors against the cytosolic off-target isoforms CA I and II and the three trans-membrane (two of which are cancer-related) isoforms CA IX, XII and XIV. The most active inhibitors are found in the series of chiral *N*-(sulfonyl)phosphoalane-derivatives, which manifest good to excellent inhibitory activity over target CAs, with compound **15** showing the best isoform-selectivity toward CA IX. We identified compound **12** as our best inhibitor in view of its high and broad efficacy on the selected metalloenzymes: the ability of this small molecule to interact with different targets involved in cancerogenesis may be beneficial to address drawbacks of the classic anti-cancer protocol. Again, activity data confirm the peculiar role of the SO₂NH

functionality in stereoelectronic recognition and binding to the metal-core of the active site.

Despite the structural and catalytic differences of these two enzyme superfamilies, it is evident that MMP and CA active sites may be targeted by common, privileged structures. Closer investigations with the aim of understanding the overlapping chemical space of these target proteins will be necessary to further explore the dual-inhibition approach.

Disclosure statement

The authors declare no conflict of interest. The authors alone are responsible for the content and writing of this article.

Funding

This work was supported by G. d'Annunzio" University of Chieti-Pescara, (MIUR 2016, ex 60% Grant).

ORCID

Antonio Laghezza  <http://orcid.org/0000-0001-6221-6155>

References

- Guler OO, Capasso C, Supuran CT. A magnificent enzyme superfamily: carbonic anhydrases, their purification and characterization. *J Enzyme Inhib Med Chem* 2016;31:689–94.
- Supuran CT. Carbonic anhydrases: novel therapeutic applications for inhibitors and activators. *Nat Rev Drug Discov* 2008;7:168–81.
- Supuran CT. Structure and function of carbonic anhydrases. *Biochem J* 2016;473:2023–32.
- Supuran CT. Carbonic anhydrases-an overview. *Curr Pharm Des* 2008;14:603–14.
- Scozzafava A, Menabuoni L, Mincione F, et al. Carbonic anhydrase inhibitors: perfluoroalkyl/aryl-substituted derivatives of aromatic/heterocyclic sulfonamides as topical intra-ocular pressure-lowering agents with prolonged duration of action. *J Med Chem* 2000;43:4542–51.
- Thiry A, Dogné JM, Supuran CT, et al. Carbonic anhydrase inhibitors as anticonvulsant agents. *Curr Top Med Chem* 2007;7:855–64.
- Neri D, Supuran CT. Interfering with pH regulation in tumours as a therapeutic strategy. *Nat Rev Drug Discov* 2011;10:767–77.
- Schwartz L, Supuran CT, Alfarouk KO. The Warburg effect and the hallmarks of cancer. *Anticancer Agents Med Chem* 2017;17:164–70.
- Monti SM, Supuran CT, De Simone G. Anticancer carbonic anhydrase inhibitors: a patent review (2008 - 2013). *Expert Opin Ther Pat.* 2013;23:737–49.
- Waheed A, Sly WS. Carbonic anhydrase XII functions in health and disease. *Gene* 2017;623:33–40.
- Nagase H, Woessner JF Jr. Matrix metalloproteinases. *J Biol Chem* 1999;274:21491–4.
- Liotta LA, Tryggvason K, Garbisa S, et al. Metastatic potential correlates with enzymatic degradation of basement membrane collagen. *Nature* 1980;284:67–8.
- Mohan V, Talmi-Frank D, Arkadsh V, et al. Matrix metalloproteinase protein inhibitors: highlighting a new beginning for

- metalloproteinases in medicine. *Metalloproteinases Med* 2016;3:31–47.
14. Tauro M, Shay G, Sansil SS, et al. Bone-seeking matrix metalloproteinase-2 inhibitors prevent bone metastatic breast cancer growth. *Mol Cancer Ther* 2017;16:494–505.
 15. Campestre C, Agamennone M, Tauro M, et al. Phosphonate emerging zinc binding group in matrix metalloproteinase inhibitors. *Curr Drug Targets* 2015;16:1634–44.
 16. Biasone A, Tortorella P, Campestre C, et al. Alpha-biphenyl-sulfonylamino 2-methylpropyl phosphonates: enantioselective synthesis and selective inhibition of MMPs. *Bioorg Med Chem* 2007;15:791–9.
 17. Pochetti G, Gavuzzo E, Campestre C, et al. Structural insight into the stereoselective inhibition of MMP-8 by enantiomeric sulfonamide phosphonates. *J Med Chem* 2006;49:923–31.
 18. Vandembroucke RE, Libert C. Is there new hope for therapeutic matrix metalloproteinase inhibition? *Nat Rev Drug Discov* 2014;13:904–27.
 19. Supuran CT, Scozzafava A, Mastrolorenzo A. Bacterial proteases: current therapeutic use and future prospects for the development of new antibiotics. *Expert Opin Ther Pat* 2001;11:221–59.
 20. Hoffman A, Qadri B, Frant J, et al. Carbamoylphosphonate matrix metalloproteinase inhibitors 6: cis-2-aminocyclohexyl-carbamoylphosphonic acid, a novel orally active antimetastatic matrix metalloproteinase-2 selective inhibitor—synthesis and pharmacodynamic and pharmacokinetic analysis. *J Med Chem* 2008;51:1406–14.
 21. Folgueras AR, Fueyo A, García-Suárez O, et al. Collagenase-2 deficiency or inhibition impairs experimental autoimmune encephalomyelitis in mice. *J Biol Chem* 2008;283:9465–74.
 22. Vandembroucke RE, Dejonckheere E, Van Lint P, et al. Matrix metalloprotease 8-dependent extracellular matrix cleavage at the blood-CSF barrier contributes to lethality during systemic inflammatory diseases. *J Neurosci* 2012;32:9805–16.
 23. Albaiceta GM, Gutierrez-Fernández A, García-Prieto E, et al. Absence or inhibition of matrix metalloproteinase-8 decreases ventilator-induced lung injury. *Am J Respir Cell Mol Biol* 2010;43:555–63.
 24. Rubino MT, Agamennone M, Campestre C, et al. Synthesis, SAR, and biological evaluation of alpha-sulfonylphosphonic acids as selective matrix metalloproteinase inhibitors. *Chem Med Chem* 2009;4:352–62.
 25. Tauro M, Loiodice F, Ceruso M, et al. Arylamino bisphosphonates: potent and selective inhibitors of the tumor-associated carbonic anhydrase XII. *Bioorg Med Chem Lett* 2014;24:1941–3.
 26. Tauro M, Loiodice F, Ceruso M, et al. Dual carbonic anhydrase/matrix metalloproteinase inhibitors incorporating bisphosphonic acid moieties targeting bone tumors. *Bioorg Med Chem Lett* 2014;24:2617–20.
 27. Scozzafava A, Supuran CT. Carbonic anhydrase and matrix metalloproteinase inhibitors: sulfonylated amino acid hydroxamates with MMP inhibitory properties act as efficient inhibitors of CA isozymes I, II, and IV, and N-hydroxysulfonamides inhibit both these zinc enzymes. *J Med Chem* 2000;43:3677–87.
 28. Supuran CT. How many carbonic anhydrase inhibition mechanisms exist? *J Enzyme Inhib Med Chem* 2016;31:345–60.
 29. Supuran CT. Advances in structure-based drug discovery of carbonic anhydrase inhibitors. *Expert Opin Drug Discov* 2017;12:61–88.
 30. Alterio V, Pan P, Parkkila S, et al. The structural comparison between membrane-associated human carbonic anhydrases provides insights into drug design of selective inhibitors. *Biopolymers* 2014;101:769–78.
 31. Winum JY, Temperini C, El Cheikh K, et al. Carbonic anhydrase inhibitors: Clash with Ala65 as a means for designing inhibitors with low affinity for the ubiquitous isozyme II, exemplified by the crystal structure of the topiramate sulfamide analogue. *J Med Chem* 2006;49:7024–31.
 32. Supuran CT, Capasso C. New light on bacterial carbonic anhydrases phylogeny based on the analysis of signal peptide sequences. *J Enzyme Inhib Med Chem* 2016;31:1254–60.
 33. Briganti F, Pierattelli R, Scozzafava A, et al. Carbonic anhydrase inhibitors. Part 37. Novel classes of carbonic anhydrase inhibitors and their interaction with the native and cobalt-substituted enzyme: kinetic and spectroscopic investigations. *Eur J Med Chem* 1996;31:1001–10.
 34. Supuran CT, Clare BW. Carbonic anhydrase inhibitors. Part 57. Quantum chemical QSAR of a group of 1,3,4-thiadiazole and 1,3,4-thiadiazoline disulfonamides with carbonic anhydrase inhibitory properties. *Eur J Med Chem* 1999;34:41–50.
 35. Supuran CT, Barboiu M, Luca C, et al. Carbonic anhydrase activators. Part 14. Syntheses of mono and bis pyridinium salt derivatives of 2-amino-5-(2-aminoethyl)- and 2-amino-5-(3-aminopropyl)-1, 3, 4-thiadiazole and their interaction with isozyme II. *Eur J Med Chem* 1996;31:597–606.
 36. Scozzafava A, Menabuoni L, Mincione F, et al. Carbonic anhydrase inhibitors. A general approach for the preparation of water-soluble sulfonamides incorporating polyamino – polycarboxylate tails and of their metal complexes possessing long-lasting, topical intraocular pressure-lowering properties. *J Med Chem* 2002;45:1466–76.
 37. Abbate F, Winum JY, Potter BV, et al. Carbonic anhydrase inhibitors: X-ray crystallographic structure of the adduct of human isozyme II with EMATE, a dual inhibitor of carbonic anhydrases and steroid sulfatase. *Bioorg Med Chem Lett* 2004;14:231–4.