Ganji Purnachandra Nagaraju Pallaval Veera Bramhachari *Editors* 

Role of Transcription Factors in Gastrointestinal Malignancies



Materiale protetto da copyrigh



# Targeting Arachidonic Acid Pathway-Associated NF-kB in Pancreatic Cancer

30

Nagendra Sastry Yarla, Olga Sukocheva, Ilaria Peluso, Swathi Putta, Pallaval Veera Bramhachari, Rajesh Yadala, Dinesh K. Tiwari, Srinivas Jagarlamudi, Luciana Scotti, Marcus T. Scotti, Marcella Reale, Mohammad Amjad Kamal, Ashraf Ghulam, Bechan Sharma, Madhukiran Parvathaneni, Chinthalapally V. Rao, Mastan Mannarapu, and Anupam Bishayee

## 30.1 Introduction

Pancreatic cancer is the fourth-leading cause of cancer-related morbidity, with 39,590 deaths and 46,420 estimated new cases in 2014 in the United States [1]. Mutations are considered to be the genetic event occurring at the beginning of the development process of pancreatic cancer, leading to cellular proliferation due to constitutive activation of intracellular pathways [2]. The latter is related to numerous alterations in growth factors and their receptors, involved in the control of growth and differentiation during transduction pathways [2]. There are several molecular signaling pathways that play a vital role in cell survival, proliferation,

O. Sukocheva (⊠) School of Health Sciences, Flinders University of South Australia, Bedford Park, Adelaide 5042, South Australia, Australia

I. Peluso Research Centre for Food and Nutrition, Council for Agricultural Research and Economics, Rome, Italy

S. Putta

P. V. Bramhachari Department of Biotechnology, Krishna University, Machilipatnam, Andhra Pradesh, India

L. Scotti · M. T. Scotti Federal University of Paraiba, João Pessoa, Paraíba, Brazil

© Springer Nature Singapore Pte Ltd. 2017

G. P. Nagaraju, P. V. Bramhachari (eds.), *Role of Transcription Factors in Gastrointestinal Malignancies*, https://doi.org/10.1007/978-981-10-6728-0\_30

N. S. Yarla (🖂) · R. Yadala · D. K. Tiwari · S. Jagarlamudi

School of Life Sciences, University of Hyderabad, Hyderabad, India e-mail: sastryyn@gmail.com

Department of Pharmaceutical Sciences, University College of Pharmaceutical Sciences, Andhra University, Visakhapatnam, Andhra Pradesh, India

angiogenesis, metastasis, and promotion of pancreatic cancer, as well as in chemoresistance, including epidermal growth factor receptor (EGFR), nuclear factorkappa B (NF- $\kappa$ B), signal transducer and activator of transcription factor 3 (STAT3), and arachidonic acid (AA) pathway [3].

### 30.2 Arachidonic Acid Pathway-Associated NF-κB Signaling in Pancreatic Cancer

The transcription factor NF- $\kappa$ B plays a critical role in pancreatic cancer as involving at the downstream stage of many signaling cascades including AA pathway. AA is an upstream mediator and regulator of NF- $\kappa$ B pathway. NF- $\kappa$ B and activation protein 1 (AP-1) can bind to the cis-acting elements in the promoter of phospholipase A2 (PLA2), cyclooxygenase (COX-2), and lipoxygenases (LOXs), and these pathways are upregulated in several cancers. The active form of PLA<sub>2</sub> stimulates the proliferation of MIAPaCa-2 pancreatic cancer cells by the activation of mitogen-activated protein kinases (MAPKs)/NF- $\kappa$ B [4]. PLA<sub>2</sub>s gene knockout and transgenic studies demonstrated the pro-tumorigenic role of PLA<sub>2</sub> in pancreatic cancer [5].

M. Reale

M. A. Kamal King Fahd Medical Research Center, King Abdulaziz University, Jeddah, Saudi Arabia

Enzymoics, 7 Peterlee Place, Hebersham, NSW, Australia

Novel Global Community Educational Foundation, Hebersham, Australia

A. Ghulam

King Fahd Medical Research Center, King Abdulaziz University, Jeddah, Saudi Arabia

B. Sharma

Department of Biochemistry, University of Allahabad, Allahabad, India

M. Parvathaneni Department of Biotechnology, Harrisburg University of Science and Technology, Harrisburg, PA, USA

C. V. Rao

Center for Cancer Prevention and Drug Development, Department of Medicine, Hematology/Oncology Section, University of Oklahoma Health Sciences Center (OUHSC), Oklahoma City, OK, USA

M. Mannarapu Department of Biotechnology, Dravidian University Kuppam, Chittoor, Andhra Pradesh 517 426, India

A. Bishayee Department of Pharmaceutical Sciences, College of Pharmacy, Larkin University, Miami, FL 33169, USA

Department of Medical, Oral and Biotechnological Sciences, University "G. D'Annunzio", Chieti, Italy

A chemical compound bromoenol lactone (BEL) caused decrease in agonistinduced activation of EGFR in pancreatic cancer cells [6]. The NF- $\kappa$ B inhibition has become a major target in natural drug discovery, and it is necessary for survival and immunity of cell because it regulates the immune, inflammatory, and carcinogenic responses [7]. Compared to normal pancreatic tissues, NF- $\kappa$ B, which is linked with COX-2, is constitutively expressed in 67% of human pancreatic cancers. Zhang et al. [5] reported the acute pancreatitis induced by PLA2 by the activation of transcription factor NF- $\kappa$ B. Gong et al. [8] reported combinatorial target of COX-2/ NF- $\kappa$ B/STAT3/prostaglandin receptor E-prostanoid 4 pathway for effective treatment of pancreatic cancer [8, 9] reported the TLR4-/NF- $\kappa$ B-/COX-2-mediated inflammatory pathway involved in tumorigenesis of pancreatic cancer.

Although COX-2 expression along with NF- $\kappa$ B is incremented in several types of malignancies including pancreatic, esophagus, colon, and breast cancers [10–12], the molecular mechanism is still unclear. It has been reported that COX-2 has inherent role in pancreatic ductal adenocarcinomas initiation and progression by the activation of the PI3K/AKT pathway [13]. In cancer, prostaglandin E2 (PGE2) the metabolite of COX-2 is overexpressed in the majority of epithelial malignancies [14, 15]. Approximately 80% of human pancreatic ductal adenocarcinomas (PDA) overexpress the tumor form of mucin 1 (tMUC1), a heavily glycosylated membrane-tethered glycoprotein generally expressed on glandular epithelial cells [16]. tMUC1 is overexpressed and aberrantly hypoglycosylated in malignant cells [16].

In a study, it has been shown that, in addition to the overexpression of tMUC1, the tumors have high levels of COX-2 and PGE2 [17]. Mucin 1 (PDA.MUC1) mice are highly resistant to celecoxib (a COX-2 inhibitor) and gemcitabine when each drug is administered independently; however, a clinical relevant antitumor response was observed after treatment with a blend of MUC1 vaccine, celecoxib, and gemcitabine [18]. Previous reports suggested that the transmembrane mucin glycoprotein mucin 1 (MUC1) is overexpressed in pancreatic ductal adenocarcinomas along with COX-2. Molecular studies by Nath et al. [17] demonstrated the MUC1 associated with the same gene locus where NF- $\kappa$ B/p65 binds with the COX-2 promoter. The study suggests that MUC1 regulates pancreatic ductal adenocarcinomas through COX-2/NF- $\kappa$ B pathway [17]. Animal LOXs were classified into five different types—5-LOX, 8-LOX, 11-LOX, 12-LOX, and 15-LOX [19]. LOXs also involved in various human cancers including colon, pancreatic, lung, breast, and prostate [10]; however, comparatively diminutive efforts have been made to explicate its role in cancer development.

Recently, it has been shown that treatment with omega-6 fatty acids increases leukotrienes B4 (a LOX metabolite) levels in human pancreatic ductal epithelial (HPDE and HPDE-Kras) and cancer (AsPC1 and Panc1) cells, in vitro [20]. Compared to EL-Kras/5LO+/+ mice, EL-Kras mice lacking 5LO (EL-Kras/5LO-/-) had decreased mast cell infiltration and developed fewer pancreatic lesions [20]. These results suggest the LOXs involvement in pancreatic cancer. Previously, the progression of human pancreatic cancer cells by leukotrienes B4 (LTB4) via MAPK and PI-3 kinase pathways was reported, although it did not affect the activity of JNK/SAPK [21]. In fact, different LOXs exhibit tumor response in a tissue-specific

manner either pro-tumorigenic or antitumorigenic activities [22]. Inhibitors of lipoxygenases have been found to be very effective in the suppression of pancreatic cancer cell lines as well as pancreatic adenocarcinoma. In a study, the inhibition of 5-lipoxygenase by zileuton (5-LOX inhibitor) in pancreatic cancer cells by inducing apoptosis, SW1990, has been demonstrated [23].

#### 30.3 Targeting Arachidonic Acid Pathway-Associated NF-κB by Phytochemical Compounds for Prevention and Therapy of Pancreatic Cancer

Phytochemical agents and their derivatives act as anticancer drugs by inhibiting the migration, invasion, growth, survival, and metastasis of cancer cell during the carcinogenesis process by multiple pathways [24]. Cannabinoid (1), lupeol (2), plumbagin (3), quercetin (4), sulforaphane (5), triptolide (6),  $\gamma$ -tocotrienol (7), boswellic acid (8), curcumin (9), and garcinol (10) are different phytochemicals exhibiting anticancer activities against various cancers, including pancreatic, by targeting AA pathway and NF- $\kappa$ B-mediated signaling pathways (Table 30.1).

Moreover, paclitaxel, etoposide and teniposide, vinblastine and vincristine, and camptothecin derivatives are reported anticancer agents [25, 26]. These compounds act by inhibiting the expression of NF- $\kappa$ B and associated genes by modulating various signal transduction pathways [25, 26]. Researchers reported that individual phytochemicals and in combination with chemotherapeutics can prevent pancreatic cancer by inhibiting several signaling pathways [27]. Phenolics can inhibit the promotion and progression of cancerous cell by modulating the activities of NF- $\kappa$ B and AP-1 [28]. Curcumin downregulates the expression of NF- $\kappa$ B, COX2, and phosphorylated STAT3 in peripheral blood mononuclear cells in some pancreatic cancer patients [29] (Fig. 30.1). Lev-Ari et al. [30] reported that combination of curcumin and gemcitabine enhanced cytotoxic effect against pancreatic adenocarcinoma in vitro, decreasing the COX-2 and p-ERK1/2 levels.

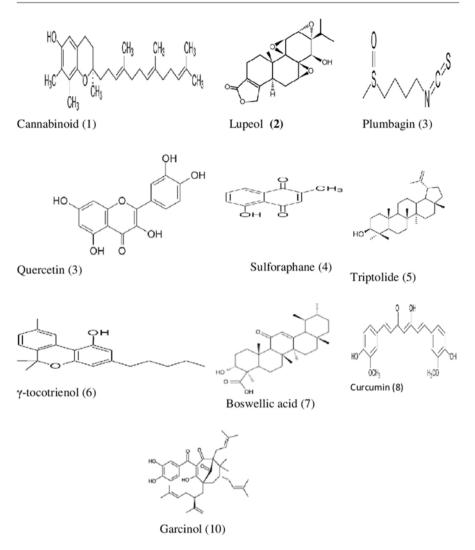
Cascinu et al. [31] conducted a clinical trial in pancreatic cancer, by using celecoxib (COX-2 inhibitor) associated to oxaliplatin and gemcitabine. The expression of COX-2 was found in 30 tumors as well as NF- $\kappa$ B expression was found in 16 tumors. Authors reveal that COX-2 inhibitor does not enhance the efficacy of gemcitabine/oxaliplatin. Nafamostat mesilate inhibits induced apoptosis in pancreatic cancer cells, but its maximum concentration ( $1.8 \times 10^{-7}$  M) is not effective to inhibit NF- $\kappa$ B [33]. The recommended dose of regional arterial infusion of nafamostat mesilate ( $1.8 \times 10^{-6}$  M), used to target gemcitabine-induced NF- $\kappa$ B activation, in combination with gemcitabine is found to be safe in pancreatic cancer patients [32]. The inhibition of gemcitabine-induced NF- $\kappa$ B activation enhances the antitumor activity of gemcitabine against pancreatic cancer [34].

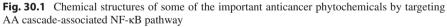
Pristimerin, a quinonemethide triterpenoid, exhibited antiproliferative and proapoptotic activities in pancreatic cancer cells by inhibiting multiple signaling pathways NF- $\kappa$ B/COX-2 [35]. Nordihydroguaiaretic acid (NDGA), a 5-LOX inhibitor,

Phytochemical	Mechanism	References
Cannabinoid (1)	Upregulation of CB2 receptor	[37]
	Upregulation of p8, ATF-4, and TRB3 genes	
Lupeol (2)	Reduced activation of NF-KB	[38]
	Reduced protein expression of PKCa/ODC, PI3K/Akt, and	
	MAPK pathways	
Plumbagin (3)	Decrease in NF-κB/p65 phosphorylation	[39]
	Inhibition of STAT3 phosphorylation	
	Upregulates the expression of IL-6	
	Inhibited expression of Cdc25A, cyclin D1, and MMP9	
	Decreased expression of proliferative markers such as PCNA and ki67	
Quercetin (4)	Diminished ALDH1	[40]
	Inhibition of NF-KB	
	Reduced proliferation, angiogenesis, cancer stem cell-marker	
	expression	
	Induction of apoptosis	
Sulforaphane (5)	Diminished NF-kB binding, induction of apoptosis	[41]
	Blockage of tumor growth and angiogenesis	
Triptolide (6)	Downregulates NF-KB	[42]
	Inhibition of HSF1 and HSP70	
	Inhibition of glycosylation of Sp1	
	Inhibition of hexosamine biosynthesis	
	Inhibition of O-GlcNAc transferase	
γ-Tocotrienol (7)	Inhibition of NF-κB activity	[43]
	Inhibition of cell growth and cell survival	
	Decreased p65 (ReIA) binding	
	Inhibition of proliferation, angiogenesis, invasion	
Boswellic acid (8)	Downregulate the expression of COX-2, MMP-9, CXCR4,	[44]
	and VEGF and inhibit the expression level of c-Myc	
Curcumin (9)	Downregulation of iNOS, COX-2, and 5-LOX expression	[45]
	Upregulation of p21 expression	
Garcinol (10)	Reduction in PGE2 expression	[46]
	Downregulation of NF-KB and COX-2	
	O2, nitric oxide (NO), iNOS, and COX2	

**Table 30.1** Phytochemicals targeting AA pathway associated transcriptional factros for cancer prevention and therapy

prevented the progression of acute pancreatitis by inhibiting multiple pathways, including NF- $\kappa$ B [36]. Nexrutine®, an anti-inflammatory nutraceutical, inhibited growth of pancreatic cancer cells and reduced levels and activity of NF- $\kappa$ B, expression of COX-2, and subsequent decreased levels of PGE2 and PGF2 [8]. 6-Shogaol (a phenol extracted from ginger) alone and in combination with gemcitabine suppressed the growth of pancreatic tumors, by suppressing the TLR4-/NF- $\kappa$ B-/COX-2-mediated pathway [9].





## 30.4 Conclusions and Future Directions

NF- $\kappa$ B is an important transcription factor and has a crucial role in pancreatic cancer as it is involved at the downstream stage of many signaling cascades including AA pathway. Metabolic enzymes and products of AA pathway are also involved in activation of NF- $\kappa$ B. Cross talk between eicosanoids and NF- $\kappa$ B plays a key role in pathophysiology of pancreatic cancer. Therefore, researchers have considered AA cascade-associated NF- $\kappa$ B pathway as novel therapeutic target in pancreatic cancer.

Moreover, several AA cascade-associated NF-kB pathways are targeted by natural products. Therefore, these compounds have been suggested as novel preventive and therapeutic agents against pancreatic cancer.

#### References

- 1. Urayama S (2015) Pancreatic cancer early detection: expanding higher-risk group with clinical and metabolomics parameters. World J Gastroenterol 21(6):1707–1717
- 2. Xiong HQ (2004) Molecular targeting therapy for pancreatic cancer. Cancer Chemother Pharmacol 54:69–77
- Gupta SC, Kim JH, Kannappan R, Reuter S, Dougherty PM, Aggarwal BB (2011) Role of nuclear factor-κB-mediated inflammatory pathways in cancer-related symptoms and their regulation by nutritional agents. Exp Biol Med 236:658–671
- Kashiwagi M, Friess H, Uhl W et al (1998) Phospholipase A2 isoforms are altered in chronic pancreatitis. Ann Surg 227(2):220–228
- Zhang MS, Zhang KJ, Zhang J, Jiao XL, Chen D, Zhang DL (2014) Phospholipases A-II (PLA2-II) induces acute pancreatitis through activation of the transcription factor NF-kappaB. Eur Rev Med Pharmacol Sci 18(8):1163–1169
- Yarla NS, Bishayee A, Vadlakonda L, Chintala R et al (2016) Phospholipase A2 isoforms as novel targets for prevention and treatment of inflammatory and oncologic diseases. Curr Drug Targets 17(16):1940–1962
- Senthilraja P, Kayitare J, Manivel G, Manikanda Prabhu S, Krishnamurthy A (2015) Potential compound derived from *Catharanthus roseus* to inhibit Non-Small Cell Lung Cancer (NSCLC). Int J Res Ayurveda Pharm 6(2):265–271
- Gong J, Xie J, Bedolla R, Rivas P, Chakravarthy D (2014) Combined targeting of STAT3/ NF-κB/COX-2/EP4 for effective management of pancreatic cancer. Clin Cancer Res 20(5):1259–1273
- Zhou L, Qi L, Jiang L, Zhou P et al (2014) Antitumor activity of gemcitabine can be potentiated in pancreatic cancer through modulation of TLR4/NF-κB signaling by 6-shogaol. AAPS J 16(2):246–257
- Knab LM, Grippo PJ, Bentrem DJ (2014) Involvement of eicosanoids in the pathogenesis of pancreatic cancer: the roles of cyclooxygenase-2 and 5-lipoxygenase. World J Gastroenterol 20(31):10729–10739
- 11. Lin DT, Subbaramaiah K, Shah JP, Dannenberg AJ, Boyle JO (2002) Cyclooxygenase-2: a novel molecular target for the prevention and treatment of head and neck cancer. Head Neck 24:792–799. https://doi.org/10.1002/hed.10108
- 12. Howe LR, Dannenberg AJ (2002) A role for cyclooxygenase-2 inhibitors in the prevention and treatment of cancer. Semin Oncol 29:111–119
- Hill R, Li Y, Tran LM, Dry S et al (2012) Cell intrinsic role of COX-2 in pancreatic cancer development. Mol Cancer Ther 11(10):2127–2137
- 14. Greenhough A, Smartt HJ, Moore AE et al (2009) The  $COX-2/PGE_2$  pathway: key roles in the hallmarks of cancer and adaptation to the tumour microenvironment. Carcinogenesis 30:377-386
- 15. Koehne C, Dubois R (2004) COX-2 inhibition and colorectal cancer. Semin Oncol 2:12-21
- Nath S, Mukherjee P (2014) MUC1: a multifaceted oncoprotein with a key role in cancer progression. Trends Mol Med 20:332–342
- Nath S, Roy LD, Grover P et al (2015) Mucin 1 regulates Cox-2 gene in pancreatic cancer. Pancreas 44(6):909–917
- Mukherjee P, Basu GD, Tinder TL et al (2009) Progression of pancreatic adenocarcinoma is significantly impeded with a combination of vaccine and COX-2 inhibition. J Immunol 182:216–224

- Ivanov I, Heydeck D, Hofheinz K, Roffeis J et al (2010) Molecular enzymology of lipoxygenases. Arch Biochem Biophys Elsevier Inc 503:161–174. https://doi.org/10.1016/j. abb.2010.08.016
- Knab LM, Schultz M, Principe DR, Mascarinas WE et al (2015) Ablation of 5-lipoxygenase mitigates pancreatic lesion development. J Surg Res 194(2):481–487
- Tong WG, Ding XZ, Talamonti MS, Bell RH, Adrian TE (2005) LTB4 stimulates growth of human pancreatic cancer cells via MAPK and PI-3 kinase pathways. Biochem Biophys Res Commun 335(3):949–956
- Comba A, Pasqualini ME (2009) Primers on molecular pathways lipoxygenases: their role as an oncogenic pathway in pancreatic cancer. Pancreatology 9(6):724–728
- Zhou GX, Ding XL, Wu SB, Zhang HF et al (2015) Inhibition of 5-lipoxygenase triggers apoptosis in pancreatic cancer cells. Oncol Rep 33(2):661–668
- Safe S, Kasiappan R (2016) Natural products as mechanism-based anticancer agents: Sp transcription factors as targets. Phytother Res 30(11):1723–1732
- 25. Sun M, Estrov Z, Ji Y, Kevin R, Coombes KR et al (2008) Curcumin (diferuloylmethane) alters the expression profiles of microRNAs in human pancreatic cancer cells. Mol Cancer Ther 7(3):464–473
- 26. Cragg GM, Newman DJ (2005) Plants as a source of anti-cancer agents. J Ethnopharmacol 100:72–79
- Banerjee S, Wang Z, Kong D, Sarkar FH (2009) 3, 30- Diindolylmethane enhances chemosensitivity of multiple chemotherapeutic agents in pancreatic cancer. Cancer Res 69:5592–5600
- Domenicoa FD, Foppolib C, Cocciaa R, Perluigi M (2012) Antioxidants in cervical cancer: chemopreventive and chemotherapeutic effects of polyphenols. Biochim Biophys Acta 1822(5):737–747
- 29. Dhillon N, Aggarwal BB, Newman RA, Wolff RA et al (2008) Phase II trial of curcumin in patients with advanced pancreatic cancer. Clin Cancer Res 14(14):4491–4499
- Lev-Ari S, Vexler A, Starr A, Ashkenazy-Voghera M, Greif J et al (2007) Curcumin augments gemcitabine cytotoxic effect on pancreatic adenocarcinoma cell lines. Cancer Investig 25:411–418
- 31. Cascinu S, Scartozzi M, Carbonari G, Pierantoni C et al (2007) COX-2 and NF-κB overexpression is common in pancreatic cancer but does not predict for COX-2 inhibitors activity in combination with gemcitabine and oxaliplatin. Am J Clin Oncol 30(5):526–530
- 32. Uwagawa T, Misawa T, Sakamoto T, Ito R et al (2009) A phase I study of full-dose gemcitabine and regional arterial infusion of nafamostat mesilate for advanced pancreatic cancer. Ann Oncol 20(2):239–243
- Yanaga K et al (2007) Mechanisms of synthetic serine protease inhibitor (FUT-175)-mediated cell death. Cancer 109:2142–2153
- 34. Banerjee S, Zhang Y, Ali S et al (2005) Molecular evidence for increased antitumor activity of gemcitabine by genistein in vitro and in vivo using an orthotopic model of pancreatic cancer. Cancer Res 65:9064–9072
- 35. Deeb D, Gao X, Liu YB, Pindolia K, Gautam SC (2014) Pristimerin, a quinonemethide triterpenoid, induces apoptosis in pancreatic cancer cells through the inhibition of pro-survival Akt/ NF-κB/mTOR signaling proteins and anti-apoptotic Bcl-2. Int J Oncol 44(5):1707–1715
- 36. Mahajan UM, Gupta C, Wagh PR, Karpe PA, Tikoo K (2011) Alteration in inflammatory/ apoptotic pathway and histone modifications by nordihydroguaiaretic acid prevents acute pancreatitis in Swiss albino mice. Apoptosis 16(11):1138–1149
- 37. Carracedo A, Gironella M, Lorente M, Garcia S et al (2006) Cannabinoids induce apoptosis of pancreatic tumor cells via endoplasmic reticulum stress-related genes. Cancer Res 66(13):6748–6755
- 38. Saleem M, Kaur S, Kweon MH, Adhami VM, Afaq F, Mukhtar H (2005) Lupeol, a fruit and vegetable based triterpene, induces apoptotic death of human pancreatic adenocarcinoma cells via inhibition of Ras signaling pathway. Carcinogenesis 26(11):1956–1964

- 39. Hafeez BB, Jamal MS, Fischer JW, Mustafa A, Verma AK (2012) Plumbagin, a plant derived natural agent inhibits the growth of pancreatic cancer cells in in vitro and in vivo via targeting EGFR, Stat3 and NF-κB signaling pathways. Int J Cancer 131:2175–2186
- Zhou W, Kallifatidis G, Baumann B, Rausch V et al (2010) Dietary polyphenol quercetin targets pancreatic cancer stem cells. Int J Oncol 37(3):551–561
- Kallifatidis G, Rausch V, Baumann B, Apel A et al (2009) Sulforaphane targets pancreatic tumour-initiating cells by NF-κB-induced antiapoptotic signaling. Gut 58(7):949–963
- 42. Banerjee S, Sangwan V, McGinn O, Chugh R, Dudeja V, Vickers SM, Saluja AK (2013) Triptolide-induced cell death in pancreatic cancer is mediated by *O*-GlcNAc modification of transcription factor Sp1. J Biol Chem 288(47):33927–33938
- 43. Husain K, Francois RA, Yamauchi T, Perez M, Sebti SM, Malafa MP (2011) Vitamin E d-tocotrienol augments the antitumor activity of gemcitabine and suppresses constitutive NF-κB activation in pancreatic cancer. Mol Cancer Ther 10:2363–2372
- 44. Park B, Prasad S, Yadav V, Sung B, Aggarwal BB (2011) Boswellic acid suppresses growth and metastasis of human pancreatic tumors in an orthotopic nude mouse model through modulation of multiple targets. PLoS One 6(10):e2694
- 45. Swamy MV, Citineni B, Jagan MR et al (2008) Prevention and treatment of pancreatic cancer by curcumin in combination with omega-3 fatty acids. Nutr Cancer 60:81–89
- Saadat N, Gupta SV (2012) Potential role of garcinol as an anticancer agent. J Oncol 2012.; 2012:647206. https://doi.org/10.1155/2012/647206