



Beyond the HLA Genes in Gluten-Related Disorders

Michele Sallese^{1,2*}, Loris Riccardo Lopetuso^{2,3}, Konstantinos Efthymakis³ and Matteo Neri^{2,3*}

¹ Department of Medical, Oral and Biotechnological Sciences, 'G. D'Annunzio' University of Chieti–Pescara, Chieti, Italy,

² Center for Advanced Studies and Technology (CAST), 'G. D'Annunzio' University of Chieti–Pescara, Chieti, Italy,

³ Department of Medicine and Aging Sciences, 'G. D'Annunzio' University of Chieti–Pescara, Chieti, Italy

OPEN ACCESS

Edited by:

Antonio Di Sabatino,
University of Pavia, Italy

Reviewed by:

Mourad Aribi,
University of Abou Bekr
Belkaid, Algeria
Keith R. Martin,
University of Memphis, United States

*Correspondence:

Michele Sallese
michele.sallese@unich.it
Matteo Neri
m.neri@unich.it

Specialty section:

This article was submitted to
Nutritional Immunology,
a section of the journal
Frontiers in Nutrition

Received: 24 June 2020

Accepted: 12 October 2020

Published: 12 November 2020

Citation:

Sallese M, Lopetuso LR, Efthymakis K
and Neri M (2020) Beyond the HLA
Genes in Gluten-Related Disorders.
Front. Nutr. 7:575844.
doi: 10.3389/fnut.2020.575844

Most common food grains contain gluten proteins and can cause adverse medical conditions generally known as gluten-related disorders. Celiac disease is an immune-mediated enteropathy triggered by gluten in individuals carrying a specific genetic make-up. The presence of the human leukocyte antigens (HLA)-DQ2 and HLA-DQ8 haplotypes together with gluten intake is a necessary, although not sufficient, condition, to develop celiac disease. Fine mapping of the human genome has revealed numerous genetic variants important in the development of this disease. Most of the genetic variants are small nucleotide polymorphisms located within promoters and transcriptional enhancer sequences. Their importance is underlined by an increased risk in DQ2/DQ8 carriers who also have these non-HLA alleles. In addition, several immune-mediated diseases share susceptibility loci with celiac disease, shedding light on the reasons for co-occurrence between these diseases. Finally, most of the genes potentially involved in celiac disease by fine genetic mapping of non-HLA loci were confirmed in gene expression studies. In contrast to celiac disease, very little is known about the genetic make-up of non-celiac wheat sensitivity (NCWS), a clinically defined pathology that shares symptoms and gluten dependence with the celiac disease. We recently identified differentially expressed genes and miRNAs in the intestinal mucosa of these patients. Remarkably, the differentially expressed genes were long non-coding RNAs possibly involved in the regulation of cell functions. Thus, we can speculate that important aspects of these diseases depend on alteration of regulatory genetic circuits. Furthermore, our finding suggests that innate immune response is involved in the pathogenic mechanism of NCWS. This review is intended to convey the idea that in order to fully understand celiac disease and its relationship with other gluten-related disorders, it is worth learning more about non-HLA variants.

Keywords: HLA, DQ2 and DQ8, non-HLA genes, gene expression, non-celiac gluten sensitivity (NCGS)

INTRODUCTION

Celiac disease is an immune-mediated enteropathy triggered by ingestion of gluten in genetically predisposed individuals (1, 2). Gluten is a protein mixture of prolamins and glutenin present in most diffused food cereals like wheat, rye, and barley. Gluten proteins are incompletely fragmented by the digestive proteases and residual peptides, crossing the intestinal epithelial barrier, reach the lamina propria (3, 4). Gluten peptides have

cytotoxic, immunomodulatory and permeabilizing effects/properties. Contact with enterocytes leads to the production of interleukin (IL)15, a key mediator of celiac disease, the release of zonulin, and the activation of tissue transglutaminase (TG2) (5, 6). IL15 upregulates human leukocyte antigen (HLA)-E and major histocompatibility complex class I chain-related protein A (MICA) on enterocytes and their corresponding receptors, as well as cluster of differentiation (CD)94 and natural killer group 2D (NKG2D) on natural killer and CD8⁺ lymphocytes, thus contributing to mucosal damage (6, 7). Zonulin leads to the loosening of tight junctions and increased intestinal permeability. Glutamine residues of gluten peptides in the lamina propria are deamidated to glutamic acid by activated tissue transglutaminase (TG2). The introduction of negative charges into gluten peptides promotes their stable interaction with HLA class II in antigen presenting cells (APC) and thus the presentation of these antigens to T lymphocytes in lamina propria (8–10). Activated CD4⁺ T-cells produce IL21 and interferon (IFN) γ that support the activation of intraepithelial CD8⁺ lymphocytes which in turn damage enterocytes and cause villous atrophy (6). Cytokines released by activated T cells also contribute to the differentiation of B lymphocytes into plasma cells and the production of antibodies, mainly of the IgA subtype, against gluten peptides and TG2. The presence of anti-TG2 autoantibodies and villous atrophy, two main features of celiac disease, are also essential diagnostic markers (1).

The onset of celiac disease, besides gluten ingestion and the expression of specific HLAs, requires additional ill-defined factors. For example, a particular stressful event like the loss of a relative, a pregnancy, or an infection have been considered potential triggers of celiac disease. Nevertheless, even the simultaneous presence of these conditions not necessarily leads to the development of the disease, likely because additional genetic factors are involved. In this context, genetic factors beyond HLA have gained consistent relevance either as predisposing to the occurrence of celiac disease or as favoring the progression toward complicated disease or the presence of extraintestinal or associated diseases.

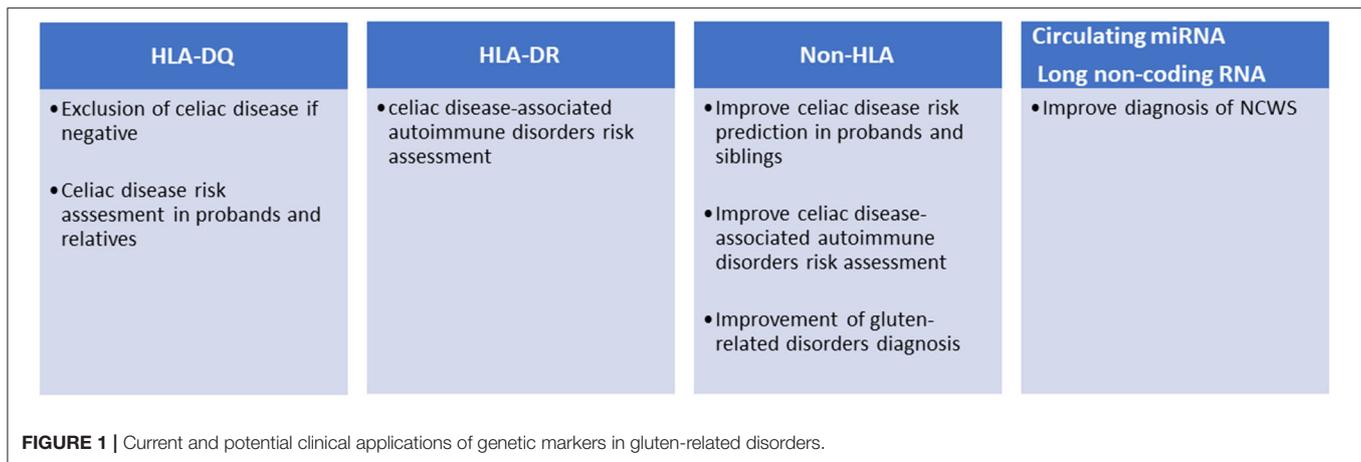
Moreover, some patients claiming persistent symptoms related to gluten consumption including abdominal distension and pain, diarrhea, tiredness, and dermatitis show negative celiac disease serology and preserved mucosal architecture. This characteristic phenotype has been defined non-celiac gluten sensitivity (NCWS), as symptoms respond to gluten exclusion (11, 12). Patients with NCWS have a normal intestinal mucosa on histological examination, and intraepithelial lymphocytes (IELs) counts are <25 per 100 enterocytes in about 60% of cases while ranging from 25 to 40 in the rest of cases. To date, very little is known about the pathological mechanisms, environmental factors, and genetic makeup predisposing to NCWS.

In this minireview we report the current knowledge regarding genetic variants and gene expression changes that have been associated to celiac disease and NCWS, focusing on non-HLA genes.

HLA LOCUS IS THE MAIN PREDISPOSING FACTOR FOR THE DEVELOPMENT OF CELIAC DISEASE

More than 90% of patients affected by celiac disease are carriers of HLA-DQ2, while the rest are carriers of HLA-DQ8 (13). The expression of these HLAs appears a necessary condition for celiac disease development, while their absence almost definitively excludes it (**Figure 1**). The presence of HLA-DQ2/DQ8 haplotypes is often used to settle borderline serological and histological tests. The prevalence of HLA-DQ2/DQ8 in the general population ranges from 30 to 40%, but only about 3% of the carriers develop celiac disease suggesting that additional genetic factors may be required (14). HLA-DQ2 is a heterodimer encoded by different DQA1 (α chain) and DQB1 (β chain) genes. Specifically, DQA1*0501 and DQB1*0201 form the HLA-DQ2.5, while the DQA1*0201 and DQB1*0202 form the HLA-DQ2.2. The DQ2.5 haplotype can be carried in *cis*-configuration, where DQA1*0501 and DQB1*0201 are both present on the same chromosome copy (paternal, maternal, or both); alternatively, it can be present in *trans*-configuration where the genes for the α and β chains are each separately present on the parental chromosomes. Haplotypes can further be homozygous or heterozygous; thus, HLA-DQ2.5 can be homozygous, heterozygous in *cis*, and heterozygous in *trans*. People carrying the DQ2.5 haplotype have a high risk to develop celiac disease, in comparison to those carrying DQ2.2 (15). DQB1*0201 is highly associated to disease risk; patients carrying two copies of DQB1*0201 show a five-fold risk to develop the disease compared to heterozygotes. The dosage of DQA1*0501 appears less tightly linked to the development of celiac disease. Thus, HLA-DQ2.5 homozygotes and HLA-DQ2.5*cis*/2.2 heterozygotes show a higher risk for CD, compared to HLA-DQ2.5*cis*/non-DQ2.2 heterozygotes (DQ2.5/X) and HLA-DQ2.5(*trans*) heterozygotes (15–18). Finally, the HLA-DQ8 genes associated with celiac disease, DQA1*0301 and DQB1*0302, confer a low risk in comparison to DQ2 genes (16). However, risk grade for CD in DQ2.5/DQ8 carriers is unclear, as some studies have classified these patients as having the highest genetic predisposition (16) with others as being intermediate risk only (19). Overall, about 35% of the genetic risk of developing celiac disease is associated with the presence of the HLA-DQ2/DQ8 haplotype (20, 21).

To further complicate the subject, HLA-DQ haplotypes are linked to HLA-DR genes, forming HLA-DR-DQ haplotypes, mainly DR3-DQ2.5, DR5/7-DQ2.2, and DR4-DQ8 in the case of celiac disease. HLA-DR haplotypes are known to confer risk for a variety of immune-mediated disorders (**Figure 1**): HLA DR3 is associated to Graves disease, myasthenia gravis, systemic lupus erythematosus (SLE), Hashimoto's thyroiditis, and type 1 diabetes, the last two also associated to DR4. HLA-DR4 is also associated to rheumatoid arthritis (22). Risk can be independently conferred by DQ and DR alleles: for instance, type 1 diabetes risk is independently associated with both DQ2.5/8 alleles (HLA-DQB1*0201/HLA-DQB1*0302) and DR3/4 alleles (HLA-DRB1*0301/HLA-DRB1*0405, HLA-DRB1*0401) (23).



Increased risk for co-occurrence of type 1 diabetes and celiac disease has been reported for DR3-DQ2.5 homozygotes as well as for DR3-DQ2.5/DR4-DQ8 vs. celiac disease alone; on the contrary, DR3-DQ2.5/DR7-DQ2.2 did not increase the risk for co-occurrence (24).

Different studies have shown that the risk of developing celiac disease among the patient's relatives is higher than that of the general population. The prevalence of the disease in first-degree relatives (FDRs) is 7.5% and varies considerably depending on the degree of kinship (e.g., sisters and daughters have higher risk than fathers and sons) (25). These potential patients should be monitored and tested periodically for anti-TG2 and anti-endomysial antibodies. However, as mentioned above, only those carrying the HLA-DQ2/DQ8 can develop the disease; therefore, FDRs are preliminarily stratified for the presence of these haplotypes (**Figure 1**). This genetic typing is generally based on the amplification of genomic DNA, extracted from blood samples or mouth swabs, by polymerase chain reaction (PCR) followed by hybridization of the PCR product with sequence-specific oligonucleotide probes (26). An alternative approach is the direct detection of haplotypes by real-time PCR (27).

Due to the high complexity of the HLA locus, only recently, through a fine mapping analysis, it was discovered that other portions of HLA besides DQ2/DQ8 are important in celiac disease (28, 29): specifically, the small nucleotide polymorphism (SNPs) rs2301226 located within the HLA class II locus and connected by expression quantitative trait loci (eQTL) to the expression HLA-DPB1 and B3GALT4; the HLA class I alleles HLA-B0801 and HLA-B3906 as well as the SNPs that influence the expression of HLA-F have all been associated to celiac disease (28). Excluding the HLA-DQ2/DQ8, genetic information within the HLA locus represents a further 18% of the risk (28).

GENETIC VARIANTS OF NON-HLA LOCI ASSOCIATED WITH CELIAC DISEASE

Although robust evidence on the role of HLA molecules and particular genotypes may explain their role in celiac disease pathogenesis, these are important yet not sufficient factors to explain the occurrence of the disease in ~3% of all individuals

harboring DQ2 or DQ8 HLA loci. Sequencing of the human genome opened the possibility of mapping genetic variants and analyzing their association with complex diseases. To date, over 40 additional genetic loci have been associated with celiac disease outside the HLA region (30). These variants are mainly SNPs found in non-coding regions, often within enhancers, suggesting an effect on gene expression rather than causing modifications of protein functions. The pathogenic mechanisms linking celiac disease with the few genetic variants found within encoding regions (e.g., MMEL1, SH2B3, IRAK1, and NCF2) are not fully understood, although these proteins play important roles in adaptive immune response, immune cell signaling, T cell maturation, and differentiation (29). Association of HLA and Non-HLA alleles may favor the occurrence of celiac disease as shown in a large population study of 2,308 cases and 4,585 controls, in which risk increased six-fold when 13 non-HLA alleles were present, compared to those with zero to five alleles (31). Non-HLA alleles may also ameliorate risk prediction. In a study involving 2,675 cases and 2,815 controls, adding 57 non-HLA variants to HLA testing showed a statistically significant improvement of the receiver operating characteristic (ROC) curve from 0.823 (for HLA only) to 0.854, which translates in a reclassification of risk in 11% of individuals (32). In a similar approach, Izzo et al. improved the prediction of the disease in a cohort of first-degree relatives, particularly in the low HLA risk groups (33). The association of such non-HLA regions with celiac disease may show some geographic variation, being statistically significant in Swedish children but insignificant in US and Finland (34). However, the possible use of non-HLA variants for the stratification of celiac patients has not yet been introduced in the clinical setting (**Figure 1**). A recent study selected 42 SNPs from a European case control study of about 12,000 patients and as many controls and showed that these SNPs can discriminate celiac disease more than HLA-DQ2/DQ8 in both adults (1,280 cases) and children (114 cases) cohorts (35). Moreover, since clinical presentation of celiac disease differs in siblings with similar HLA haplotypes, it has been suggested that non-HLA genes may play a significant role more so than HLA-concordance in siblings (36). It has also been reported that risk variants in the IL2-IL21 region may further contribute to celiac disease susceptibility (37). The same study

group proposed seven other regions selected from 1,164 non-HLA SNPs, which may modulate innate and adaptive immunity (20). Others have demonstrated the presence of non-HLA risk variants that act by influencing regulatory regions consistent with those previously reported in other immune-mediated diseases (29).

These observations may be relevant also to the occurrence of autoimmune disorders in celiac disease. In fact, celiac disease may present with a number of associated autoimmune conditions, and some of these, namely, multiple sclerosis (MS), autoimmune thyroiditis, psoriasis, vitiligo, alopecia, inflammatory bowel disease (IBD), autoimmune hepatitis, primary biliary cholangitis, type I diabetes, rheumatoid arthritis (RA), ankylosing spondylitis, and systemic lupus erythematosus (SLE) show some genetic overlap with celiac disease; in particular, IBD, MS, and RA show a substantial number of associated loci (110, 97, and 101, respectively) (38).

Several studies have observed a shared genetic pathway for non-HLA genes that may predispose to autoimmune diseases in children and adults (39–43). Although relevant to its occurrence, non-HLA loci do not appear to favor progression of the disease toward severe complications such as refractory celiac disease type II (RCD II), a condition that frequently evolves to intestinal lymphoma (44). In a 2018 study, none out of 39 known non-HLA genetic loci associated to celiac disease were found to be significantly associated with susceptibility to RCD II.

Finally, intestinal biopsies and immune cells eQTL analysis, aimed at understanding the possible functional role of the SNPs, led to the identification of over 100 candidate genes. Integrated analysis of data related to genetic regulome elements suggested that SNPs associated with celiac disease mainly impact on the regulatory layer of the cells. For example, over 75% of the SNPs belong to DNase I hypersensitive regions and might influence the binding of numerous transcription factors (30).

GENE EXPRESSION ALTERED IN THE SMALL INTESTINE OF PATIENTS AFFECTED BY CELIAC DISEASE

Genome-wide gene expression studies were also used to further our understanding of celiac disease. These studies, using intestinal biopsies, revealed a number of differentially expressed genes (DEG) including APOC3, CYP3A4, OCLN, MAD2L1, MKI67, CXCL11, and IL17A among the most representative (45). A more recent study identified over 1,000 DEGs; interestingly, of the 79 genes potentially involved in celiac disease by the genetic mapping of non-HLA loci (30), 65 were among the DEGs of this study (46). Pathway analysis suggested that these DEGs are involved in carbohydrate, lipid, and amino acid metabolism, microorganisms infection, native and adaptive immune response, intestinal permeability, and autoimmune conditions. Twenty-seven DEGs following validation by qPCR were investigated by principal component analysis (PCA). The PC1 summarizing 79% of gene expression variance strongly correlated with the severity of mucosal damage. It is interesting to note that five genes (CXCL10, GBP5, IFI27, IFNG, and UBD) were differentially expressed in patients with potential celiac disease at the time

of the investigation and who developed overt celiac condition afterwards (46).

CLINICAL PRESENTATION OF NON-HLA GLUTEN-RELATED DISORDERS

Non-HLA gluten-related disorders share similar clinical characteristics with irritable bowel syndrome (IBS). Indeed, some studies have included this condition in the IBS spectrum because of their overlapping symptoms. In this scenario, fermentable oligo-, di-, and monosaccharides and polyols (FODMAPs) could have a critical role in driving the pathogenesis and triggering symptoms (47). Conversely, other authors depicted a key ability of gluten alone in promoting the clinical aspects in specific clusters of patients (47–50).

Overall, patients with non-HLA gluten-related disorders usually report IBS-like gastrointestinal symptoms. Clinical manifestations often occur a short period after gluten assumption and can last from hours to few days (50). Abdominal pain and bloating, altered bowel habits compatible with IBS-D (diarrhea), IBS-C (constipation) or IBS-M (mixed), nausea, and reflux are commonly described (51, 52). Moreover, some subjects can also report extra-intestinal manifestations such as headache, fatigue, foggy mind, fibro-myalgia, anxiety, depression, altered sleep pattern, weight gain, joint pain, skin rash, and dermatitis (48, 51–54). These manifestations can also be described in celiac disease and are not connected to FODMAPs mechanisms of action. Thus, they could mainly represent a potential specific effect of gluten (50). Similarly, skin manifestations are often described in an important percentage of subjects and improve when on a gluten-free diet (55, 56).

Finally, a potential relationship between non-HLA gluten-related disorders and neuropsychiatric diseases has been suggested. In particular, autism spectrum and schizophrenia have been mainly involved (50). This connection may be due to the increased intestinal permeability (i.e., leaky-gut syndrome) often described in non-HLA gluten-related disorders. In this context, gluten and gluten-related molecules can pass into the bloodstream along the gut–brain axis and have a strong impact on the endogenous opioid and neurotransmission brain system (57, 58). Gluten molecules could have the ability to interact with opioid receptors, cause immune cells activation, trigger neuro-inflammation, and overall modulate patients' behavior. Furthermore, gluten may affect serotonin production and thus play a potential role in depression spectrum disorders. Although the exact mechanisms remain unclear, a possible rationale could be linked to gut microbiota activity, a major direct and indirect source of serotonin, which could be negatively influenced by gluten-containing foods (50).

GENE AND MICRORNA EXPRESSION IS DYSREGULATED IN THE SMALL INTESTINE OF PATIENTS AFFECTED BY NCWS

In contrast with celiac disease, very little is known about the genetics of NCWS. First of all, patients affected by NCWS do

not carry any characteristic HLA (59). HLA DQ2 and DQ8 are present in about 40–50% of NCWS patients, a percentage slightly above the prevalence of these haplotypes in the general population which suggests that DQ2/DQ8 has minor relevance, if any, in the development of NCWS (11). Furthermore, genotyping of NCWS is not yet available. The examination of a few genes involved in the immune response demonstrated the downregulation of the T-regulator marker FOXP3 and the upregulation of Toll like receptor 2 suggesting a possible role of innate immunity in NCWS. Claudin was also upregulated, in agreement with the increase of intestinal barrier function (12).

In a recent study by our laboratory we identified seven miRNA (hsa-miR-19b-3p, hsa-miR-19a-3p, hsa-miR-186-5p, hsa-miR-17-5p, hsa-miR-145-5p, hsa-miR-30e-5p, and hsa-miR-143-3p) whose expression was higher in the intestinal mucosa of NCWS patients than in controls with dyspeptic symptoms or celiac disease (60). Remarkably, six of these miRNA were also dysregulated in peripheral blood leukocytes of NCWS. In addition, from an analysis of gene expression on a genomic scale in intestinal biopsies of NCWS patients, we have identified 300 DEGs (61). The vast majority of these transcripts were long non-coding RNA (lncRNA), while only 36% were protein-coding transcripts. Although still poorly investigated, lncRNA are considered regulatory RNAs involved in various pathophysiological functions including the immune response and inflammation (62–64). Similarly to celiac disease, non-coding regions of the genome might be important also for the development of NCWS. These lncRNAs can exert their effects by controlling the organization of chromatin, the transcription, the splicing, the stability of mRNAs, or the availability of miRNAs (65). In fact, further investigation is needed to understand their

role in NCWS. Functional analysis of the coding transcripts revealed the presence of genes of inflammation, innate immunity, or previously associated with autoimmune diseases. Ingenuity pathway analysis suggested that hedgehog signaling pathway and regulators of circadian rhythm may also have a role in NCWS (61). Finally, these biomarkers (miRNAs and lncRNAs) after appropriate validation in a clinical setting could be potentially useful for the diagnosis of NCWS (Figure 1).

CONCLUSIONS

Identification and characterization of non-HLA celiac disease genes is an important tool to the clarification of the pathophysiology of celiac disease as well as for risk prediction and diagnosis. It may also help identify patients with a borderline disease or with discordant clinical, laboratory, and pathology features. The risk of developing extraintestinal manifestations of the disease or the progression toward more severe stages of celiac disease might also be predicted. Finally, disentangling the biologic role of non-HLA genes may clarify the disease process and enlighten the road to potential new treatments.

AUTHOR CONTRIBUTIONS

MS and MN conceived the manuscript and contributed to its organization, creation of a draft, revision, and editing. LL and KE contributed to the writing and editing of the manuscript, as well as the literature revision. All authors contributed to the article and approved the submitted version.

REFERENCES

- Hujoel IA, Reilly NR, Rubio-Tapia A. Celiac disease: clinical features and diagnosis. *Gastroenterol Clin North Am.* (2019) 48:19–37. doi: 10.1016/j.gtc.2018.09.001
- Green PH, Cellier C. Celiac disease. *N Engl J Med.* (2007) 357:1731–43. doi: 10.1056/NEJMra071600
- Fasano A, Not T, Wang WL, Uzzau S, Berti I, Tommasini A, et al. Zonulin, a newly discovered modulator of intestinal permeability, and its expression in coeliac disease. *Lancet.* (2000) 355:1518–9. doi: 10.1016/S0140-6736(00)02169-3
- Schuppan D, Junker Y, Barisani D. Celiac disease: from pathogenesis to novel therapies. *Gastroenterology.* (2009) 137:1912–33. doi: 10.1053/j.gastro.2009.09.008
- Fasano A, Catassi C. Celiac disease. *New Engl J Med.* (2012) 367:2419–26. doi: 10.1056/Nejmcp1113994
- Meresse B, Ripoche J, Heyman M, Cerf-Bensussan N. Celiac disease: from oral tolerance to intestinal inflammation, autoimmunity and lymphomagenesis. *Mucosal Immunol.* (2009) 2:8–23. doi: 10.1038/mi.2008.75
- Meresse B, Chen ZG, Ciszewski C, Tretiakova M, Bhagat G, Krausz TN, et al. Coordinated induction by IL15 of a TCR-independent NKG2D signaling pathway converts CTL into lymphokine-activated killer cells in celiac disease. *Immunity.* (2004) 21:357–66. doi: 10.1016/j.immuni.2004.06.020
- Vader LW, de Ru A, van der Wal Y, Kooy YMC, Benckhuijsen W, Mearin ML, et al. Specificity of tissue transglutaminase explains cereal toxicity in celiac disease. *J Experimental Med.* (2002) 195:643–9. doi: 10.1084/Jem.20012028
- Petersen J, Montserrat V, Mujico JR, Loh KL, Beringer DX, van Lummel M, et al. T-cell receptor recognition of HLA-DQ2-gliadin complexes associated with celiac disease. *Nat Struct Mol Biol.* (2014) 21:480–8. doi: 10.1038/nsmb.2817
- Petersen J, van Bergen J, Loh KL, Kooy-Winkelaar Y, Beringer DX, Thompson A, et al. Determinants of gliadin-specific T cell selection in celiac disease. *J Immunol.* (2015) 194:6112–22. doi: 10.4049/jimmunol.1500161
- Sapone A, Bai JC, Ciacci C, Dolinsek J, Green PH, Hadjivassiliou M, et al. Spectrum of gluten-related disorders: consensus on new nomenclature and classification. *BMC Med.* (2012) 10:13. doi: 10.1186/1741-7015-10-13
- Sapone A, Lammers KM, Casolaro V, Cammarota M, Giuliano MT, De Rosa M, et al. Divergence of gut permeability and mucosal immune gene expression in two gluten-associated conditions: celiac disease and gluten sensitivity. *BMC Med.* (2011) 9:23. doi: 10.1186/1741-7015-9-23
- Lionetti E, Catassi C. Co-localization of gluten consumption and HLA-DQ2 and -DQ8 genotypes, a clue to the history of celiac disease. *Dig Liver Dis.* (2014) 46:1057–63. doi: 10.1016/j.dld.2014.08.002
- Liu E, Rewers M, Eisenbarth GS. Genetic testing: who should do the testing and what is the role of genetic testing in the setting of celiac disease? *Gastroenterology.* (2005) 128:S33–7. doi: 10.1053/j.gastro.2005.02.013
- Vader W, Stepniak D, Kooy Y, Mearin L, Thompson A, van Rood JJ, et al. The HLA-DQ2 gene dose effect in celiac disease is directly related to the magnitude and breadth of gluten-specific T cell responses. *Proc Natl Acad Sci USA.* (2003) 100:12390–5. doi: 10.1073/pnas.2135229100
- Megiorni F, Mora B, Bonamico M, Barbato M, Nenna R, Maiella G, et al. HLA-DQ and risk gradient for celiac disease. *Hum Immunol.* (2009) 70:55–9. doi: 10.1016/j.humimm.2008.10.018
- Medrano LM, Dema B, Lopez-Larios A, Maluenda C, Bodas A, Lopez-Palacios N, et al. HLA and celiac disease susceptibility: new genetic factors bring open questions about the HLA influence and gene-dosage effects. *PLoS ONE.* (2012) 7:e48403. doi: 10.1371/journal.pone.0048403

18. Almeida LM, Gandolfi L, Pratesi R, Uenishi RH, de Almeida FC, Selleski N, et al. Presence of DQ2.2 associated with DQ2.5 increases the risk for celiac disease. *Autoimmune Dis.* (2016) 2016:5409653. doi: 10.1155/2016/5409653
19. Piccini B, Vascotto M, Serracca L, Luddi A, Margollicci MA, Balestri P, et al. HLA-DQ typing in the diagnostic algorithm of celiac disease. *Rev Esp Enferm Dig.* (2012) 104:248–54. doi: 10.4321/s1130-01082012000500005
20. Hunt KA, Zhernakova A, Turner G, Heap GA, Franke L, Bruinenberg M, et al. Newly identified genetic risk variants for celiac disease related to the immune response. *Nat Genet.* (2008) 40:395–402. doi: 10.1038/ng.102
21. Mooney PD, Hadjivassiliou M, Sanders DS. Coeliac disease. *BMJ.* (2014) 348:g1561. doi: 10.1136/bmj.g1561
22. Gough SC, Simmonds MJ. The HLA region and autoimmune disease: associations and mechanisms of action. *Curr Genomics.* (2007) 8:453–65. doi: 10.2174/138920207783591690
23. Jones EY, Fugger L, Strominger JL, Siebold C. MHC class II proteins and disease: a structural perspective. *Nat Rev Immunol.* (2006) 6:271–82. doi: 10.1038/nri1805
24. Smigoc Schweiger D, Mendez A, Kunilo Jamnik S, Bratanic N, Bratina N, Battelino T, et al. High-risk genotypes HLA-DR3-DQ2/DR3-DQ2 and DR3-DQ2/DR4-DQ8 in co-occurrence of type 1 diabetes and celiac disease. *Autoimmunity.* (2016) 49:240–7. doi: 10.3109/08916934.2016.1164144
25. Singh P, Arora S, Lal S, Strand TA, Makharia GK. Risk of celiac disease in the first- and second-degree relatives of patients with celiac disease: a systematic review and meta-analysis. *Am J Gastroenterol.* (2015) 110:1539–48. doi: 10.1038/ajg.2015.296
26. Fasano ME, D'Amato E, D'Alfonso S. HLA genotyping: methods for the identification of the HLA-DQ2,-DQ8 Heterodimers Implicated in Celiac Disease (CD) susceptibility. *Methods Mol Biol.* (2015) 1326:79–92. doi: 10.1007/978-1-4939-2839-2_9
27. D'Ippolito S, Gasbarrini A, Castellani R, Rocchetti S, Sisti LG, Scambia G, et al. Human leukocyte antigen (HLA) DQ2/DQ8 prevalence in recurrent pregnancy loss women. *Autoimmun Rev.* (2016) 15:638–43. doi: 10.1016/j.autrev.2016.02.009
28. Gutierrez-Achury J, Zhernakova A, Pulit SL, Trynka G, Hunt KA, Romanos J, et al. Fine mapping in the MHC region accounts for 18% additional genetic risk for celiac disease. *Nat Genet.* (2015) 47:577–8. doi: 10.1038/ng.3268
29. Trynka G, Hunt KA, Bockett NA, Romanos J, Mistry V, Szperl A, et al. Dense genotyping identifies and localizes multiple common and rare variant association signals in celiac disease. *Nature Genetics.* (2011) 43:1193–U45. doi: 10.1038/ng.998
30. Withoff S, Li Y, Jonkers I, Wijmenga C. Understanding celiac disease by genomics. *Trends Genet.* (2016) 32:295–308. doi: 10.1016/j.tig.2016.02.003
31. Romanos J, van Diemen CC, Nolte IM, Trynka G, Zhernakova A, Fu J, et al. Analysis of HLA and non-HLA alleles can identify individuals at high risk for celiac disease. *Gastroenterology.* (2009) 137:834–40, 840 e1-3. doi: 10.1053/j.gastro.2009.05.040
32. Romanos J, Rosen A, Kumar V, Trynka G, Franke L, Szperl A, et al. Improving coeliac disease risk prediction by testing non-HLA variants additional to HLA variants. *Gut.* (2014) 63:415–22. doi: 10.1136/gutjnl-2012-304110
33. Izzo V, Pinelli M, Tinto N, Esposito MV, Cola A, Sperandeo MP, et al. Improving the estimation of celiac disease sibling risk by non-HLA genes. *PLoS ONE.* (2011) 6:e26920. doi: 10.1371/journal.pone.0026920
34. Sharma A, Liu X, Hadley D, Hagopian W, Liu E, Chen WM, et al. Identification of Non-HLA genes associated with celiac disease and country-specific differences in a large, international pediatric cohort. *PLoS ONE.* (2016) 11:e0152476. doi: 10.1371/journal.pone.0152476
35. Sharp SA, Jones SE, Kimmitt RA, Weedon MN, Halpin AM, Wood AR, et al. A single nucleotide polymorphism genetic risk score to aid diagnosis of coeliac disease: a pilot study in clinical care. *Aliment Pharm Ther.* (2020) 52:1165–73. doi: 10.1111/apt.15826
36. Kauma S, Kaukinen K, Huhtala H, Kivela L, Pekki H, Salmi T, et al. The phenotype of celiac disease has low concordance between siblings, despite a similar distribution of HLA haplotypes. *Nutrients.* (2019) 11:479. doi: 10.3390/nu11020479
37. van Heel DA, Franke L, Hunt KA, Gwilliam R, Zhernakova A, Inouye M, et al. A genome-wide association study for celiac disease identifies risk variants in the region harboring IL2 and IL21. *Nat Genet.* (2007) 39:827–9. doi: 10.1038/ng2058
38. Ricano-Ponce I, Wijmenga C, Gutierrez-Achury J. Genetics of celiac disease. *Best Pract Res Clin Gastroenterol.* (2015) 29:399–412. doi: 10.1016/j.bpg.2015.04.004
39. Ostensson M, Montén C, Bacelis J, Gudjonsdottir AH, Adamovic S, Ek J, et al. A possible mechanism behind autoimmune disorders discovered by genome-wide linkage and association analysis in celiac disease. *PLoS ONE.* (2013) 8:e70174. doi: 10.1371/journal.pone.0070174
40. Gutierrez-Achury J, Coutinho de Almeida R, Wijmenga C. Shared genetics in coeliac disease and other immune-mediated diseases. *J Intern Med.* (2011) 269:591–603. doi: 10.1111/j.1365-2796.2011.02375.x
41. Gutierrez-Achury J, Zorro MM, Ricano-Ponce I, Zhernakova DV, Coeliac Disease Immunochip Consortium RC, Diogo D, et al. Functional implications of disease-specific variants in loci jointly associated with coeliac disease and rheumatoid arthritis. *Hum Mol Genet.* (2016) 25:180–90. doi: 10.1093/hmg/ddv455
42. Parkkola A, Laine AP, Karhunen M, Harkonen T, Ryhanen SJ, Ilonen J, et al. HLA and non-HLA genes and familial predisposition to autoimmune diseases in families with a child affected by type 1 diabetes. *PLoS ONE.* (2017) 12:e0188402. doi: 10.1371/journal.pone.0188402
43. Zhernakova A, Stahl EA, Trynka G, Raychaudhuri S, Festen EA, Franke L, et al. Meta-analysis of genome-wide association studies in celiac disease and rheumatoid arthritis identifies fourteen non-HLA shared loci. *PLoS Genet.* (2011) 7:e1002004. doi: 10.1371/journal.pgen.1002004
44. Hrdlickova B, Mulder CJ, Malamut G, Meresse B, Platteel M, Kamatani Y, et al. A locus at 7p14.3 predisposes to refractory celiac disease progression from celiac disease. *Eur J Gastroenterol Hepatol.* (2018) 30:828–37. doi: 10.1097/MEG.0000000000001168
45. Bragde H, Jansson U, Jarlsfelt I, Soderman J. Gene expression profiling of duodenal biopsies discriminates celiac disease mucosa from normal mucosa. *Pediatr Res.* (2011) 69:530–7. doi: 10.1203/PDR.0b013e318217ecec
46. Bragde H, Jansson U, Fredrikson M, Grodzinsky E, Soderman J. Celiac disease biomarkers identified by transcriptome analysis of small intestinal biopsies. *Cell Mol Life Sci.* (2018) 75:4385–401. doi: 10.1007/s00018-018-2898-5
47. Vazquez-Roque M, Oxentenko AS. Nonceliac gluten sensitivity. *Mayo Clinic Proc.* (2015) 90:1272–7. doi: 10.1016/j.mayocp.2015.07.009
48. De Giorgio R, Volta U, Gibson PR. Sensitivity to wheat, gluten and FODMAPs in IBS: facts or fiction? *Gut.* (2016) 65:169–78. doi: 10.1136/gutjnl-2015-309757
49. Capannolo A, Viscido A, Barkad MA, Valerii G, Ciccone F, Melideo D, et al. Non-Celiac Gluten Sensitivity among Patients Perceiving Gluten-Related Symptoms. *Digestion.* (2015) 92:8–13. doi: 10.1159/000430090
50. Dale HF, Biesiekierski JR, Lied GA. Non-coeliac gluten sensitivity and the spectrum of gluten-related disorders: an updated overview. *Nutrition Res Rev.* (2019) 32:28–37. doi: 10.1017/S095442241800015X
51. Catassi C, Elli L, Bonaz B, Bouma G, Carroccio A, Castillejo G, et al. Diagnosis of non-celiac gluten sensitivity (NCGS): the salerno experts' criteria. *Nutrients.* (2015) 7:4966–77. doi: 10.3390/nu7064966
52. Catassi C, Bai JC, Bonaz B, Bouma G, Calabro A, Carroccio A, et al. Non-Celiac Gluten sensitivity: the new frontier of gluten related disorders. *Nutrients.* (2013) 5:3839–53. doi: 10.3390/nu5103839
53. Khalid AN, McMains KC. Gluten sensitivity: fact or fashion statement? *Curr Opin Otolaryngol Head Neck Surg.* (2016) 24:238–40. doi: 10.1097/MOO.0000000000000263
54. Peters SL, Biesiekierski JR, Yelland GW, Muir JG, Gibson PR. Randomised clinical trial: gluten may cause depression in subjects with non-coeliac gluten sensitivity - an exploratory clinical study. *Alimentary Pharmacol Therapeutics.* (2014) 39:1104–12. doi: 10.1111/apt.12730
55. Bonciolini V, Bianchi B, Del Bianco E, Verdelli A, Caproni M. Cutaneous manifestations of non-celiac gluten sensitivity: clinical histological and immunopathological features. *Nutrients.* (2015) 7:7798–805. doi: 10.3390/nu7095368
56. Picarelli A, Borghini R, Di Tola M, Marino M, Urciuoli C, Isonne C, et al. Intestinal, systemic, and oral gluten-related alterations in patients with nonceliac gluten sensitivity. *J Clin Gastroenterol.* (2016) 50:849–58. doi: 10.1097/MCG.0000000000000515
57. Czaja-Bulsa G. Non coeliac gluten sensitivity - A new disease with gluten intolerance. *Clin Nutrition.* (2015) 34:189–94. doi: 10.1016/j.clnu.2014.08.012

58. de Magistris L, Familiari V, Pascotto A, Sapone A, Froli A, Iardino P, et al. Alterations of the intestinal barrier in patients with autism spectrum disorders and in their first-degree relatives. *J Pediatric Gastroenterol Nutrition*. (2010) 51:418–24. doi: 10.1097/MPG.0b013e3181dcc4a5
59. Volta U, Bardella MT, Calabro A, Troncone R, Corazza GR, Study Group for Non-Celiac Gluten S. An Italian prospective multicenter survey on patients suspected of having non-celiac gluten sensitivity. *BMC Med*. (2014) 12:85. doi: 10.1186/1741-7015-12-85
60. Clemente E, Efthymakis K, Carletti E, Capone V, Sperduti S, Bologna G, et al. An explorative study identifies miRNA signatures for the diagnosis of non-celiac wheat sensitivity. *PLoS ONE*. (2019) 14:e0226478. doi: 10.1371/journal.pone.0226478
61. Efthymakis K, Clemente E, Marchioni M, Di Nicola M, Neri M, Sallese M. An exploratory gene expression study of the intestinal mucosa of patients with non-celiac wheat sensitivity. *Int J Mol Sci*. (2020) 21:1969. doi: 10.3390/ijms21061969
62. Carpenter S, Aiello D, Atianand MK, Ricci EP, Gandhi P, Hall LL, et al. A long noncoding RNA mediates both activation and repression of immune response genes. *Science*. (2013) 341:789–92. doi: 10.1126/science.1240925
63. NE II, Heward JA, Roux B, Tsitsiou E, Fenwick S, Lenzi L, et al. Long non-coding RNAs and enhancer RNAs regulate the lipopolysaccharide-induced inflammatory response in human monocytes. *Nat Commun*. (2014) 5:3979. doi: 10.1038/ncomms4979
64. Ma S, Ming Z, Gong AY, Wang Y, Chen X, Hu G, et al. A long noncoding RNA, lincRNA-Tnfaip3, acts as a coregulator of NF-kappaB to modulate inflammatory gene transcription in mouse macrophages. *FASEB J*. (2017) 31:1215–25. doi: 10.1096/fj.201601056R
65. Yao RW, Wang Y, Chen LL. Cellular functions of long noncoding RNAs. *Nat Cell Biol*. (2019) 21:542–51. doi: 10.1038/s41556-019-0311-8

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Sallese, Lopetuso, Efthymakis and Neri. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.